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(71) Applicant (for all designated States except US): COUNCIL OF SCIENTIFIC AND INDUSTRIAL RESEARCH [IN/IN]; Rafi Marg, New Delhi 110 001 (IN).

(72) Inventors; and

(75) Inventors/Applicants (for US only): VERMA, Sunil, Kumar [IN/IN]; Centre for Cellular and Molecular Biology, Hyderabad 500 007 (IN). SINGH, Lalji [IN/IN]; Centre for Cellular and Molecular Biology, Hyderabad 500 007 (IN).

(74) Agent: GABRIEL, Devadoss, Calab; Kumaran & Sagar, 84-C, C6 Lane, (Off Central Avenue), Sainik Farms, New Delhi 110 062 (IN).

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(54) Title: UNIVERSAL PRIMERS FOR WILDLIFE IDENTIFICATION

(57) Abstract: The invention provides novel universal primers that can amplify the fragment of cytochrome b gene of any animal species in polymerase chain reaction (PCR) and reveal the identity of the biological material of any unknown animal origin and a method for identification of the specific animal from a given biological sample.

UNIVERSAL PRIMERS FOR WILDLIFE IDENTIFICATION TECHNICAL FIELD

The invention relates to the identification of novel universal primers that can amplify the fragment of cytochrome b gene of any animal species in polymerase chain reaction (PCR) and reveal the identity of the biological material of any unknown animal origin at species and sub-species sources. The invention also provides a method for the identification of fragments on mitochondrial cytochrome b gene in biological material of unknown origin.

BACKGROUND ART

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A large number of studies in evolutionary biology utilize phylogenetic information obtained from mitochondrial cytochrome b gene. It has been identified a potent molecule to distinguish the phylogenetic depth of different lineages to family, genus and species in molecular taxonomy1-66. A vast database of the sequences of cytochrome b gene of different animal species has accumulated in public databases such as GenBank, NCBI (http://www.ncbi.nlm.nih.gov) etc. We have utilized this capacity of cytochrome b gene in establishing the identity of the origin of animal parts and product to its family, genus and species sources. The technique developed is based on a pair of universal primer that can amplify a small fragment of cytochrome b gene from a vast range of animal species. Establishing identity of confiscated animal parts and products is a great challenge to law enforcement agencies because none of the methods available till date is too efficient to reveal the identity of animal remains beyond a reasonable doubt. Morphological markers, described for certain species allow the identification of complete specimen of animals⁶⁷. However, a complete specimen is confiscated very rarely by the investigation agencies; therefore, these marker are not practical in wildlife forensics. The biochemical traits such as the bile characteristics⁶⁸ blood heam analysis^{69,70} etc. have also been employed in wildlife forensic for identification of individual species. The difficulty of these markers are that these markers are limited in number and are rarely found in their natural forms in which these were originally described as the characteristic of a particular species. The molecular approaches such as micro-satellite based identification⁷¹, Restriction

The molecular approaches such as micro-satellite based identification, Restriction fragment length polymorphism analysis of mitochondrial genes or PCR based species specific STS markers require the prior information of the species to establish the identity^{72,73}. These methods also need a significant amount of DNA material to be analysed. We may not have the prior information about the species origin of confiscated animal parts and product in forensics, therefore, these methods are not really useful and

practical in wildlife identification. The technique invented by us is universal, therefore does not require any background information to establish the identity of any unknown confiscated remains at family, genus and species sources. Being a PCR based procedure it can be applied with trace amount of any biological material. Because the amplicon length is small (472 bp); therefore, it can work perfectly with the mutilated remains, which are commonly seized by the crime investigation agencies. It does not require the large amount of genetic material i.e. DNA to be analyzed to establish the identity, hence, can detect a minute amount of adulteration in food products. The procedure described is simple and very fast. Due to the said advantages, the procedure invented by us is most suited for forensic wildlife identification.

OBJECTS OF THE INVENTION

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The main object of the invention is to identify a fragment on mitochondrial cytochrome b gene capable of significantly discriminating among various evolutionary lineages of different animal species.

Another object is to identify a fragment on mitochondrial cytochrome b gene which is flanked by the highly conserved sequences at a vast range of animal species.

Yet another object is to detect a fragment on mitochondrial cytochrome b gene which is polymorphic inter-specifically, but monomorphic at intra species sources.

Still another object is to develop the universal primers to amplify the fragment on mitochondrial cytochrome b gene using polymerase chain reaction.

Another object is to develop a PCR protocol that works universally with DNA template of any unknown origin (i.e. all the animal species).

Yet another object is to provide a universal method for identification of species of analyzed material (i.e. the DNA isolated from confiscated animal remain of unknown origin) using the public databases such as GenBank, NCBI etc.

Still another object is to provide a universal method of animal identification to establish the crime with the criminal beyond a reasonably doubt.

Another object is provide a universal method to establish the identity of biological materials such as skin, horns etc confiscated from animal poachers, if it is that of an endangered species.

Yet another object is to provide a universal method for establishment of the identity of confiscated animal parts and products of endangered animal species for the purpose of production of molecular evidence of animal hunting and related crime in the court of law, so that the human violation to the wildlife resources could be controlled.

Still another object is to provide a universal technique to have an idea of the geographical location of the commitment of wildlife crime based on the haplotype of poached animal—identified by the universal primer invented.

Another object is to provide a universal technique of animal identification to detect the adulteration of animal meat/products in vegetarian food product for the purpose of food fortification, by the food fortification agencies.

Yet another object is to provide a universal technique for detection of the origin of blood or blood stains etc collected from the scene of crime related to offences such as murder, rape etc, in order to establish the origin of blood found at scene of crime when it sounds as if criminals have wontedly spread the blood of an animal at the scene of crime, to confuse the crime investigation agencies and forensic scientists with human blood.

Another object is to invent and authenticate a universal technique that can be converted to a (a) 'MOLECUALR KIT' and (b) 'DNA CHIPS' based application to meet the requirements of above objectives.

15 SUMMARY OF THE INVENTION

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Accordingly, the invention provides novel universal primers that can amplify the fragment of cytochrome b gene of any animal species in polymerase chain reaction (PCR) and reveal the identity of the biological material of any unknown animal origin

DETAILED DESCRIPTION OF THE INVENTION

Keeping in view the above objectives, the cytochrome b gene sequences (1140 bp) of 221 distantly related animal species (listed in Table 1) representing various families were obtained from public database NCBI (http://www.ncbi.nlm.nih.gov). These sequences were aligned using the software Clustal X(1.8)(NCBI, USA) and a fragment (of 472 bp, alignment shown in Table 2) of gene was identified which had all the features mentioned above under column 1, 2 and 3 of sub-heading 'Objectives of invention'. As for the identity of this fragment we would like to mention that it includes the nucleotides between 398 to 869 in Antilope cervicapra and Felis catus; however, 399 to 870 in Homo sapiens sapiens species. Except at few positions (marked as star (*) in Table 2, the nucleotide sequences of this fragment are highly variable amongst the animal species, giving rise to their unique molecular signature. These molecular signatures are characteristic of its species and form the basis of revealing the identity of the biological material of an unknown animal origin by the procedure invented by us. Considering Antilope cervicapra as a representative species, the sequence of this fragment is mentioned herewith:

Mitochondrial cytochrome b gene sequence (398-869 bp) of Antilope cervicapra:

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"taccatgaggacaaatatctttttgaggagcaacagtcatcaccaatctcctttcagcaatcccatacatcggtacaaacctagtaga atgaatctgaggagggttctcagtagataaagcaacccttacccgatttttcgccttcactttatcctcccatttatcattgcagccctt accatagtacacctactgtttctccacgaaacaggatccaacaaccccacaggaatctcatcagaacgcagacaaaattccattcaac ccctactacactatcaaagatatcctaggagctctactattaattttaaccctcatgcttctagtcctattctaaccggacctgcttggag acccagacaactatacaccagcaaacccacttaatacacccccacatatcaagccgaatgatacttcctatttgcatacgcaatcct ccgatcaattcctaacacaacaaactaggagg".

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A pair of universal primer was designed to amplify this fragment in polymerase chain reaction (PCR). These primers were named as 'mcb398' and 'mcb869' because of its property to amplify a region of mitochondrial cytochrome b gene between nucleotides 398 to 869 of Antilope cervicapra, a representative animal species for this invention. We took this animal species as representative species because the idea of developing such a novel primers came in the mind of inventors while they were working on the genome of this animal in Centre for Cellular and Molecular Biology, Hyderabad, India. These primers work universally because its 3' end are highly conserved amongst a vast range of animal species (shown in Table 2). As mentioned above, the DNA fragment (sequence of which is shown above) targeted by these primers is highly polymorphic inter-specifically; however, it is monomorphic among the individual of same species (Tables 6, 7a, 7b, 7c, 1d and 8, respectively). These unique features of the targeted region enable these primers to generate the molecular signatures of an individual species; thereby, enabling them to differentiate amongst the animals of different species (see in Figure 1c). The variation within the fragment amplified by these primers increase with increasing distances of evolutionary lineages of two animals (Table 8). These unique features of the fragment amplified by the universal primers 'mcb398' and 'mcb869' invented by the applicants fulfill the objectives of invention.

Thus, the primers invented by us can generate the molecular signature from any biological material of unknown animal origin, which actually is the characteristic of its family, genus and more precisely, the species. When these signatures are compared *in-silico* with the signatures already available in public databases (viz., GenBank, NCBI database etc) using 'BLAST software⁷³, it indicates identity of the family, genus or species of the analyzed material, which in turn is confirmed practically by comparing with the reference unimals of the revealed family, genus or species, by including them in the further analysis by the primers 'mcb398' and 'mcb869'. The complete procedure involved in the analyses (the word, 'analyses' should be understood with the stepwise procedure to establish the identity of the biological remain of any unknown animal origin for the aims mentioned in columns

1-13 under sub-heading 'Objectives of invention') is briefed under 'Examples 5 and 6, respectively, as well as illustrated in Figures 1a, 1b and 1c, respectively.

BRIEF DESCRIPTION OF DRAWING AND TABLES

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Figure 1a. Illustration of the step-wise procedure involved in *analyses*. The unknown biological material i.e. 'adil.flesh' refers to the confiscated skin mentioned in 'Example 6'. The arrow marks indicate the stepwise procedure involved. The brief description of Figure 1a is as follows:

The biological material i.e. the confiscated skin 'adil.flesh' was subjected to DN. isolation using the standard procedures ⁷⁴. The DNA obtained was amplified using the primers 'mcb398' and 'mcb869' in PCR, fractionated in 2% (w/v) agarose gel, visualized and photographed under UV light using Gel Documentation System (Syngene, USA). The lane 'M' shown in the photograph represents the molecular weight marker (Marker XIII, Boehringer mannheim). Lane 1 shows the PCR amplicon (472 bp) obtained from 'adil.flesh' using primers 'mcb398' and 'mcb869'. The PCR amplicon obtained were sequenced at both the strand using "ABI Prism 3700 DNA Analyzes, PE-Applied Biosystems). The chromatogram shows the sequences (about 80 bp long, i.e. between 150-230 bp of sequence (328 bp), revealed from the PCR product of 472 bp length) obtained from 'adil.flesh'.

Figure 1b. Illustrates the further steps involved in analyses. The sequence (328 bp) revealed from 'adil.flesh' was subjected to homology search in nr (i.e. non-redundant) database of Netional Centre for Biological Information (NCBI), USA. The sequences producing significant alignments are shown along with its bits score and E values. It indicates the extent of homology amongst the sequence enquired (i.e. the 328 bp sequence from adil.flesh) and the sequences registered in nr database of NCBI. BLAST analysis revealed the highest homology of the sequence revealed from 'adil.flesh' with the sequence of Panthera pardus (gene bank registration number 'AY005809'), indicating the identity of adil.flesh as that of a leopard (Panthera pardus) origin. Figure 1b further illustrates the multiple alignments of the sequences obtained from reference animals (listed in Table 5) along with the sequence obtained from 'adil.flesh'. The sequences of 'adil.flesh' is similar to the sequences of 'gz1L' further confirming the identity of the source of confiscated remain 'adil.flesh' as that of a Panthera pardus origin.

Figure 1c illustrates the NJ-tree (Neighbor Joining tree) constructed using CLUSTAL X (1.8) from the sequences revealed from 'adil.flesh' and reference animals listed in Table 5.

The animals belonging to similar species cluster together; however, the animals of different species group in different clusters. The confiscated material under investigation (i.e. 'adil.flesh') clusters with 'gzlL' (i.e. the known normal leopard 'Panthera pardus') indicating the identity of the species of 'adil.flesh' as that of a Panthera pardus source.

- Figure 2 shows the Agarose gel electrophorogram showing the PCR amplicons (472 bp) obtained from the reference animals of family felidae listed in Table 5, using universal primers 'mcb398 and 'mcb869'. Description of different lanes is as follows:
 - Lanes 1-21: The PCR profiles of the animals 1-21, respectively, listed in Table 5.
 - Lane 22: The PCR profiles of DNA isolated from confiscated skin of unknown animal origin 'i.e. adil.flesh'
 - Lane 23: Negative control (no DNA)

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- Lane M: Molecular weight marker (marker XIII, Boehringer mannheim)
- Figure 3. Shows PCR amplicons obtained from animals listed in Table 9. The primers used in PCR are 'AFF' and 'AFR'. The description of different lanes shown is as follows:
- Lane 1-4: The PCR profiles of animals 1-4, respectively, listed in Table 9, showing amplicons of 354 bp.
 - Lane M: Molecular weight marker (marker XIII, Boehringer mannheim)
 - Figure 4. Shows PCR amplicons obtained from animals listed in Table 12. This experiment demonstrates the universal nature of our primers among a vast range of animal species. Description of different lanes shown is as follows:
 - Lanes 1-23: The PCR profiles of the animals 1-23, respectively, listed in Table 12. The PCR product of 472 bp is amplified universally from all the animal species analyzed.
 - Lane 24: Negative control (no DNA)
 - Lane M: Molecular weight marker (marker XIII, Boehringer mannheim)
- Table 1. List of 221 animal species used for *In-silico* analysis to design the universal primers 'mcb398' and 'mcb869'. Table also demonstrate the 'P,S scores' of 'mcb398' and 'mcb869' for different templates. The descriptions of various symbols used in this table are as follows:
 - Symbol (#) refers to Number
- Symbol (*) refers to the animal species which is either protected species (listed in Wildlife (Protection) Act, 1972 (Central Act NO 53 of 1972), or an endangered/rare animal species Symbol (⁵P,S/F) refers to Probability of match and Stability of match of primer 'mcb398' with different templates (i.e. the cytochrome b gene from different species origin). A higher P, S score refers to the higher probabilities of significant amplification of specific

template by the primer. It is calculated by Amplify (1.2) software.

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Symbol ($^{\Psi}P,S/R$) refers to Probability of match and Stability of match of primer 'mcb869' with different templates. A higher P,S score refers to the higher probabilities of significant amplification of specific template by the primer. It is calculated by *Amplify* (1.2) software.

Table 2. Multiple sequence alignment of 472 bp fragment of mitochondrial cytochrome b gene (identified by inventors to fulfill the requirements of column 1, 2 and 3 mention under sub-heading 'Objectives of invention') of 221 animal species listed in Table 1. Alignments also show the binding sites for universal primers 'mcb398' and 'mcb869'. The symbol (*) refers to the nucleotide bases which are conserved amongst 221 animal species listed in Table 1). The alignments have been done using software CLUSTAL X (1.8). The nucleitide positions that are unmarked are variable amongst 221 animal species analyzed. These variable sites together constitute the molecular signature of an individual species, giving rise to molecular basis of species identification by our primers.

Table 3. Results of the blast analysis of the sequence revealed from 'adil.flesh' in 'mito' database of NCBI. It shows the most significant alignment of cytochrome b sequence (328 bp) revealed from confiscated skin piece 'adil.flesh' with felis catus cytochrome b gene sequence (genbank registration number NC_001700.1, bits score 365, E value, e-101) registered in NCBI database (bits score 365 and E value e-101). It gives an indication that the species of analyzed material belongs to family felidae. It also fulfills the requirements of column 6 mention above under sub-heading 'Objectives of invention'.

Table 4. Results of the blast analysis of the sequence revealed from 'adil.flesh' in 'nr' database of NCBI. It shows the most significant alignment of cytochrome b sequence (328 bp) revealed from confiscated skin piece 'adil.flesh' with Panthera pardus cytochrome b gene sequence (genbank registration number AY005809, bits score 603, E value, e-170) registered in NCBI database. It gives an indication that the species of analyzed material belongs to Panthera paurdus origin. It also fulfills the requirements of column 6 mention above under sub-heading 'Objectives of invention'.

Table 5. Reference animal belonging to family felidae selected for comparison with 'adil.flesh' to confirm the findings of BLAST analysis results of which are mentioned in Table 3 and 4, respectively. The animals listed in SN. 1-21 represent different species of family felidae. SN. 22 and 23 are primate species taken for out-group comparisons.

Table 6 Multiple sequence alignments of cytochrome b sequences (328 bp) revealed from 'adil.flesh' and reference animals listed in Table 5. The positions that have a common nucleotide in all the animal species under investigation are shown with a star (*) mark;

however, the positions that are variable in any of the animals under investigation are unmarked. The nucleotides at these positions constitute the molecular signature of an individual species, which are unique and highly specific for its species. These signatures are the molecular basis of identification of individual animal species using our primers 'mcb398' and 'mcb869'.

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Table 7 (Tables 7a, 7b, 7c and 7d). The comparison of the molecular signatures of different animal species investigated along with 'adil.flesh', the confiscated skin of unknown animal origin. This table demonstrates the variable positions (i.e. the positions which are not marked with star (*) symbol in Table 6), amongst the 328 bp fragment revealed from the animals listed in Table 5. The dot (.) mark represents the presence of the similar nucleotide as listed in lane 1 i.e. the sequence from "adil flesh' at that position. It demonstrates that the signatures of each species are unique and specific to its species. The molecular signatures of 'adil.flesh' are comparable (except for position 37 which has a transition from 'T' to 'C') to the molecular signature of 'gz1L' i.e. the known leopard 'Panthera pardus' source, indicating the identity of the source of confiscated skin 'adil.flesh' as that of a leopard 'Panthera pardus' source. The nucleotide variations (at the positions 153, 198, 223, 264, among the known leopards, (i.e. gz1L, gz2L, and gz3L, respectively)), give an idea about the geographical habitat of each animals. Various studies referring to molecular evolution of different animal species support this hypothesis⁷⁵; however, it could further be confirmed by taking the reference animals from different geographical areas and analyzing by our primers 'mcb 398' and 'mcb869'. If we could generate the database of different haplotypes (i.e. habitat specific molecular signatures) of the animal species, it would also enable our primers to reveal the geographical location of the commitment of wildlife crime.

Table 8. Percent similarity matrix calculated by pair-vise comparisons of nucleotide sequences aligned (illustrated in Table 6). The cytochrome b gene sequence of DNA isolated from confiscated material had maximum similarity (99.7% and 98.2%, with the lineages of animals 'gz2L' and 'gz3L', respectively) with the sequences obtained from known normal leopard source, indicating its identity as that of a leopard origin. The similarity matrix has been calculated using the software *PHYLIP* (3.5).

Table 9. Animals selected for validation of minimum P,S score for efficient amplification of cytochrome b gene of different origin by the primers 'mcb398' and 'mcb869'. P,S score of primers 'AFF' and 'AFR' for these animals are shown.

Table 10. BLAST analysis of primers 'mcb398' in nr database of NCBI. It demonstrates

that the 3' end of this primer is highly conserved among a vast range of animal species. It also shows the significant homology among the primer and templates (i.e. the cytochrome b gene fragment of different animal species), confirming the universal nature of our primer **Table 11**. BLAST analysis of primers 'mcb869' in *nr* database of NCBI. It demonstrates that the 3' end of this primer is highly conserved among a vast range of animal species. It also shows the significant homology among the primer and templates (i.e. the cytochrome b gene fragment of different animal species), confirming the universal nature of our primer.

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Table 12. Other animal belonging to distantly related animal species, investigated to confirm the universal nature of primers 'mcb398' and 'mcb869'. Gel photograph showing the PCR amplicons from these animals are shown in Figure 4.

The mitochondrial cytochrome b gene has very widely been used in molecular taxonomic studies. It has immense capabilities to reveal different evolutionary lineages of animals in family, genus and species specific manner. It has also been used to classify the population of a particular species according to its demographic distributions⁷⁵. The vast database of cytochrome b sequences of different animal species has accumulated in public databases such as Genbank and NCBI¹⁻⁶⁵. We have explored these unique characteristics of cytochrome b gene to establish the identity of confiscated remains of any unknown animal by inventing a pair of novel primers, 'mcb398' and 'mcb869', that can amplify a small fragment (472 bp) of cytochrome b gene of wide range of animal species in universal manner. These primers work universally because its 3' ends target within a highly conserved region.

The fragment of cytochrome b gene identified had all the features mentioned in columns 1, 2 and 3 listed under sub-heading 'Objective of invention'. We identified this fragment by aligning the cytochrome b gene sequences (1140 bp) of 221 different animal species listed in Table 1. These sequences are publicly available in NCBI DNA databases. These sequences were aligned using the software CLUSTAL X (1.8). As mentioned before, the 472 bp fragment of cytochrome b gene identified by us to have the features mentioned in columns 1, 2 and 3 listed under sub-heading 'Objective of invention' includes the nucleotides between 398 to 869 in Antilope cervicapra and Felis catus; however, 399 to 870 in Homo sapiens sapiens species. Except at few positions (marked as star (*) in Table 2, the nucleotide sequences of this fragment are highly variable amongst the animal species, revealing the identity of the biological material belonging to that of an unknown animal origin by the procedure invented by us. As for identity of this fragment we are

considering Antilope cervicapra as a representative species, and the sequence the above fragment of cytochrome b gene of Antilope cervicapra is mentioned herewith:

Mitochondrial cytochrome b gene sequence (398-869 bp) of Antilope cervicapra

"taccatgaggacaaatatetttttgaggagcaacagteateaceaateteettteageaateeeatacateggtacaaacetagtaga at gaatet gaggagggt tet cagtagataa ag caaccett accegatt ttte geet te caett ta teet ce catt ta teet caet tta teet caett ta teetccctactacactatcaaagatatcctaggagctctactattaattttaaccctcatgcttctagtcctattctcaccggacctgcttggag acceagacaactatacaccagcaaacccacttaatacacccccacatatcaagcccgaatgatacttcctatttgcatacgcaatcctccgatcaattcctaacaaactaggagg

10 Table 2 presents the alignment of the above fragment of cytochrome b gene of 221 animal species. Each species in table 2 has been represented by a unique code, which is decoded in Table 1. We selected these species to represent the vast range of animal families of distant orders. Of 221 species, about 65 were the protected/endangered or rare species listed in Wildlife (Protection) Act, 1972 (Central Act NO 53 of 1972). These species are marked with symbol (*) in Table 1. The NCBI accession number refers to its registration number in NCBI database and the number in superscript represent the reference cited. Based on the aligned cytochrome b sequences of different 221 animal species the primers designed were as follow:

Primers name

Sequence (5'-3')

20 'mcb398'

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"TACCATGAGGACAAATATCATTCTG"

'mcb869'

"CCTCCTAGTTTGTTAGGGATTGATCG"

Tables 2, 10 and 11, respectively, demonstrates that the 3' ends of the primers are highly conserved amongst all the animal species analyzed in-silico (In total 221 animal species listed in Table 1 and about 500 species listed in Tables 10 and 11, respectively) Also, the 5' end of the primers were selected within the conserved region of cytochrome b gene to improve the probability and stability of match of the primers to their target sequences (i.e. the above mentioned 472 bp fragment of cytochrome b gene). The primers were thoroughly checked for internal stabilities, loop or dimmer formation using different software viz.. 'Amplify (1.2)', 'Primer3' (http://www.genome.wi.mit.edu/cgi-<u>bin/primer/primer3.cgi</u>) as well as manually. We assigned the P,S score (P=Probability of match, S=Stability of match) to the primers for each template using the software Amplify (1.2). The higher scores of P and S ensure a good amplification if all other conditions standard (which are mentioned under 'Example 3') are optimum. The Highest score for 'mcb398' was 98,63 (i.e. the situation where the primer has perfect match with template);

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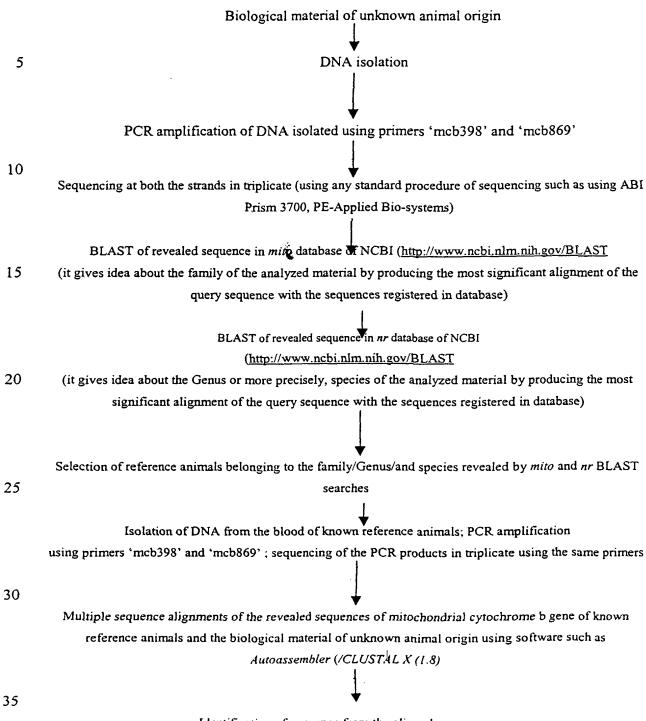
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however, the highest P, S for 'mcb869' was recorded as 98, 68 for a complete match between the primer and template. The lowest P,S score observed for 'mcb398' was 81,50 for species Talpa europaea whereas 'mcb869' had a high P, S score for this species (92, 57). The another species which have lowest P, S score for one of the two primers were Eumeces egregious and Equus ainus. Eumeces egregious had P, S score 86, 55 and 73,51 for 'mcb398' and 'mcb869', respectively; however, the P, S score of Equus ainus was calculated as 91,61 and 73, 51 for 'mcb398' and 'mcb869', respectively. All other animals had higher P, S scores then the above mentioned species. To ensure that these primers would work efficiently with the DNA template from the animals having the lowest P, S score for one of the primers, we designed an another experiment to validate the lower limits of one of the two primers sufficient for efficient amplification in PCR. We designed another primer pair (AFF= 5'tagtagaatgaatctgaggagg3' and an AFR=5'atgcaaataggaagtatcattc3'.) having more mis-pairing at their annealing sites (but not at ends), therefore have less internal stability and lower P, S scores for its templates (listed in Table 9). The P,S scores of 'AFF' and 'AFR' were as calculated as low as 41 and 49 for Platanista gangetica and Sus scrofa These species were amplified efficiently using the primers 'AFF' and 'AFR' (results shown in Figure 3) (keeping all other conditions standard i.e. the conditions mentioned in 'Example 3'). The lowest P,S scores (86, 55 and 73,51 for species Eumeces egregious) for our primers 'mcb398' and 'mcb869', respectively, were higher then the above range of combined P, S scores of 'AFF' and 'AFR' for species Sus scrofa (87, 52 and 87, 41), which was efficiently amplified by the primers 'AFF' and 'AFR'. It gives an indication that the primers 'mcb 398' and 'mcb 869' would work with all the species including Eumeces egregious efficiently to give rise to the expected product in PCR. This experiment confirmed that the primers 'mcb398' and 'mcb 869' are capable of amplifying the cytochrome b fragment of most of the animal species in a universal manner.

For further confirmation of universal nature of our primers, we blasted the sequence of our primers against the *mito* and *nr* databases of NCBI using BLAST software. The results of these analyses are shown in Tables 10, and 11, respectively.

Finally, the universal nature of the primers was tested in our laboratory with some more animal species listed in Table 12. These primers amplified all the animal species efficiently, giving rise to the band of expected size (472 bp). The results are shown in Figure 4. This experiments substantiated the results of P,S analysis and other *in-silico* analyses to show that the primers 'mcb398' and 'mcb 869' are universal primers.

The flow chart of establishing identity of the species of biological material of unknown animal origin using primers 'mcb398' and 'mcb869'



Identification of sequence from the aligned sequences

that is homologous (or significantly similar) to the cytochrome b gene sequence of the DNA obtained from biological material of unknown animal origin.

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The species of homologous sequence would be the species of the biological material under investigation

Examples

Example 1

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Example for identification of a fragment of cytochrome b gene fulfilling the requirements of columns 1, 2 and 3 mentioned under sub-heading 'Objectives of invention' of heading 'Brief summary of invention'

The cytochrome b molecule has very vastly been used in molecular taxonomic studies. Being a slow evolving gene, It has a tremendous information in its nucleotide sequences to distinguish the animals to their family, genus and species sources 1-65. A vast database of the sequences of cytochrome b gene of different animal species has accumulated in the nr and mito databases of NCBI. We have explored these qualities of cytochrome b gene to establish the identity of confiscated remains of unknown animal origin to its family, genus and species sources. For this purpose, we have identified a fragment of cytochrome b gene which is highly polymorphic inter-specifically, however, it is monomorphic among the individual of same species, therefore it can group the individual of an unknown species with the known individuals of reference species to which it belongs. In order to amplify this fragment from DNA isolated form any unknown origin, it was necessary that it remain flanked with the highly conserved sequences amongst a vast range of animal families. To identify such a unique fragment within the cytochrome b gene, we aligned the sequences of 221 distantly related animal species (listed in Table 1) representing various families using software CLUSTAL X (1.8). These sequences were obtained from public database NCBI (http://www.ncbi.nlm.nih.gov). The aligned data was examined carefully for the conserved sites amongst all the species included in in-silico analysis. We identified a fragment (472 bp) of cytochrome b gene that was fulfilling all the requirements mentioned above and also under column 1, 2 and 3 of sub-heading 'Objectives of invention'.

As for the identity of this fragment we would like to mention that it includes the nucleotides between 398 to 869 in Antilope cervicapra and Felis catus; however, 399 to 870 in Homo sapiens sapiens species. Except at few positions marked as star (*) in Table 2, the nucleotide sequences of this fragment are highly variable amongst the animal species, giving rise to their unique molecular signature. These molecular signatures are characteristic of its species and form the basis of revealing the identity of the biological material of an unknown animal origin by the procedure invented by us. Considering Antilope cervicapra as a representative species, the sequence of this fragment is mentioned herewith:

Mitochondrial cytochrome b gene sequence (398-869 bp) of Antilope cervicapra

"taccatgaggacaaatatctttttgaggagcaacagtcatcaccaatctcctttcagcaatcccatacatcggtacaaacctagtaga atgaatctgaggagggttctcagtagataaagcaacccttacccgatttttcgccttccactttatcctcccatttatcattgcagccctt accatagtacacctactgtttctccacgaaacaggatccaacaaccccacaggaatctcatcagacgcagacaaaattccattccac ccctactacactatcaaagatatcctaggagctctactattaattttaaccctcatgcttctagtcctattctacacggacctgcttggag acccagacaactatacaccagcaaacccacttaatacacccccacatatcaagcccgaatgatacttcctatttgcatacgcaatcct ccgatcaattcctaacaaactaggagg"

Example 2:

Example for development of universal primers to amplify the fragment identified mentioned under 'Example 1'.

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A pair of universal primer was designed which has the following features:

- 1. It targets the fragment identified (mentioned under 'Example 1') to amplify it in polymerase chain reaction (PCR).
- 2. Its 3' and 5' ends that are highly conserved (marked as star (*) in Table 2), amongst a vast range of animal species ensuring the amplification of the fragment mentioned above in a universal manner. The sequencing of the fragment amplified by these primes reveals the molecular signature of the species of analyzed material, which on comparison with the sequences of the known reference animals reveals the identity of the species of unknown biological material under investigation.
- 3. The tm (melting temperature) of both primers was almost similar (about 58 degree centigrade) ensuring the significant annealing of both the primers to its template, therefore significant amplification of targeted region in PCR.
 - 4. The internal stability and P, S, score of the primers were ensured higher while designing it. The possibilities of internal loop formation, dimmer formation etc were also excluded by selecting its sequence uniquely. This ensured that the primer would be a good primer to be used in PCR for amplification of DNA from unknown animal origin.
 - 5. The 3' end of the primers were ensured to have either 'G' or 'C' to increase the probability of strong bonding at its 3'ends, which is necessary for efficient amplification of DNA template in PCR. It also strengthens the universal nature of the primer.
 - 6. The sequences of the primers were ensured to be unique so that it does not give rise to non-specific and spurious products in PCR leading to confusion. It improved the efficiency and quality of the technique invented by us.

7. These primers were named as 'mcb398' and 'mcb869' because of its property to amplify a region of mitochondrial cytochrome b gene between nucleotides 398 to 869 of Antilope cervicapra, a representative animal species for this invention. We took this animal species as representative species because the idea of developing such a novel primers came in the mind of inventors while they were working on the genome of this animal in Centre for Cellular and Molecular Biology, Hyderabad, India.

8. The sequences of the universal primers invented are as follows:

Primers name

Sequence (5'-3')

10 'mcb398'

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"TACCATGAGGACAAATATCATTCTG"

'mcb869'

"CCTCCTAGTTTGTTAGGGATTGATCG"

Example 3:

Example for development of universal PCR conditions to ensure the amplification of a template of any unknown origin in PCR, hence strengthening the universal nature of the technique invented by us

The PCR conditions developed had the following unique features:

- 1 These were capable of amplifying the DNA template of any animal origin in an universal manner using the universal primers mentioned under 'Example 2'.
- 2. The conditions were selected to ensure the comparable annealing temperature for both the primers i.e. 'mcb398' and 'mcb869'.
- 3. The PCR conditions standardized herewith are universal; therefore, the possibility of PCR failure with a template of unknown origin due to non-standard conditions is excluded. It ensures the universal nature of our technique to be used in wildlife forensics.
- 4. The universal conditions mentioned above are:

Amplification reactions should be carried out in 20 μl reaction volume containing approximately 20 ηg of template DNA, 100μm each of dNTPs, 1.25 pmole of each primer, 1.5mM MgCl₂, 0.5 unit of Ampli*Taq* Gold (Perkin-Elmer-Cetus, USA) DNA polymerase and 1X PCR buffer (10mM Tris-HCl, pH 8.3, and 50mM KCl). The amplification profiles followed should be: an initial denaturation at 95°C for 10 min, followed by 35 cycles each of denaturation at 95°C for 45 s, annealing at 51°C for 1 min, and extension at 72°C for 2 min. The extension step at 35th cycles should be held for 10 min.

Example 4:

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Establishing the universal nature of our primer and experimental evidences to demonstrate the universal nature of primers:

The universal nature of the primers 'mcb398' and 'mcb 869' was ensured by the following measures:

(a) Selecting the primers from the aligned cytochrome b gene sequences of 221 animal of distantly related species:

The cytochrome b gene sequences (1140 bp) were aligned using software CLUSTAL X (1.8). The region of cytochrome b gene that was most conserved amongst 221 animal species was selected to design the primers.

(b) Selecting the 3' and 5' ends of the primers at the highly conserved positions of cytochrome b gene:

The 3' and 5' ends of the primers were ensured to anneal to a highly conserved position amongst 221 animal species representing a vast range of animal families. It was done to ensure an efficient amplification of all the species in PCR. These positions are shown with star (*) mark in Table 2.

(c) Ensuring either 'G' or 'C' at the 3' end of the primers:

It was ensured the primers to have either 'G' or 'C' at its 3' ends as these are the nucleotides that ensure the strong bonding at the 3' ends of the primers due to three hydrogen bonds while pairing with each other. The strong bonding at 3' ends nelps the primers to anneal properly with its template resulting in significant amplification in PCR.

(d) Selecting the sequences of the primers to ensure a higher internal stability, higher P, S score, and no primer dimmer and loop formation:

The sequences of the primers were selected to have a high P, S score for a vast range of animal species (Shown in Table 1). The care was taken to exclude the possibilities of loop or primer dimmer formation that could reduce the efficiency of the primers in PCR.

(e) Selecting the sequence of the primers with a comparable melting temperature:

The sequences of the primers were selected to have a comparable melting temperature so that these could work together to amplify a DNA template in PCR at a similar annealing temperature. The melting temperature of both the primers was about 58 degree centigrade and the annealing temperature used in PCR is 51 degree centigrade.

Experimental evidences to demonstrate the universal nature of primers:

(1) Evidence from In-silico analysis:

(a) Selecting the primers within the most conserved region of mitochondrial cytochrome b

As mentioned above, the primers were designed to anneal within a highly conserved region of mitochondrial cytochrome b gene fragment of 472 bp. Table 2 presents the alignment of the above fragment of cytochrome b gene of 221 animal species representing a vast range of animal families. The conserved positions of nucleotide sequences are shown with star (*) mark in Table 2

Table 2 also demonstrates that the 3' ends of the primers are highly conserved amongst all the animal species analyzed *in-silico*. In the aligned sequences, the conserved nucleotides are marked with symbol (*). Also, the 5' end of the primers were selected within the conserved region of cytochrome b gene to improve the probability and stability of match of the primers to their target sequences (i.e. the above mentioned 472 bp fragment of cytochrome b gene). The primers were thoroughly checked for internal stabilities, loop or dimmer formation using different software viz., 'Amplify (1.2)', 'Primer3' (http://www.genome.wi.mit.edu/cgi-bin/primer/primer3.cgi) as well as manually.

(b) P, S, score analysis:

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We assigned the P,S score (P=Probability of match, S=Stability of match) to the primers for each template using the software Amplify (1.2). The higher scores of P and S ensure a good amplification if all other conditions standard (which are mentioned under 'Example 3') are optimum. The Highest score for 'mcb398' was 98,63 (i.e. the situation where the primer has perfect match with template); however, the highest P, S for 'mcb869' was recorded as 98, 68 for a complete match between the primer and template. The lowest P,S score observed for 'mcb398' was 81,50 for species Talpa europaea whereas 'mcb869' had a high P, S score for this species (92, 57). The another species which have lowest P, S score for one of the two primers were Eumeces egregious and Equus ainus. Eumeces egregious had P, S score 86, 55 and 73,51 for 'mcb398' and 'mcb869', respectively; however, the P, S score of Equus ainus was calculated as 91,61 and 73, 51 for 'mcb398' and 'mcb869', respectively. All other animals had higher P, S scores then the above mentioned species. To ensure that these primers would work efficiently with the DNA template from the animals having the lowest P, S score for one of the primers, we designed an another experiment to validate the lower limits of one of the two primers sufficient for PCR. efficient amplification in We designed an another (AFF= 5'ctagtagaatgaatctgaggagg^T and AFR= 5'tatgcaaataggaagtatcattc³'.) that have more mis-pairing at their annealing sites (but not at ends), therefore have less internal stability

and lower P, S scores for its templates (listed in Table 9). The P,S scores of 'AFF' and 'AFR' were as calculated as low as 41 and 49 for *Platanista gangetica* and *Sus scrofa* These species were amplified efficiently using the primers 'AFF' and 'AFR' (results shown in Figure 3) (keeping all other conditions standard i.e. the conditions mentioned in 'Example 3'). The lowest P,S scores (86, 55 and 73,51 for species *Eumeces egregious*) for our primers 'mcb398' and 'mcb869', respectively, were higher then the above range of combined P, S scores of 'AFF' and 'AFR' for species *Sus scrofa* (87, 52 and 87, 41), which was efficiently amplified by the primers 'AFF' and 'AFR'. It gives an indication that the primers 'mcb 398' and 'mcb 869' would work with all the species including *Eumeces egregious* efficiently to give rise to the expected product in PCR. This experiment confirmed that the primers 'mcb398' and 'mcb 869' are capable of amplifying the cytochrome b fragment of most of the animal species in a universal manner.

© BLAST analysis:

The sequences of primers 'mcb398' and 'mcb869' were blasted against mito and nr databases of NCBI to see its significant alignments with the sequences registered in GenBank. As expected, the most significant alignments of the sequences were found with the cytochrome b gene regions (within the 472 bp fragment mentioned in 'Example 1') of different animal species. This analysis also showed that the 3' as well as 5' ends of the primers were highly conserved amongst a vast range of animal species, confirming the universal nature of the primers (Tables 10 and 11, respectively)

(2) Evidence from bench work/experiments done in laboratory conditions:

The DNA from different animals belonging to distantly related species (mentioned in Table 12) was isolated and subjected to PCR amplification using the primers invented by us i.e. the primers 'mcb398' and 'mcb869' The PCR products amplified were resolved in agarose gel by electrophoresis and visualized under UV light. The PCR products of expected size (472bp) were obtained from all the animals confirming the universal nature of our primers. These results are shown in Figure 4.

Example 5:

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Example to establish the identity of confiscated remains from unknown animal origin using the universal primers 'mcb398' and 'mcb869'.

The step-vise procedure to establish the identity of the biological material from an unknown animal source is mentioned below:

Biological material of unknown animal origin

DNA isolation

PCR amplification of DNA isolated using primers 'mcb398' and 'mcb869'

Sequencing at both the strands in triplicate (using any standard procedure of sequencing such as using ABI Prism 3700, PE-Applied Bio-systems)

BLAST of revealed sequence in mito database of NCBI

(http://www.ncbi.nlm.nih.gov/BLAST

(it gives idea about the family of the analyzed material by producing the most significant alignment of the query sequence with the sequences registered in database)

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BLAST of revealed sequence in nr database of NCBI

(http://www.ncbi.nlm.nih.gov/BLAST

(it gives idea about the Genus or more precisely, species of the analyzed material by producing the most significant alignment of the query sequence with the sequences registered in database)

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Selection of reference animals belonging to the family/Genus/and species revealed by mito and nr BLAST searches

Isolation of DNA from the blood of known reference animals;
PCR amplification using primers 'mcb398' and 'mcb869'; sequencing of the PCR
products in triplicate using the same primers

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Multiple sequence alignments of the revealed sequences of mitochondrial cytochrome b gene of known reference animals and the biological material of unknown animal origin using software such as Autoassembler/CLUSTAL X (1.8)

Identification of sequence from the aligned sequences

that is homologous (or significantly similar) to the cytochrome b gene sequence of the

DNA obtained from biological material of

unknown animal origin.



The species of homologous sequence would be the species of the biological material under investigation



Application of the above information for the objectives mentioned in columns 7-13 under sub-heading 'Objective of invention' of heading 'Summary of invention'

Example 6:

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The actual execution of the technique invented

As a first application and to demonstrate the ease and utility of this method, we investigated a case of forensic identification submitted at our laboratory to seek scientific opinion on animal hunting evidence. In this case, we received the half burned remains of an unknown animal, confiscated by the crime investigation agencies. The DNA was isolated from the above material following standard methods⁷⁴ and subjected to PCR amplification using the primers mentioned above (viz., 'mcb398' and 'mcb869'). Amplification reactions were carried out in 20 µl reaction volume containing 20 ηg of template DNA, 100µm each of dNTPs, 1.25 pmole of each primer, 1.5mM MgCl₂, 0.5 unit of AmpliTaq Gold (Perkin-Elmer-Cetus, USA) DNA polymerase and 1X PCR buffer (10mM Tris-HCl, pH 8.3, and 50mM KCl). The amplification profiles followed were: an initial denaturation at 95°C for 10 min, followed by 35 cycles each of denaturation at 95°C for 45 s, annealing at 51°C for 1 min, and extension at 72°C for 2 min. The extension step at 35th cycles was held for 10 min.

The PCR products obtained were sequenced in automated work station (ABI Prism 3700, PE-Biosystems) on both strands in triplicate and the sequence resolved (328 bp, shown in Figure 1a) was blasted against *mito* databases of NCBI using BLAST program⁷³. The most significant alignment (bits Value 365, E value e⁻¹⁰¹) of this sequence was produced with the cytochrome b gene sequence of *Felis catus*, (Table 3) indicating that species of

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analyzed material belongs to family felidae. Further, the above sequence revealed from the confiscated remain was blasted against nr databases of NCBI using BLAST program. The most significant alignment (bits Value 603, E value e⁻¹⁷⁰) of this sequence was produced with the cytochrome b gene sequence of Panthera pardus (Table 4), indicating the identity of the analyzed material as that of a Panthera pardus source. Based on this information, we selected the reference animals listed in Table 5 representing different species and subspecies of felidae. The DNA isolated from reference animals was amplified and sequenced on both strands in triplicate using the primer pair mentioned above. Consensus sequences obtained were aligned using program CLUSTAL X (1.8) (Table 6). Sequence comparisons identified 113 variable sites in total amongst all animals analyzed (Table 7). Pair-vise comparisons of sequences were performed to find out the variation among different animals investigated. All the species investigated were differentiated by a their unique nucleotides sequences. The molecular signatures of different reference animals were compared with the molecular signature of the confiscated skin 'adil.flesh'. Table 7 demonstrate that the maximum similarity of the adil.flesh with 'gzll' i.e. known Leopard (Panthera pardus) species, indicating the identity of the adil flesh, the confiscated skin, as that of a Panthera pardus origin. We also calculated the similarity matrix showing the pairvise similarity amongst the animal species under investigation using PHYLIP software This matrix is shown in Table 8. It demonstrates that the animals belonging to different species had more variation; however, the animals of same species had maximum similarity among their cytochrome b sequences. The cytochrome b gene sequence of DNA isolated from confiscated material had maximum similarity with the sequences obtained from known Leopard source(99.7%, and 98.2 with 'gz11' and 'gz21', respectively); establishing the identity of the source of confiscated material as that of a Normal leopard (Penthera pardus) species. The step-vise procedure involved in above analysis is illustrated in Figure la, lb and lc, respectively.

Thus, the primers invented by us can generate the molecular signature from any biological material of unknown animal origin, which actually is the characteristic of its family, genus and more precisely, the species. When these signatures are compared *in-silico* with the signatures already available in public databases (viz., GenBank, NCBI database etc) using *BLAST* software⁷³, it indicates identity of the family, genus or species of the analyzed material, which in turn is confirmed practically by comparing with the reference animals of the revealed family, genus or species, by including them in the further analysis by the primers 'mcb398' and 'mcb869'. Application of the information revealed could be in

fulfilling the requirements of objectives mentioned in columns 7-13 under sub-heading 'Objective of invention' of heading 'Summary of invention'

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The method of the invention can be used to establish the identity of confiscated animal parts and products is one of the key requirements of wildlife identification in forensics. It is needed to establish the crime with the criminal beyond a reasonable doubt to avoid the human violation of wildlife resources. Various morphological biochemical and molecular approaches have been given for this purpose; however, none of the current methods is universally applicable to detect the mutilated animal remains of unknown origin. We have identified a fragment on the mitochondrial cytochrome b gene, which has enormous information to differentiate among various animal species back to the family, genus and species sources. We have also found that this fragment is flanked by the highly conserved sequences amongst a vast range of animal species. We invented a pair of universal primer that can amplify this fragment of DNA isolated from the biological material of an unknown animal origin in polymerase chain reaction (PCR) to reveal its identity at species and sub-species sources. This novel invention has great potential to revolutionize the whole scenario of wildlife forensic identification and crime investigation.

Table 1. The animal species included in the study for in-silico analysis

SN.	Code	Name	NCBI accession #	^{\$} P,S/F	*P,S/R
1	aep.mel	Aepyceros melampus	AF0362891	97, 60	94, 62
2	ore.ore	Oreotragus oreotragus	AF0362881	88, 52	94, 62
3	add.nas	Addax nasomaculatus	AF034722 ²	97, 60	95, 66
4	ory.dam	Oryx damah	AJ2226851	90, 58	95, 66
5	hip.equ	Hippotragus equinus	AF022060 ³	98, 63	85, 55
6	alc.bus	Alcelaphus buselaphus	AJ222681 ^t	97, 60	98, 68
7	sig.lic	Sigmoceros lichtensteinii	AF034967 ⁴	97, 60	98, 68
8	bea.hun	Beatragus hunteri	AF0349684	97, 60	94, 62
9	dam.lun	Damaliscus lunatus	.AF0166353	97, 60	77,55
10	con.tau	Connochaetes taurinus	AF016638 ³	82, 56	93, 62
11	bis.bon	Bison bonasus	Y150055	90, 58	87, 63
12	bos.gru	Bos grunniens *	AF0916316	90, 58	94, 62
13	bos.tra	Bos tragocamelus*	AJ2226791.	90, 58	95, 66
	buba.bub	Bubalus bubalis *	D34637 ⁷	97, 60	93, 64
	bub.min	Bubalus mindorensis	D82895 ⁸	97, 60	87, 62
	tralang	Tragelaphus angasii	AF0916336	97, 60	87, 63
17	tracur	Tragelaphus eurycerus	AF0362761	90, 58	97, 64
_	nem.cau	Nemorhaedus caudatus*	U17861°	95, 61	93, 59
19	pse.nay	Pseudois nayaur	AF034732 ²	89, 55	89, 59
20	amm.ler	Ammotragus lervia	AF034731 ²	94, 58	97, 63
21	cap.fal	Capra falconeri*	D8420210	98, 63	95, 66
22	cap.ibe	Capra ibex*	AF034735 ²	98, 63	89, 58
23	hem.jem	Hemitragus jemlahicus*	AF034733 ²	95,61	90, 61
24	пир.руг	Rupicapra pyrenaica	AF034726 ²	95, 61	89, 59
	rup.rup	Rupicapra rupicapra	AF034725 ²	95, 61	94, 64
	pan.hod	Pantholops hodgsoni	AF034724 ²	98, 63	95, 66
	bud.tax.tax	Budorcas taxicolor taxicolor*	U17868°	90, 53	95, 66
28	ovi.amm	Ovis ammon*	AF034727 ²	98, 63	97.64
29	ovi.vig	Ovis vignei*	AF034729 ²	98, 63	97, 64 94, 63
30	cap.cri	Capcornis crispus •	AJ30450211	98, 63	94, 63 92, 61
- 31	ovi.mos	Ovibos moschatus	U17862°	98, 63	94, 62
	ore.ame	Oreamnos americanus	AF19063212	98, 63 97, 58	90, 61
33	c cp .dor	Cephalophus dorsalis	AF091634 ⁶	97, 38 97, 60	88, 53
	ccp.max	Cephalophus maxwellii	AF096629 ¹³	97, 60 95, 61	93, 59
35	alc.alc	Alces alces	AJ000026 ¹⁴ AJ000028 ¹⁴	97, 60	90, 63
36	hyd.ine	Hydropotes inermis	AF042718 ¹⁵	90, 58	93. 64
37	mun.mun	Muntiacus muntjak*	AB02109816	98, 63	82, 59
38	cer.ele.kan	Cervus elaphus kansuensis*	AB02109716	98, 63	82. 59
39 40	cer.ele.xan	Cervus elaphus xanthopygus * Cervus elaphus canadensis *	AB02109616	98, 63	90.61
41	cer.ele.can cer.nip.ce	Cervus nippon centralis	AB02109415	98, 63	90.61
	cer.nip.ye	Cervus nippon vesoensis	AB0210951*	98, 63	90.61
	cer.nip.ke	Cervus nippon keramae	AB021091"	93, 63	90.61
-	···	• • •			

4.1	cec ain au	Cervus nippon pulchellus	AB02109016	98, 63	90, 61
	cer.nip.pu	Cervus nippon nippon	AB02109316	98, 63	90, 61
45	•		AB02109916	98, 63	90, 61
46	· · - ·	Cervus elaphus scoticus	AJ00002214	98, 63	88, 53
47	·	Cervus dama	AJ000022	98, 63	89, 57
48		Rangifer tarandus	AF026888 ¹⁷	90, 59	90, 61
49		Moschus fuscus	AF026889 ¹⁷	90, 59	90, 61
50		Moschus leucogaster*	AF026887 ¹⁷	90, 59	90, 61
51		Moschus chrysogaster*	AF026886 ¹⁷	90, 59	90, 61
52		Moschus berezovskii*	AF026883 ¹⁷	90, 59	92, 61
53		Moschus moschiferus*		91,61	95, 66
54		Kobus ellipsiprymnus	AF022059 ³	91,61	83, 56
55	kob.meg	Kobus megaceros	AJ2226861	91,61	94, 62
56		Redunca arundinum	AF096628 ¹³	89, 57	94, 62
57	red.ful	Redunca fulvorufula	AF036284 ¹	•	94, 62
58	neo.mos	Neotragus moschatus	AJ2226831	89, 57	
59	pel.cap	Pelea capreolus	AF0220551	91,61	90, 61
60	anticer	Antilope cervicapra*	AF022058 ³	82, 56	93, 64
61	sai.tat	Saiga tatarica	AF064487 ¹⁸	91,61	92, 61
62	gaz.dam	Gazella dama	AF0259543	91,61	92, 61
63	our.our	Ourebia ourebi	AF036288 ¹	82, 56	82, 59
64	gaz.gaz	Gazela gazella*	AJ2226821	91,61	89, 57
65	rap.mel	Raphicerus melanotis	AF022053 ³	81, 54	80, 50 97, 65
66	mad.kir	Madoqua kirkii	AF022070 ³	90, 58	98, 68
67	antame	Antilocapra americana	AF0916296	98, 63	96, 59
68	tra.jav	Tragulus javanicus *	D32189 ¹⁹	86, 57	93, 58
69	tra.nap	Tragulus napu *	X56288 ²⁰	81, 52	93, 33 97, 61
70	bal.acu	Balaenoptera acutorostrata	X75753 ²¹	89, 56	93, 59
71	bal.bon	Balaenoptera bonaerensis	X75581 ²¹	89, 56 89, 56	93, 59
72	bal.bor	Balaenoptera borealis*	X75582 ²¹	89, 56	83, <i>5</i> 4
73	bal.edi	Balaenoptera edeni	X75583 ²¹	97, 61	86, 57
74	esc.rob	Eschrichtius robustus	X75585 ²¹	97, 57	93, 59
75	bal.mus	Balaenoptera musculus*	NC_001601 ²² X75584 ²¹	97, 61	94, 63
76	meg.nov	Megaptera novaeangliae	NC_001321 ²³	97, 57	94, 63
77	bal.phy	Balaenoptera physalus*	X75586 ²¹	93, 55	91,53
78	cap.mar	Caperea marginata	AF084073 ²⁴	85, 51	38, 55
	cep.com	Cephalorhynchus commersonii Cephalorhynchus eutropia •	AF084072 ²⁴	85, 51	92, 59
80	cep.eut	Lagenorhynchus obliquidens	AF084067 ²⁴	94, 59	92, 59
81 82	lag.obl cep.hea	Cephalorhynchus heavisidii	AF084070 ²⁴	89, 56	97, 63
83	cep.hec	cephalorhynchus hectori*	AF08407124	89, 56	92. 59
84	lag.aus	Lagenorhynchus australis	AF08406924	86.54	92, 59
85	lag.cru	Lagenorhynchus cruciger	AF08406824	86. 54	92, 59
86	lag.obs	Lagenorhynchus obscurus	AF08406624	86. 54	92, 59
	lis.bor	Lissodelphis borealis	AF08406434	85.51	92, 59
	lis.per	Lissodelphis peronii	AF08406514	86. 54	92, 59
	glo.mac	Globicephala macrorhynchus	AF084055*4	94, 59	88, 55
	glo.mel	Globicephala melas	AF08405614	94, 59	\$8, 55
	fer.att	Feresa attenuata*	AF08405224	94. 59	92, 59

			AF08405324	94, 59	88, 55
92	pep.ele	Peponocephala electra*	AF084059 ²⁴	97,61	89, 59
93	gra.gri	Grampus griscus	AF084057 ²⁴	94, 59	92, 59
94	pse.cra	Pseudorca crassidens*	AF084075 ¹⁴	98, 63	89, 59
95	lag.acu	Lagenorhynchus acutus	AF084061 ²⁴	86, 57	82, 52
96	arci.bre	Orcinus orca	AF084063 ²⁴	86. 57	91, 54
97	orca.bre	Orcaella brevirostris	AF084087 ²⁴	96, 54	97, 63
98	del.cap	Delphinus capensis	AF084085 ²⁴	97, 57	97, 63
99	del.tro	Delphinus tropicalis		97, 57	97, 63
100	del.del	Delphinus delphis	AF084085 ²⁴	97, 57	97, 63
101	sten.cly	Stenella clymene	AF084083 ²⁴	97, 57	97, 66
102	sten.coe	Stenella coeruleoalba	AF084082 ²⁴	97, 57	97, 63
103	tur.adu	Tursiops aduncus	AF084092 ²⁴	97, 57	97, 63
104	sten.fro	Stenella frontalis	AF084090 ²⁴	97, 57	88, 59
105	saus.chi	Sousa chinensis	AF084080 ²⁴	97, 61	97, 63
106	sten.lon	Stenella longirostris	AF084103 ²⁴	97, 57	96, 59
107	turs.tru	Tursiops truncatus	AF084095 ²⁴	97, 61	97,66
108	lage.alb	Lagenorhynchus alborostris	AF084074 ²⁴	97, 61	94, 64
109	sten.bre	Steno bredanensis	AF084077 ²⁴	97, 61	97,63
110	sota_flu	Sotalia fluviatilis	AF304067 ²⁵	97, 61	95, 66
111	del.leu	Delphinapterus leucas	U72037 ²⁶	97, 61	95, 66
112	mono.mon	Monodon monoceros	U72038 ²⁶ AF304070 ²⁵	97, 61	86, 59
113	plat.gan	Platanista gangetica*	X92543 ²⁷	97, 61	86, 59
114	platmin	Platanista minor*	X92543 U72040 ²⁶	97, 59	90, 63
115	kogi.bre	Kogia breviceps	AF304072 ²³	96, 55	92, 63
	kogi.sim	Kogia simus	AF304072 ²⁵	97, 57	80, 58
	phys.cat	Physeter catodon	AF304071 ²⁵	89, 56	88, 53
	lipo.vex	Lipotes vexillifer*	AF084051 ²⁴	87, 49	92, 62
	phoc.sin	phocoena sinus	X92541 ²⁷	96, 55	90, 59
	bera.bai	Berardius bairdii	X92540 ²⁷	97,61	89,57
	ziph.car	Ziphius cavirostris Mesoplodon europaeus	X92537 ²⁷	97, 57	90, 61
	meso.eur	Mesoplodon bidens	X92538 ²⁷	97, 61	92, 61
	meso.bid	Mesoplodon densirostris	X92536 ²⁷	91,61	94, 63
	meso.den	Hyperoodon ampullatus*	X92539 ²⁷	97,61	90,65
	hype.amp	Mesoplodon peruvianus	AF304074 ²⁵	97, 61	86, 58
	meso.per pont.bla	Pantaparia blainvillei	AF304069 ²⁵	92, 59	88, 55
	hipp.amp	Hippopotamus amphibius	Y0881329	92, 58	95, 66
	hex.lib	Hexaprotodon liberiensis	Y0831429	98, 63	97, 66
	rhin.son	Rhinoceros sondaicus*	AJ245725 ³⁰	90, 59	87, 61
	cera	Ceratotherium simum	MC_00180215	90, 59	90, 63 86, 57
	dic.sum	Dicerorhinus sumatrensis	AJ24572310	90, 59	73, 51
133	eduu	Equus asinus	NC_00178531	91,61 39,56	85. 56
134	baby.bab	Babyrousa babyrussa	Z50106 ¹¹	90, 59	87, 54
135	phac.afr	Phacochoerus africanus	Z50090 ¹³	97. 57	83.54
	sus.scr.ew	Sus scrofa haplotype EWB3*	AF136549 ¹⁴	97, 57	85.55
	sus.bar	Sus barbatus	Z50107 ¹¹ U06429 ¹¹	89, 55	85. 53
	lama.gla	Lama glama	A08215.	83, 54	86. 57
139	lama,gua	lama guanicae	1.0001 =	•	

				89, 55	85, 53
140	vic.vic	Vicugna vicugna	U06430 ³⁵	94,58	86, 58
141	cam.bac	Camelus bactrianus	U06427 ³⁵	97, 60	87, 64
142	arc.for	Arctocephalus forsteri	XS2293 ³⁶	94, 58	87,64
143	arc.gaz	Arctocephalus gazella	X82292 ³⁶	97, 57	86, 57
144	eum.jub	Eumetopias jubatus	X82311 ¹⁶		86, 57
145	zal.cal	Zalophus californianus	XS2310 ³⁶	89, 55	81,52
	odo.ros	Odobenus rosmarus	X82299 ³⁶	91,61	87, 64
	pho.vit	Phoca vitulina	X32306 ¹⁶	90, 58	
	pho.fascia	Phoca fasciata	X82302 ³⁶	98, 63	95, 66
	pho.gro	Phaca graenlandica	X82303 ³⁶	92, 59	90, 61
	cys.cri	Cystophora cristata	X82294 ³⁶	89, 56	87, 64
	hyd.lep	Hydrurga leptonyx	X32297 ³⁶	89, 55	82, 54
	lep.wed	Leptonychotes weddelli	X72005 ³⁷	98, 63	91,66
	mir.leo	Mirounga leonina	X82298 ³⁶	89, 55	82, 59
	eri.bar	Erignathus barbatus	X82295 ³⁶	89, 56	87, 63
	mon.sch	Monachus schauinslandi	X72209 ³⁷	91,61	87, 60
	hela.mal	Helarctos malayanus*	U18899 ¹⁸	84, 54	90, 63
	sel.thi	Selenarctos thibetanus*	AB0209!0 ³⁹	89, 57	87, 64
	ail.ful	Ailurus fulgen*s	X94919 ¹⁰	93, 55	87, 64
159		Felis catus	NC_00170041	85, 56	90, 63
	can	Canis familiaris	NC_00200842	98, 58	84, 54
161		Talpa europaea	NC_00239143	81,50	92, 57
	gla.sab	Glaucomys sabrinus	AF011738**	90, 59	82, 54
	gla.vol	Glaucomys volans	AB03026145	90, 59	87, 60
	hyl.pha	Hylopetes phayrei*	AB03025945	91,61	81,50
	pet.set	Petinomys setosus "	AB030260 ⁴⁵	91,61	81, 50 87, 64
	bel.pea	Belomys pearsonii*	AB03026245	91,61	90, 63
	pte.mom	Pteromys momonga*	AB03026345	97, 61	87, 64
	gala.demi	Galagoides demidojj	AF27141146	97, 58	87,63
	pero.pot	Perodicticus potto	AF271413 ⁴⁶	97,60	90.61
	gala.mat	Galago matschiei	AF271409 ⁴⁶	97,60 07.57	95, 66
	gala.moh	Galago moholi	AF271410 ⁴⁶	97, 57 92, 58	87, 60
	oto.gar	Otolemur garnettii	AF27141246	97, 60	93, 59
173	lor.tar	Loris tardigradus*	U53581 ⁴⁷	97, 60 97, 60	95, 66
174	nyc.cou	Nycticebus coucang*	US3580 ⁴⁷	97,60	86, 59
175	mus	Mus musculus	NC_00156948	89, 57	80, 58
176	gon	Gorilla gorilla	NC_001645 ⁴ °	96, 55	84, 64
	homo	Homo sapiens sapiens	NC_001807 ³⁰	97, 60	89, 59
	dug.dug	Dugong dugong*	AB002412 ⁵²	97,60	76, 57
	ele.max	Elephas maximus*	AF013760'	97, 58	87, 63
	afr.con	Afropavo congensis	AF013763''	97, 57	87,63
	pavo.mut	Pava muticus*	AF2007225*	89.55	35, 57
	tra.bly	Tragopan blythii*	AF229837**	39, 55	86, 61
	tra.sat	Tragopan satyra* Tragopan cabott	AF200723 **	39, 55	\$6,61
	tra.cob	Ггадоран сантински *	AF028802''	89, 55	31,56
	tra.tem arg.arg	Argusianus argus	AF013761"	89, 55	87, 63
		Catrous wallicht	AF028792"	88. 24	85.57
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	Crossoptilon crossoptilan*	AF02879453	89,55	85, 57
			89.55	85, 57
•			80, 48	94, 64
			97.58	86, 61
			98. 63	85.57
	-		85. 56	82, 58
•				82, 52
ant.vir				87, 63
gru.ant.an		U11000		87, 63
gru.ant.gi		U1100+		87, 63
gru.any.sh		U11001	•	87, 63
gru.leu				87, 63
gru.can.pr				87, 63
gru.can.ro				87, 63
gru.can.ta		U275517	-	87, 63
gru.can.ca	Grus canadensis canadensis	U275543°	•	
gru.ame	Grus americana	U27555 ²⁸	•	87, 63
gru.gru	Grus grus		•	87, 63
-	Grus monacha*			87, 63
_	Grus nigricollis*			87, 63
-			•	87, 63
	Ciconia boyciana*		•	79, 60
-	Rhea americana	AF090339 ⁵⁹		79, 60
	Anthracoceros albirostris*	U89190 ⁶⁰	-	86, 59
		U83310 ⁶¹		86, 60
	-			85, 57
•				84, 52
•	• •			80, 51
•			•	94, 62
-	Smithornis sharpei			90, 61
	Vidua chalybeata	NC_00083023		87, 64
	and the second s	NC_002073 ⁶³		86, 57
,			-	94, 63
	Chelonia mydas *			94, 63
	Eumeces egregius	AB01660653	86, 33	73, 51
	cro.cro sym.ree bam.tho fra.fra ith.cru ant.par ant.vir gru.ant.an gru.ant.gi gru.any.sh gru.leu gru.can.ro gru.can.ta gru.can.ca gru.can.ca gru.ame gru.gru gru.mon gru.nig gru.jap cic.boy rhe.ame ant.alb fal.fam fal.ver fal.par fal.spa ayt.ame smi.sha vid.cha chry.pic emy.orb.ku che.mud eum.egr	sym.ree Syrmaticus reevesi bam.tho Bambusicola thoracica fra.fra Francolinus francolinus ith.cru Ithaginis cruentus ant.par Anthropoides paradisea ant.vir Anthropoides virgo gru.ant.an Grus antigone antigone gru.ant.gi Grus antigone gillae gru.any.sh Grus antigone sharpei gru.can.pr Grus canadensis pratensis gru.can.to Grus canadensis rowani gru.can.ta Grus canadensis rowani gru.can.ca Grus canadensis canadensis agru.ame Grus americana gru.gru Grus grus grus gru.mon Grus monacha gru.jap Grus japonensis cic.boy Ciconia boyciana ant.alb Anthracoceros albirostris fal.fam Falco femoralis fal.spa Falco sparverius ayt.ame Aythya americana smi.sha Smithornis sharpei vid.cha Vidua chalybeata chry.pic Chrysemys pic:a emy.orb.ku Emys orbicularis che.mud Chelonia mydas antigone grus cichenus che.mud Chelonia mydas antigone grus cichenus che.mud Chelonia mydas antigone cichenus cichemud Chelonia mydas antigone cichemus cichemud chelonia mydas antigone cichemus cichemud chelonia mydas antigone cichemus cichem	sym.ree Symaticus reevesi* AF028801 ³³ bam.tho Bambusicola thoracica* AF028790 ³³ fra.fra Francolinus francolinus AF013762 ³³ ith.cru Ithaginis cruentus* AF068193 ³³ ant.par Anthropoides paradisea U27557 ³⁶ ant.vir Anthropoides virgo U27545 ³⁶ gru.ant.an Grus antigone antigone U11060 ³⁷ gru.ant.gi Grus antigone sharpei U11061 ³⁷ gru.any.sh Grus antigone sharpei U17549 ³⁶ gru.can.pr Grus canadensis pratensis U27549 ³⁶ gru.can.pr Grus canadensis pratensis U27553 ³⁶ gru.can.to Grus canadensis rowani U27552 ³⁶ gru.can.ca Grus canadensis tabida U27551 ³⁶ gru.anme Grus americana U27555 ³⁶ gru.mon Grus monacha* U27546 ³⁶ gru.nig Grus nigricollis* U27544 ³⁶ gru.nig Grus japonensis U27545 ³⁶ gru.jap Grus japonensis U27550 ³⁶ cic.boy Ciconia boyciana* NC_002196 ³⁸ rhe.ame Rhea americana AF090339 ³⁹ rhe.ame Rhea americana W33311 ⁶¹ fal.par Falco peregrinus* U33307 ⁶¹ fal.par Falco peregrinus* U33307 ⁶¹ fal.spa Falco sparverius U33306 ⁶¹ syt.ame Aythya americana NC_000877 ⁶² smi.sha Smithornis sharpei NC_000879 ⁵⁹ vid.cha Vidua chalybeata NC_000879 ⁵⁹ vid.cha Vidua chalybeata NC_000830 ⁵⁹ cemy.orb.ku Emys orbicularis AB012104 ⁵⁵ AB012606 ⁴⁵ AB012104 ⁵⁵ AB012606 ⁴⁵	sym.ree Syrmaticus reevesi

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Table 2. Multiple sequence alignment of 471 bp fregment of mitochondrial cytochrome b gene of 221 animal species

PRIMER 'mcb398'	TACCATGAGGACAAATATCATTCTG	
aep.mel	TGCCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATTACAAATCTCGTCTCAGCAA	
		50
ore.ore	managa maga ang ang ang ang ang ang ang ang ang	ن ن
add.nas	The control of the second formation of the control	3.7
ory.dam	TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATCACCAACCTCCTCTCAGGAA TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATCACCAACCTCCTCTCAGGAA	50
hip.equ	TO THE TOTAL COLUMN TO THE	3 4
alc.bus	TGCCATGAGGACAAATATCATTCTGAGGGGCAACAGTCATCACCAATCTCCTCTCAGCAA	50
sig.lic	TGCCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATCACCAACCTCCTCTCAGCAA TGCCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATCACCAACCTCCTCTCAGCAA	60
bea.hun	TGCCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATCACTAACCTCCTCTCAGCAA TGCCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATCACTAACCTCCTCTCAGCAA	5 0
dam.lun	TGCCATGAGGACAAATATCCTTTTGAGGAGCAACAGTCATCAACCAAC	60
con.tau	TACCATGAGGACAAATATCCTTTGAGGAGCAACAGTCATCTACTAGCAACCTTCTCTCAGCAA	60
amm.ler	TACCATGAGGACAGATATCATTCTGAGGGGCAACAGTCATCACCAACCTTCTCTCAGCAA	60
pse.nay	TGCCATGAGGACAGATATCATTTTGAGGGGCAACAGTCATCACCAACCTTCTCTCAGCAA TGCCATGAGGACAAATATCATTTTGAGGGGCAACAGTCATCACCAACCTTCTCTCAGCAA	60
cap.ibe	TGCCATGAGGACAAATATCATTCTGAGGGGCAACAGTCATCACTAACCTTCTCTCAGCAA TACCATGAGGACAAATATCATTCTGAGGGGCAACAGTCATCACTAACCTTCTCTCAGCAA	60
hem.jem	TACCATGAGGACAAATATCATTCTGAGGGGCAACAGTCATGACCAACCTTCTCTCAGGAA TACCATGAGGACAGATATCATTCTGAGGGGCAACAGTCATGACCAACCTTCTCTCAGCAA	60
cap.fal	TACCATGAGGACAAATATCATTCTGAGGGGCAACAGTCATCACCAATCTCCTCTCAGCAA TACCATGAGGACAAATATCATTCTGAGGGGCAACAGTCATCACCAATCTCCTCTCAGCAA	60
rup.pyr		
rup.rup		
nem.cau		60
bud.tax.tax		• -
pan.hod		63
ovi.amm		
ovi.vig		- 1
cap.cri		
cvi.mos		
ore.ame		
cep.dor		
rep.del		-
bis.bon		
bos.gru		
bos.tra		
bub.min		
buba.bub		
tra.ang		
tra.eur	THE COLUMN TO SELECT THE TOTAL CONCENTRATION OF THE COLUMN TO THE COLUMN	
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red.ful	TGCCATGGGGACAAATATGCTTCTGAGGAGCAACAGTTATGAGTAACTTTGTGAGCAA	
neo.mos		
pel.cap		50
gaz.dam		
our.our		
ant.cer		
sai.tat		
mai.kir		
rap.mel		
gaz gaz		
ant.ame	TACCATGAGGACAAATATUU. JASUAGUAA LAGITATTA TTAACCTACTCTGAGGAA TACCATGAGGACAAATATGATGTGAGGAGGAGTACTGTGAGGAA	50
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ceclela kan	TACCATGAGGAGATATTA ITC COAGGAGGAGGAGGATATTATCAGGAGTTGTGTGAGGAA	

TACCATGAGGACAAATATCATTCTGAGGAGCAACGGTCATTACCAACCTTCTCTCAGCAA 60 cer.ela.xan TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATTACCAACCTTCTCTCAGCAA 60 cer.ela.can TACCATGAGGACAATATCATTCTGAGGAGCAACAGTCATTACCAACCTCCTCTCAGCAA 60 cer.nip.cent TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATTACCAACCTCCTCTCAGCAA 60 cer.nip.yes TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATTACCAACCTTCTCTCAGCAA 60 cer.nip.ker TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATTACCAACCTTCTCTCAGCAA 60 cer.nip.pul cer.nip.nip TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATTACCAACCTCCTCTCAGCAA 60 TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTCATCACCAACCTTCTCTCAGCAA 60 cer.ela.sco; TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTTATTACCAATCTTCTCTCAGCAA 60 cer.dam TACCATGAGGACAAATATCATTCTGAGGAGCAACAGTTATCACAAACCTCCTCTCAGCAA 60 ran.tar TACCTTGAGGACAAATATCTTTCTGAGGAGCGACAGTTATTACCAATCTTCTCTCAGCAA 60 mos.fus TACCTTGAGGACAAATATCTTTCTGAGGAGCAACAGTTATTACCAATCTTCTCTCAGCAA 60 mos.leu TACCTTGAGGACAAATATCTTTCTGAGGAGCAACAGTTATTACCAATCTTCTCTCAGCAA 60 mos.chr TACCTTGAGGACAAATATCTTTCTGAGGAGCAACAGTTATTACCAATCTTCTCTCAGCAA 60 mos.ber TACCTTGAGGACAAATATCTTTCTGAGGAGCAACAGTCATCACTAACCTTCTCTCAGCAA 60 mos.mos tra.jav TACCCTGAGGACAGATATCTTTCTGAGGAGCCACAGTCATCACCAACCTCTTATCAGCTA 60 TACCCTGAGGGCAAATATCTTTTTGAGGAGCTACAGTCATCACTAACCTTCTTTCAGCAA 60 trag.nap bala.acu TACCCTGAGGACAAATATCATTTTGAGGTGCAACCGTCATCACCAACCTCCTATCAGCAA 60 TACCCTGAGGACAAATATCATTTTGAGGCGCAACCGTCATCACCAACCTCCTATCAGCAA 60 bala.bon TACCCTGAGGACAATATCATTTTGAGGCGCAACCGTCATCACCAACCTCTTATCAGCAA 60 bala.bor TACCCTGAGGACAAATATCATTTTGAGGCGCAACCGTCATCACCAACCTCTTATCAGCAA 60 bala.edi TACCCTGAGGACAATATCATTCTGAGGCGCAACCGTTATCACCAACCTCCTATCAGCAA 60 bala.mus mega.nov bala.phy esch.rob TGCCCTGAGGACAAATATCATTCTGAGGCGCAACCGTCATCACCAACCTCCTATCAGCAA 60 TACCCTGAGGACAATATCATTCTGAGGCGCAACCGTCATCACCAACCTTCTATCAGCAA 60 TGCCCTGAGGACAAATATCATTCTGAGGCGCAACTGTAATCACTAACCTCCTATCAGCAA 60 TGCCCTGAGGACAGATATCATTCTGAGGCGCAACCGTCATCACCAACCTCCTATCAGCAA 60 cap.mar TACCCTGGGGACAGATATCATTTTGAGGTGCAACAGTCATCACCAACCTCCTATCAGCAA 60 ceph.com TACCCTGGGGACAGATATCATTTTGAGGTGCAACAGTCATCACCAACCTCCTATCAGCAA 60 TACCCTGAGGACAGATATCATTCTGAGGTGCAACAGTCATCACCAACCTCCTATCAGCAA 60 lage.obl TACCCTGAGGACAAATATCATTTTGAGGCGCAACAGTCATCACCAACCTCCTATCAGCAA 60 ceph.hea TACCCTGAGGACAAATATCATTTTGAGGTGCAACAGTCATCACCAACCTCCTATCAGCAA 60 ceph.hec TACCCTGAGGACAGATATCATTTTGAGGTGCAACAGTCATCACCAACCTCCTATCAGCAA 60 lage.aus TACCCTGAGGACAGATATCATTTTGAGGTGCAACAGTCATCACCAACCTCCTATCAGCAA 60 lage.cru TACCCTGAGGACAGATATCATTTTGAGGTGCAACAGTCATCACCAACCTCCTATCAGCAA 60 lage.obs TACCCTGAGGGCAGATATCATTTTGAGGTGCAACCGTCATCACCAACCTCCTATCAGCAA 60 lisso.bor TACCCTGAGGACAGATATCATTTTGAGGTGCAACCGTCATCACCAACCTCCTATCAGCAA 60 lisso.per TACCCTGAGGACAGATATCATTCTGAGGCGCAACCGTCATCACCAATCTCCTATCAGCAA 60 clo.mac TACCCTGAGGACAGATATCATTCTGAGGCGCAACCGTCATCACCAATCTCCTATCAGCAA 60 glo.mel TACCCTGAGGACAGATATCATTCTGAGGCGCAACCGTCATCACCAATCTCCTATCAGCAA 60 fere.att TACCCTGAGGACAGATATCATTCTGAGGCGCAACCGTCATCACCAATCTCCTATCAGCAA 60 pepo.ele TACCCTGAGGACAAATATCATTCTGAGGCGCAACCGTCATCACCAATCTCCTATCAGCAA 60 gram.gri TACCCTGAGGACAGATATCATTCTGAGGCGCAACCGTCATCACCAATCTTCTATCAGCAA 60 pse.cra TACCATGAGGACAAATATCATTCTGAGGCGCAACCGTTATCACCAATCTCCTATCAGCAA 60 lage.acu TACCCTGAGGACAGATATCTTTCTGAGGCGCAACCGTCATTACTAATCTCCTATCAGCAA 60 ordi.bre TACCCTGAGGACAGATATCCTTCTGAGGTGCAACCGTCATCACCAATCTCCTATCAGCAA 60 orca.bre TGCCCTGGGGACAAATATCATTCTGAGGGGGCAACCGTCATCACCAACCTCTTATCAGCAA 60 del.cap TGCCCTGAGGACAAATATCATTCTGAGGCGCAACCGTCATCACCAACCTCTTATCAGCAA 60 del.tro TGCCCTGAGGACAAATATCATTCTGAGGGGCAACCGTCATCACCAACCTCTTATCAGCAA 60 del.del TGCCCTGAGGACAAATATCATTCTGAGGCGCAACCGTCATCACCAACCTCCTATCAGCAA 60 sten.cly TGCCCTGAGGACAAATATCATTCTGAGGCGCAACCGTCATCACCAACCTCTTATCAGCAA 60 stem.coe TGCCCTGAGGACAAATATCATTCTGAGGCGCAACCGTCATCACCAACCTCTTATCAGCAA 60 tur.adu TGCCCTGAGGACAAATATCATTCTGAGGCGCAACCGTCATCACCAACCTCTTATCAGCAA 60 sten.fro TGCCCTGAGGACAAATATCATTCTGAGGGGCAACCGTTATCACCAACCTCCTATCAGCAA 60 saus.chi sten.lon TACCCTGAGGACAAATATCATTCTGAGGTGCAACCGTCATCACCAACCTCCTATCAGCAA 60 TGCCCTGAGGACAAATATCATTCTGAGGGGGAACCGTCATCACCAACCTCTTATCAGCAA 60 turs.tru lage.alb TACCCTGAGGACAAATATCATTGTGAGGGGGCAACCGTCATCACTAATCTGCTATCAGCAA 60 TACCCTGAGGACAAATATCATTCTGAGGTGCAACCGTCATTACCAACCTCCTGTCAGCAA 60 sten.bre TACCCTGAGGACAAATATCATTCTGAGGGGGAACCGTCATTACCAATCTCCTATCAGCAA 69 sota.flu

		63
del.leu	TACCCTGAGGACAAATATCATTCTGAGGCGCAACCGTCATTACCAATCTCCTATCAGCAA	. 0 .0
mono.mon	TACCCTGAGGACAAATATCATTCTGAGGTGCAACCGTCATCACCAACCTCCTATCAGCAA	
plat.gan	TACCCTGAGGACAAATATCATTCTGAGGTGCAACCGTCATCACCAACCTTTTATCAGCAA	60
plat.min	TACCCTGAGGACAAATATCATTCTGAGGTGCAACCGTCATCACCAACCTTTTATCAGCAA	60
kogi.bre	TACCCTGAGGCCAAATATCATTCTGAGGAGCAACCGTCATCACCAACCTTATATCCGCAA	63
kogi.sim	TGCCCTGAGGCCAAATATCATTCTGAGGAGCAACCGTCATCACAAACCTTATATCCGCAA	5.3
phys.cat	TGCCCTGAGGACAAATATCATTCTGAGCCGCAACCGTTATCACAAACCTTCTATCAGCAA	60
lipo.vex	TACCCTGAGGACAAATATCATTTTGAGGGGCAACCTTATCACTAATCTTTTTATCACTAACACTACTACT	63
phoc.sin	TGCCCTGGGGACAAATATCATTTTGAGGTGCTACCGTCATCACAAACCTCTTATCAGCAA	60
bera.bai	TGCCTTGAGGGCAAATATCATTCTGAGGTGCAACCGTCATCACCAACCTCTATCCGCTA	60
ziph.car	TACCTTGAGGACAAATATCATTCTGAGGTGCAACCGTCATCACAAACCTCTTATCCGCTA	60
meso.eur	TTCCCTGAGGACAAATATCATTCTGAGGCGCAACCGTTATTACCAACCTCCTATCCGCCA	60
meso.bid	TACCCTGAGGACAATATCATTCTGAGGGGCAACTGTTATTACTAACCTCCTATCCGCTA	60
meso.den	TACCATGAGGACAAATATCCTTCTGAGGTGCAACTGTCATTACCAATCTTCTATCCGCTA	60
hype.amp	TACCCTGAGGACAAATATCATTCTGAGGCGCAACCGTCATCACCAATCTCCTATCCGCCA	60
meso.per	TACCTTGAGGACAAATATCATTCTGAGGCGCAACTGTCATTACTAATCTTTTTATCTGCTA	60
pont.bla	TACCCTGAGGACAAATGTCATTCTGAGGTGCCACTGTCATCACTAACCTCCTATCAGCGA	63
hex.lib	TACCATGAGGACAAATATCATTCTGAGGGGCAACAGTCATCACCAACTTACTATCAGCTA	63
hipp.amp	TGCCATGAGGACAAATGTCATTCTGAGGGGCAACAGTCATTACCAACTTACTGTCAGCTA	60
dic.sum	TACCATGAGGTCAAATATCCTTCTGAGGAGCCACAGTTATCACAAATCTCCTCTCAGCCA	60
rhin.son	TACCATGAGGTCAAATATCCTTCTGAGGGGGCTACAGTCATTACAAATCTCCTCTCAGCCA	60
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sus.bar	TGCCCTGAGGACAATATCATTCTGAGGAGCTACGGTCATCACAAATCTACTATCAGCTA	60
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pho.gro	TACCATGAGGGCAAATGTCATTCTGAGGAGCAACAGTTATCACTAATCTACTATCAGCAA	50
pho.vit	TACCATGAGGACAAATATCATTTTGAGGAGCAACAGTCATCACCAATCTACTATCAGCAA	60
cys.cri	TACCGTGAGGACAAATATCATTTTGAGGAGCGACAGTCATCACCAACCTACTATCAGCAA	60
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red.aru	CAGATGTAGACAAAATGGGATTGGAGGCNTACTACACCATGAAAGAYATTGTAGGTGGGC	300
red.ful	CAGACGCAGACAAAATCCCATTCCACCCCTACTACACCATTAAAGACATTCTAGGCGCCA	300
neo.mos	CCGACATAGACAAAATTCCATTCCACCCATACTACACCATTAAAGATATTCTAGGCGCCT	300
pel.cap	CCGACATAGACAAAATTGCGTTCCACCCCTACTACACCATCAAAGACATTCTAGGAGCAC	300
gas/dam	CAGATGCAGACAAGGTCCCATTCCACCCCTACTACACCATTAAAGACATCCTAGGCGCCT	300
our.our	CAGACGCAGACAAAATTGCATTCCACCCCTACTACACTATCAAAGATATCCTAGGAGCTC	300
ant.cer	CAGACGCAGACAGAC ICCATICCACCCCTACTACACCATTAAAGACATTCTAGGCGCCC	300
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rap.mel	CAGATATAGACAAAATCCCATTTCACCCCTACTACACCATCAAGGACATTCTAGGAGCAC	300
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cer.ela.kan	CAGACGCAGACAAAATCCCCTTCCATCCTTACTATACCATTAAAGATATCTTAGGCATCT CAGACGCAGACAAAATCCCCTTCCATCCTTACTATACCATTAAAGATATCTTAGGCATCT	300
cer.ela.xan	CAGACGCAGACAAAATCCCCTTCCACCCTTACTATACGATTAAAGATATCTTAGGTATCT	300
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cer.nip.cent	CGGACGCAGACAAATCCCCTTCCATCCTTACTACACCATTAAAGATATCTTAGGCATCT CGGACGCAGACAAAATCCCCTTCCATCCTTACTACACCATTAAAGATATCTTAGGCATCT	300
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gaph.hag	GGAACATAGACATAATGGGATTCGACGGTTAGTAGACAAGAGAAGACATGGTAGGGGGTT GGAACATAGACATAATGGGATTCGACGGTTTAGTAGAGAAGAGAAGACATGGTAGGGGGTT	300
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gram.gri	CCAACATAGACATAATICCATICACCOTATTACACAATTAAAGATATCCTAGGGGGCTCCCCAACATAAAGATAATCCACACTTATTCCACACATTAAAGATATCCTAGGGGCTT	300
pse.cra	CCANCATAGACATAATTCCACLACCCAACACAATTAGATATCCTAGGCGCTT	300
lage.acu	CCARCALAGACA ANTICCATICACCCTTATTATACAATTAAAGATATCCTAGGCGCTT CTAACATAGATATAATCCCGTTCCACCCTTATTATACAATTAAAGATATCCTAGGCGCCC	300
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del.cap	CCAACATAGACATAATCCCATTCCACCCTTATTATACAATCAAAGATATCCTAGGTGCCT	300
del.tro	CCAACATAGACATAATCCCATTCCACCCTTATTATACAATCAAAGATATCCTAGGTGCCC	300
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lipo.vex	CTAACATAGACAAAATUUCAI ICCACCCTTACTATACAATCAAAGATATCCTAGGCGCCCCTTAACAATAAAGATAATCCTAGGAGCGCCCTTACTATACAATCAAAGATATCCTAGGAGCGCCCTTACTATACAATCAAAGATATCCTAGGAGCGCCCTTACTAACAATCAAAGATATCCTAGGAGCGCCCTTACTACTACAATCAAAGATATCCTAGGAGCGCCCTTACTAACAATCAAAGATATCCTAGGAGCGCCCTTACTACTACAATCAAAGATATCCTAGGAGCGCCCTTACTACTACAATCAAAGATATCCTAGGAGCGCCCTTACTACTACAATCAAAGATATCCTAGGAGCGCCCTTACTAACAATCAAAGATATCCTAGGAGCGCCCCTTACTACTACAATCAAAGATATCCTAGGAGCGCCCCTTACTACTACAATCAAAGATATCCTAGGAGCGCCCCTTACTACTACAATCAAAGATATCCTAGGAGCGCCCCTTACTACAATCAAAGATATCCTAGGAGCGCCCCTTACTACAATCAAAGATATCCTAGGAGCGCCCCTTACTACAATCAAAGATATCCTAGGAGCGCCCCTTACTACAATCAAAGATATCCTAGGAGCGCCCCTTACTACAATCAAAGATATCCTAGGAGCGCCCCTTACTACAATCAAAGATATCCTAGGAGCGCCCCTTACTACAATCAAAGATATCCTAAGGAGATATCCTAGGAGCGCCCCTTACTACAATCAAAGATATCCTAAGGAGCGCCCCTTACTACAATCAAAGATATCCAAAGATATCCAAAGATAATCAAAGATAATCAAAGATAATCAAAGATAATCAAAGATAATCAAAGATAATCAAAGATAATCAAAGATAACAATCAAAGATAATCAAAATCAAATCAAATCAAAGATAATCAAAAGATAATCAAAATCAAATCAAATCAAATAATAATAATAATAA	300
phoc.sin	CTAACATAGACATAATCCCT I CCACCC TACTATACAATCAAAGATATCCTAGGAGCCC CCAATATAGATAAAATTCCATTCC	300
bera.bai	CCAATATAGATAAATTCCATTCCACCCTTACTACACAATCAAAGACATCCTAGGAGCCC CCGATATAGACAAAATCCCATTCCACCCTTACTACACAATCAAAGACATCCTAGGGGCTC	300
ziph.car	CCGATATAGACAAAATCCCATTCCATCCTTACTACACAATCAAAGATATCCTAGGGGCTC CTGATATAGACAAAATCCCATTCCATCCTTACTACACAATCAAAGATATCCTAGGAGCCC	300
meso.eur	CTGATATAGACAAAATCCCATTCATCTTATACATCTTATAGATATCCTGGGAGCCC	300
meso.bid	DODACATATAGACAAATCCATTCCACCCTACTACACAATTAAAGATATCTTCAGACCCCCCCTACTACACAATTAAAGATATCTTTAGAGACCCCCCCTACTACACAATTAAAGATATCTTTAGAGACCCCCCTACTACAAATTAAAGATATTTTTTAGAACCCCCCCC	300
meso.den	DODDADDATTARATARATARATATATATATATATATATATAT	300
hype.amp	CTGACATAGATAAAATTUCATTTUAGGGTTACTACACAATCAAAGACACTGTAGGGGGGCC CTGACATAGACAAAATGCGGTTGGACGCATACTACAAATCAAAGACACTGTTAGGAGCGC CTGACATAGACAAAATTCCATTTCATGCTTACTATACAATTAAAGATATGTTAGGAGCGC	300
meso.per	CTGACATAGACAAAATTCCATTTCATCCTTACAAAAAAAA	300
pont.bla	CTOACATAGACAAAATTCCATTTCACCCTTACTACACAATTAAAGATATCCTAGGGGGCCC CTAACATAGATGCCATCCCATTTCACCCTTACTACACAATTAAAGATATCCTAGGGGGTAC	300
hex.lib	CTARCATROATOCCATCCCATTTACACCATTACACAATCAAAGATATCCTGGGCTACC CAAACGCAGACAAAATCCCATTCCACCCCTATTACACAATCAAGAACATCCTAGGTATCC CAAACGCAGACAAAATCCCATTCCACCCCTTATTACACAATCAAGGACATCCTAGGAGCCC	300
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sus.bar	CAGACATAGACAAAA 1000A. 1000A 1000A 1000AAAGACATTSTAGGAGGGT CAGACATAGACAAAATTGGATTTGAGGGATA STAGACTATGAAAGACATTGTAGGAGGGT CAGACATAGACAAAATTGGATTTGAGGGATA STAGACTATTAAGACATTGTAGGAGGAG	
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gru.nig	CALACTGCGATAAAATTCCATTCCACCCCTATTTTTCCTTAAAAGATACCCTAGGALICA 300
	CAAACTGTGATAAAATCCCATTCCACCCCTATTTTTCCTTAAAAGATATCTTAGGATTTA 300
gru.jap	CAAACTGCGACAAAATTCCATTCCACCCCTACTTCTCCCTCAAAGATATCCTAGGCCTTA 300
cic.boy	CTCACTCTGACAAAATCCCATTCCACCCCTACTTCTCCCTAAAAGATGCCCTAGGACTAG 300
rhe.ame	CCAACTGCGACAAAATCCCATTCCACCGATACTTTGCCCTAAAGGACATCCTAGGATTCA 300
ant.alb	CAAACTGCGATAAAATCCCATTCCATCCCTATTACTCTCTCAAAGACCTCCTAGGATTCA 300
fal.fam	CAAACTGCGACAAAATCCCATTCCATCCCTACTACTCTCTAAAAGACCTTTTTAGGAGTCA 300
fal.ver	CANACTGCGACAAAA CCCAA CCCAA CCCAACTACTACTACTA TAAAAAAAA
fal.per	CAAATTGCGACAAAATCCCATTCCACCCATACTACTCTCTCAAAGATATCCTAGGATTTA 300
fal.spa	CAAACTGTGACAAAATCCCATTCCACCCCTACTACTCTCTCAAAGACCTCCTAGGTTTTA 300
ayt.ame	CAGACTGCGACAAAATCCCATTTCACCCCTACTTCTCCTTCAAAGACATCCTAGGATTTA 100
smi.sha	CTAACTCCGATAAAATCCCATTCCACCCATACTTCTCCATAAAAGACATTCTAGGCTTTG 300
vid.cha	CAGACTGTGACAAAATTCCATTCCACCCATACTACACCACAAAGGACATCCTAGGCTTCG 300
chry.pic	CAAACACTGACAAAATCCCATTCCACCCTTATTTCTCATATAAAGACCTTTTTAGGCGTCA 300
emy.orb.kur	CAAACACCGATAAAATCCCTTTCCATCCCTACTTCTCATACAAGACCTATTAGGACTCA 300.
che.mud	CAAATACCGACAAAATCCCCTTCCACCCCTACTTCTCCTACAAAGACTTACTAGGACTCA 300
eum.egr	CTAGCACAGATAAGGTGCCATTCCACCCATATTACACATACAAAGACCTTCTTGGTTTCA 300
aep.mel	TATTAATAATTCTAGTCCTAATACTCCTAGTACTATTCATACCCGACCTACTAGGAGACC 360
ore.ore	TATTACTAATTCTAGCTTTATTACTCTTAGTATTATTCACACCTGACCTACTTGGAGACC 360
add.nas	TACTACTAATTCTAGTCCTCATACTACTAGTATTATTCACACCCGACCTACTTGGAGACC 360
ory.dam	TACTACTAATCCTAGCCCTTATGTTGCTAGTATTATTCGCACCCGACCTACTTGGAGACC 360
hip.equ	TACTACTAATTCTAGCCCTCATACTACTAGTACTATTCGCACCCGACCTACTTGGAGACC 360
alc.bus	TATTACTAATCCTAGCCCTCATACTACTAGTACTATTCGCACCCGACCTGCTCGGAGACC 360
sig.lic	TATTACTAATTCTAGCCCTCATACTACTAGTACTATTCGCACCCGACCTGCTCGGAGACC 360
bea.hun	TACTACTAATTCTAGCCCTCATATTACTAGTACTATTTGCACCCGACCTGCTCGGAGACC 360
dam.lun	TACTACTAATTCTAGCCCTCATACTACTAGTACTATTTGCACCCGACCTGCTCGGAGACC 360
con.tau	TATTACTAATTCTAGCCCTAATACTACTAGTACTATTCGCGCCCGATTTACTTGGAGACC 360
amm.ler	TGCTACTAATCCTCACCCTCACACTACTAGTACTATTTACACCCGATCTACTCGGGGACC 360
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cap.ibe	TGCTACTAATTCTTGTCCTAATATTACTAGTACTATTCACACCCGACCTACTCGGGGACC 360
hem.jem	TACTACTAATTCTTGTCCTAATATTACTAGTACTATTTATACCCGACCTACTTGGAGACC 360
cap.fal	TACTACTACTCGCCCTGATGCTACTAGTACTATTCACACCTGACCTACTCGGAGACC 360
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ovi.amm	TCCTACTAATCCTCATCCTCATGCTGCTAGTACTATTCACGCCTGACTTACTT
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cap.cri	TACTACTAATCCTTACCCTTATACTACTACTATTCACACCCGACCTACTTGGAGACC 360
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pel.cap		350
gaz.dam	TACTATTAATTCTAGCCCTCATACTCCTAGTTCTATTCACACCAGACCTGCTTGGAGACC TCCTACTAATTCTAGCCCTCATGCTCCTAGTCCTATTCACACCAGACCTGCTTGGAGACC	350
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sai.tat		
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cer.ela.xan	TACTTCTAGTACTCTTAGTATTACTAGTATTATTCGCACCAGATCTGCTTGGAGACC TACTTCTAATACTCTTCCTAATATTACTAGTATTATTCGCACCAGATCTGCTTGGAGATC	360
cer.ela.can	TACTTCTAATACTCTTAATATTACTAGTATTATTCGCACCAGACCTGCTTGGAGATC TACTTCTAGTACTCTTAATATTACTAGTATTATTCGCACCAGACCTGCTTGGAGATC	350
cer.nip.cent	TACTTCTAGTACTCTTCCTAATATTACTAGTATTATTCCCACACACA	360
cer.mip.yes	TACTTCTAGTACTCTTCCTAATATTACTAGTATTATTCGCACCAGACCTGCTTGGAGATC TACTTCTAGTACTCTTCCTAATATTACTAGTATTATTCGCACCAGACCTGCTTGGAGATC	350
cer.nip.ker	TACTTCTAGTACTCTTAGTATTACTAGTATTATTCGCACCAGAGCTGCTTGGAGATC TACTTCTAGTACTCTTGGTATATTACTAGTATTATTCGCACCAGAGCTGCTTGGAGATC	360
cer.nip.pul		350
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esch.rcb		350
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mcs.fus	CGGACAATTATACCCCAGCAAACCCATTAAATACGCCCCCACATATTAAACCCGAATGAT	420
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tra.cob	ACTICCTATIOGCOTTATGCCATCCTACGCTCAATCCCAAACAAACTCGGAGG	47
tra.tem	ATTTTCTGTTCGCTTATGCCATCCTGCGCTCAATTCCAAACAAA	470
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cat.wal	ACTICCTATIOGCOTACGCTATCCTACGCTCAATCCCAAATAAACTCGGAGG	472
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Reference:

Altschul, Stephen F., Thomas L. Madden, Alejandro A. Schäffer, Jinghui Zhang, Zheng Zhang, Webb Miller, and David J. Lipman (1997), "Gapped BLAST and PSI-BLAST: a new generation of protein database search programs", Nucleic Acids Res. 25:3389-3402.

RID: 984593689-1224-27770

Query=

(328 letters)

Database: Sequences from complete mitochondrial genomes 129 sequences; 3,164,247 total letters

If you have any problems or questions with the results of this search please refer to the BLAST FAOS

Taxonomy reports

Distribution of 80 Blast Hits on the Query Sequence

Mouse-over to show defline and scores. Click to show alignments	ᆜ
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Sequences produci	ing significant alignments:	(bi	(2 5)	Value
	Felia catus mitochondrion, complete genome	365	e-10	
ref NC 001700 1	Phoca vitulina mitochondrion, complete genome	198	1e-5	
ref[NC_001325.1]	Canis familiaris mitochondrion, complete g	190	4e-4	19
ref NC 002008.1	Halichoerus grypus mitochondrion, complete	180	3e-4	
ref NC 001602.1	Cavia porcellus complete mitochondrial genome	175	5e-4	15
ref[NC_000884.1]	Ceracotherium simum mitochondrion, complet	155	2e-4	11
ref NC 001808.1	Myoxus glis mitochondrion, complete genome	153	8e-3	8
ref NC 001892.1	Equus asinus mitochondrion, complete genome	1,5,1	3e-3	37
ref NC_001788.1	Orycteropus afer complete mitochondrial ge	149	1e-3	16
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ref NC 001821.1	R.unicornis complete mitochondrial genome	135	2e-3	12
ref NC 001779.1	Mus musculus mitochondrian, complete genome	133	7e-3	12
ref NC 001569.11	Hippoporamus amphibius mirochondrion, comp	125	2e-2	29
ref NC 000889.1	Equus caballus mitochondrion, complete genome	125	2e-7	29
ref NC 001640.1	Macropus robustus mitochondrion, complete	123	7e-2	29
== E NC 001794 . 1	Sus scrofa mitochondrion, complete genome	121	3e-2	28
ref NC 000845.1	Rattus norvegicus mitochondrial genome	121	3e-2	28
ref NC 001665.1	Bos taurus mitochondrion, complete genome	121	3e-2	28
ref NC 001567.1	Pan troglodytes mitochondrion, complete ge	117	4e-2	27
res NC 001643.1	Ovis aries mitochondrion, complete genome	109	1e-2	24
ref NC 001941.1	Talpa europaea mitochondrion, complete genome	103	7e-2	23
ref NC 002391.1	Orycrolagus cuniculus mitochondrion, compl	103	7e-2	23
ref NC 001913.1	Pan paniscus mitochondrion, complete genome	101	3e-2	22
ref NC 001644.1	Human mitochondrion, complete genome	99	4e-2	21
ref NC 001807.2	Balaenoptera musculus mitochondrion, compl	98	4e-2	21
ref NC 001601.1	Artibeus jamaicensis mitochondrion, comple	95	2e-2	
ref NC 002009.1	Gorilla gorilla mitochondrion, complete ge	<u>92</u>	2e-1	19
ref NC 001645.1	Balaenoptera physalus mitochondrion, compl	90	1e-1	18
ref NC 001321.1	Didelphis virginiana mitochondrion, comple	_80	9e-1	
ref NC 001610.1	Hylobates lar mitochondrion, complete genome	_70	9e-1	
ref NC 001727.1	Crosseroma lacustre mitochondrion, comple	70	9e-1	
ref NC 001804.1	rarimania chalumnae mitochondrion, complet	<u> 68</u>	4e-1	
ref NC 000880.1	vidua chalvheata mitochondrion, complete 9	<u> 56</u>	1e-1	
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ref NC 000886.11	Chalonia mydas mitochondrial DNA, complete	<u> 62</u>	2e-1	
ref NC 001646.1	Bonco numaeus mitochondrion, complete genome	<u> 62</u>	2e-1	
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NC 001645 14955 .gcac	14974
NC 001610 14963cgca	14990
NC 002082 14953	14972
NC 001960 16170 Ct t. a C	16197
NC 001953 14416c	14437
NC 001733	
Database: Sequences from complete mito	chondrial genomes
pogred date: Jun 28, 2000 10:50 AM	
Number of letters in database: 3,164,2	47
Number of sequences in database: 129	
Lambda K H	
1.37 0.711 1.31	
Gapped .	
Lambda K H	
1.37 0.711 1.31	
Matrix: blastn matrix:1 -3	· ·
Gap Penalties: Existence: S, Extension: : Number of Hits to DB: 788	_
Number of Alex to DB: 700 Number of Sequences: 129	
Number of extensions: 788	•
Number of successful extensions: 168	
Number of sequences better than 10.0: 77	
length of query: 328	
length of database: 3,164,247	
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effective length of query: 313	•
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effective search space used: 989803656	
T: 0	
A: 30	
X1: 6 (11.9 bits)	
XZ: 15 (29.7 bits)	•
S1: 12 (24.3 bits) S2: 14 (28.2 bits)	
24: 14 /40.4 11/21	

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BLASTN 2.1.2 [Nov-13-2000]

Reference:

Altschul, Stephen F., Thomas L. Madden, Alejandro A. Schäffer, Jinghui Zhang, Zheng Zhang, Webb Miller, and David J. Lipman (1997), "Gapped BLAST and PSI-BLAST: a new generation of protein database search programs". Nucleic Acids Res. 25:3389-3402.

RID: 984593400-28182-3122

Query-

(328 letters)

Database: nt

807,597 sequences; 2,863,827,885 total letters

If you have any problems or questions with the results of this search please refer to the BLAST FAQS

Taxonomy reports

Distribution of 50 Blast Hits on the Query Sequence

Mouse-over to show defline and scores. Click to show alignments	==
×	
	:

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Score
                                                                                                                 (bics) Value
 Sequences producing significant alignments:
                                                                                                                603 e-170
 gb[AY005809.1] Panchera pardus cytochrome b gene, partial c...
gb|AY005809.1| Panthera pardus cytochrome b gene, partial c...
gb|AF053054.1|AF053053
gb|AF053053.1|AF053053
gb|AF053049.1|AF053049
gb|AF053025.1|AF053025
gb|AF053024.1|AF053024
gb|AF053023.1|AF053024
gb|AF053023.1|AF053023
gb|AF053022.1|AF053022
gb|AF053021.1|AF053021
gb|AF053021.1|AF053021
gb|AF053021.1|AF053021
gb|AF053021.1|AF053021
gb|AF053018.1|AF053018
                                                                                                               527 e-147
527 e-147
476 e-132
                                                                                                               475 e-132
                                                                                                               450 e-127
                                                                                                                450 e-127
                                                                                                                450 e-127
                                                                                                                450 e-127
                                                                                                                460 e-127
Gb[AF05]018.1[AF05]018 Panthera tigris tigris isolate 82 cy...
                                                                                                               450 e-127
 gb|AF053051.1|AF053051 Panthera tigris corbetti isolate C3 ...
                                                                                                               452 e-125
                                                                                                              452 e-125
 gb]AF053048.1|AF053048 Panchera tigris sumacrae isolate Sul...
 gb/AF053047.1/AF053047 Panthera tigris sumacrae isolate Su9...
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        Panthera tigris sumatrae isolate Su7 | 452 e-125

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        Panthera tigris sumatrae isolate Su6 | 452 e-125

        GD | AF053044 | 1 | AF053044
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        Panthera tigris altaica isolate S15 | 452 e-125

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        Panthera tigris altaica isolate S14 | A52 e-125

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452 e-125
452 e-125
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452 e-125
452 e-125
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 gb|AF053035.1|AF053035 Panthera tigris altaica isolate S11 ...
gb|AF053034.1|AF053034

gb|AF053033.1|AF053032

gb|AF053032.1|AF053032

gb|AF053031.1|AF053031

panthera tigris altaica isolate S6 c...

gb|AF053031.1|AF053031

Panthera tigris altaica isolate S7 c...

panthera tigris altaica isolate S7 c...
                                                                                                                <u>452</u> e-125
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 SD|AF053029.1|AF053029 Panthera tigris altaica isolate S4 C...
cb|AF053028.1|AF053028 panthera tigris altaica isolate 51 c...
cb|AF053027.1|AF053027 panthera tigris altaica isolate 52 c...
cb|AF053026.1|AF053026 panthera tigris altaica isolate 52 c...
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cb|AF053020.1|AF053020 panthera tigris tigris isolate 84 cy...
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                                                                                                               452 e-125
444 e-122
444 e-122
440 e-121
gb/AF053019.1/AF053019 Panthera tigris tigris isolate B3 cy...
gb/AF053043.1/AF053041 Panthera tigris sumatrae isolate Su4...
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GD/AF053052.1/AF053052 Panthera leo cytochrome b (cytb) gen...
emb/X82300.1/MIPLCYTBG P.leo mitochondrial cytochrome b gene
                                                                                                                438 e-121
                                                                                                               399 e-106
 dbi|AB004238.1|AB004238 Felis catus mitochondrial DNA for c...
                                                                                                               381 e-103
177 e-102
165 le-98
165 le-98
 dbi[A3004237.1[A3004237] Felis catus mitochondrial DNA for G...
 emb|X82296.1|MIFDCYTB F.domesticus mitochondrial cytochrome...
ref[NC 001700.1] Felis catus mitochondrion, complete genome gb[U20753.1]FCU20753 Felis catus mitochondrion, complete ge...
QD: UZU/D3: LIFCUZU/D3Felia catus mitochondrion, complete ge...1651e-98QD: AF125145. LIAF125145Viverricula indica cytochrome b gene...2767e-72QD: AF125144. LIAF125144Chrotogale owstoni cytochrome b gene...2704e-70QD: AF154975. LIAF154975Martes martes specimen voucher AF175...2557e-66QD: AB051237. LIAB051237Martes martes mitochondrial cyto ge...2367e-56QD: AF125149. LIAF125149Viverra tangalunga cytochrome b gene...2466e-63
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                            98
 cmpseq_0 I
                            545
 AY305809 19
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AF053050
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AFOSJOIS
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 AFQ53Q45
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 AF251947
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AF053040	487		546
AF053039	487	gt	546
AF053038	487	# P	
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AF053037		gt	546
AF053036	487	gg	546
AF053035	487	gc	546
AF053034	487	gg	370
	487	m m	240
AF053033	-	gE	546
AF053032	487	gcgc	546
AF053031	487		546
AF053030	487	gc	E 1 E
AF053029	487	gt	240
	487	— •	340
AF053028		7 P 	340
AF053027	487		240
<u> AF053026</u>	487		546
AF053020	487	gt	546
AF053019	487		
AF053043	487		
X82301	487		_
AF053052	487		
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A3004237	487		
X82296	487		
NC 001700	-		
<u>U20753</u>	15524		
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<u> AF125144</u>	357	tgggcag	546
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	487		
A3051237	487		
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A3051237 AF125149 Cmpseq_0 AY005809 AF053054	487 357 61 99 547	ttcatccttccatttatcatctcagctctagcagcagtccacctcctattccttcacgag	415 120 153 606 606
A3051237 AF125149 Cmpseq_0 AY005809	487 357 61 99	ttcatccttccatttatcatctcagctctagcagcagtccacctcctattccttcacgag	415 120 153 606 606
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AB051237 AF125149 EMPSEQ 0 AY005809 AF053054 AF053050 AF053049 AF053025 AF053024 AF053024 AF053021 AF053021 AF053021 AF053045 AF053045 AF053045 AF053045 AF053044 AF053045 AF053045 AF053045 AF053045 AF053045 AF053041	435 61954477777777777777777777777777777777777	g	415 123666666666666666666666666666666666666

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AF053020	547		
AF053019	547		
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AF053043	547		
X82301	54.7		
<u> AF053052</u>	547	gggggg	606
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X82296	547	t	15641
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U20753	15584		230.0
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AF125144	417		4 / 5
AF154975	547		000
AB051237	547		800
AF125149	417	t.att.a.	4/6
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AY005809	607		666
AF053054			666
AF053053	607 607	- h	666
AF053050	607 607	# # # # # # # # # # # # # # # # # # #	666
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AF053039	607		666
AF053038	607 607		666
AF053037	607		666
AF053036	607	gggg	666
AF053035	607		600
<u>AF053034</u> AF053033	607	gggg	600
AF053033	607		
AF053031	607		
AF053031	607		
AF053029	607		
AF053028	607	gggg	666
AF053027	607		
AF053026	607		
AF053020	607		666
AF053019	607		
AF053043	607		
X82301	607		666
AF053052	607		666
X32300	607		666
AB004238	607	tactac	666
AB004237	607	tact.	555
X82295	607	tactac	12107
	15644		614 72107
<u>U2075]</u> AF125145	477		3.00 4.16
AF123144	477		666
AF154275	607		
A8951217	607	- management P	
1513112	477	g.,c.,c.,	
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AF053025	-		
AF053020	667		725
<u>AF053019</u>	667		725
AF053043	667		725
X32301	667		725
AF053052	667		726
X82300	667		725
A9004238	667		726
A3004237	667		726
X82296	667		15763
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NC 001700			15763
<u>U20753</u>	1570-	t	596
AF125145	537		
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AF154975	667	cagccc.c	
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AF125149	537	ttt.atct.ct.agc.agc.a.	
cmpseq_0	241	geocattocoaccagacctgttaggagaccccgataactacatccctgccaaccctcta	338
AY005809	279		785
AF053054	727		785
AF053053	727		785
AF051050	727		786
AF053049	727		
AF051025	727		785
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AF051024	727		785
AF053073	727		786
AF051022	727		785
AF051021	727		785
AF051018	727		785
AFOS 1051	727		786
AF751049	727		785
AF951947	727		785
AF751945	727		785
AF751745	727		75%
AF951944	727		786
AFDSJD42	727		796
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AF053038	727	* · · · · · · · · · · · · · · · · · · ·	786
AF053037	727	P	786
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AF053036	727		786
AF053035	727		786
AF053034	727		786
AF053033	727	A	786
AF053032	727		
AF053031	727	a	786
	727	tcccc	785
AF053030	_		786
<u> AF053029</u>	727		786
AF053028	727		786
AF053027	727		786
AF053026	727		
AF053020	727	A	785
AF051019	727	a	786
AF053043	727		786
			786
X82301	727	t.ct.c	786
<u>AF053052</u>	727	t.ct.ct.	786
X82300	727	aa.	786
A3004238	727	*	786
AB004237	727		786
X82296	727		
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AF125145	597		656
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AF154975	727	acccgacc	786
A3051237	727		6 56
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	747	.gca	812
AF053052	787	.gc	814
X82100	787	.gcaa	
AB004238	787		874
	787		814
AB004237			814
X82296	787		10001
NC 001700	15824	ta	13031
	15824		15851
<u>U20753</u>			664
AF125144	657	.,C	803
AF154975	787	⊂aa	
	787	caa	803
AB051237			664
AF125149	657	c	

Database: nt

Posted date: Mar 2, 2001 12:20 AM

Number of letters in database: 2,861,827,885 Number of sequences in database: 807,597

Lambda K H 1.37 0.711 1.31

Gapped

Lambda K H

S1: 12 (24.3 bits) S2: 19 (38.2 bits)

1.37 0.711 1.31

Matrix: blastn matrix:1 -3 Gap Penalties: Existence: 5, Extension: 2 Number of Hits to DB: 460542 Number of Sequences: 807597 Number of extensions: 460542 Number of successful extensions: 22671 Number of sequences better than 10.0: 6487 length of query: 328 length of database: 2,863,827,885 effective HSP length: 20 effective length of query: 308 effective length of database: 2,847,675,945 effective search space: 877084191060 effective search space used: 877084191060 T: 0 A: 30 X1: 6 (11.9 bits) X2: 15 (29.7 bits)

Table 5. Reference animals and the allocated code numbers included in the study

SN	I. Code number	Name of the animal	Zoological name
1	bhz25t	Indian tiger	Panthera tigris tigris
2	bhz26t	Indian tiger	Panthera tigris tigris
3	bhz30t	Indian tiger	Panthera tigris tigris
4	bhz45t	Indian tiger	Panthera tigris tigris
5	bhz56t	Indian tiger	Panthera tigris tigris
6	bhz63t	Indian tiger	Panthera tigris tigris
7	bhz20wt	Indian white tiger	Panthera tigris bengalensis
8	bhz22wt	Indian white tiger	Panthera tigris bengalensis
9	bhz23wt	Indian white tiger	Panthera tigris bengalensis
10	bhz28wt	Indian white tiger	Panthera tigris bengalensis
11	gz1l	Normal leopard	Panthera pardus
12	gz2l	Normal leopard	Panthera pardus
13	gz3l	Normal leopard	Panthera pardus
14	gz21cl	Clouded leopard	Neofelis nebulosa
15	gz22cl	Clouded leopard	Neofelis nebulosa
16	darz14sl	Snow leopard	Panthera unicia
17	darz15si	Snow leopard	Panthera unicia
18	darz16sl	Snow leopard	Panthera unicia
19	sbz22al	Asiatic lion	Panthera leo persica
20	sbz38al	Asiatic lion	Panthera leo persica
21	sbz39al	Asiatic lion	Panthera leo persica
22	humsk	Human	Homo sapiens sapiens
23	chimss	Chimpanzee	Pan sp.

Table 6. Multiple sequence alignments of the cytochrome b sequences of reference animals with the sequence obtained from confiscated animal remain

	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	o
sbz22al	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	0
sb::38al	TGAXTCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6 TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	o
sbz39al	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCTACCCTGACACGATTCTTTGCCTTCCAC 6 TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCTACCCTGACACGATTCTTTGCCTTCCAC 6	o
adil.flesh	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCTACCTTGACACGATTCTTTGCCTTCCAC 6 TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCTACCTTGACACGATTCTTTGCCTTCCAC 6	0
gzini	TGAATCTGAGGAGGCTICTCAGTAGACAAAGCTACCTTGACACGATTCTTTGCCTTCCAC 6 TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCTACCTTGACACGATTCTTTGCCTTCCAC 6	0
gz2nl	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCTACCTTGACACGATTCTTTGCCTTCCAC 6 TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCTACCTTGACACGATTCTTTTGCCTTCCAC	0
gz3nl	TGAATCTGAGGAGGCTTCTCAGIAGACAAAGCIACCIIGACACGAIIGTTGCCTTCCAC	0
bh=23wt	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	a
bh=28wt	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	a
bh=22wt	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6 TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	0
bh=20wt	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	a
bhz63t	The state of the s	_
bhzset	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6 TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTTGCCTTCCAC 6	i a
bhz26t	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	a
bhz30t	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	3
bhz45c	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	0
bhz25t	TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6	a
dz14sl	TGAXICIGAGAGGCTTCTCAGTACACAAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6 TGAATCTGAGGAGGCTTCTCAGTACACAAAAGCCACCCTGACACGATTCTTTTGCCTTCCAC 6	a
d215sl	TGAATCTGAGGAGGCTTCTCAGTACACAAAGCCACCCTGACACGATTCTTTGCCTTCCAC 6 TGAATCTGAGGAGGCTTCTCAGTACACAAAAGCCACCCTGACACGATTCTTTTGCCTTCCAC 6	0
d:16sl		-
gz21cl	men a memera cen ceremperor l'estaga calabecca celle GALALGAL i i i l'occi i l'occi i l'occi i l'occi i l'occi	-
gz22cl		-
chimss	TGAATCTGAGGAGGCTACTCAGTAGACAGCCCTACCCTTACACGATTCTTCACCTTCAC 6 TGAATCTGAGGAGGCTACTCAGTAGACAGCCCTACCCTTACACGATTCTTCACCTTCAC 6	a
humsk	TGAATCTGAGGAGGCTACTCAGTAGACAGTCCCCACACCCTCACACGATTCTTACACGACTCACACGATTCTTACACGACTCACACACA	•
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sbz22al	TTCATCCTTCCATTTATCATCTCAGCCCTAGCAGCAGTCCACCTCCTGGTTCCTCCATGAA 1 TTCATCCTTCCATTTATCATCTCAGCCCTAGCAGCAGTCCACCTCCTGGTTCCTCCATGAA 1	.20
sbz38al	TTCATCCTTCCATTTATCATCTCAGCCCTAGCAGCAGCAGTCCACCTCCTGTTCCTCCATGAA 1 TTCATCCTTCCATTTATCATCTCAGCCCTAGCAGCAGCAGTCCACCTCCTTGTTCCTTCACGAG 1	20
sbz39al	TTCATCCTTCCATTTATCATCTCAGCCCTAGCAGCAGCAGCTCCACCTCCTATTCCTTCACGAG 1 TTCATCCTTCCATTTATCATCTCAGCTCTAGCAGCAGCTCTACCTCCTTATTCCTTCACGAG 1	20
adil.flesh	TTCATCCTTCCATTTATCATCTCAGCTCTAGCAGCAGCAGCACCTCCTATTCCTTCACGAG 1 TTCATCCTTCCATTTATCATCTCAGCTCTAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCAGCA	20
gzinl	TTCATCCTTCCATTTATCATCTCAGCTCTAGCAGCAGCAGTCCACCTCCTATTCCTTCACGAG 1 TTCATCCTTCCATTTATCATCTCAGCTCTAGCAGCAGCAGTCCACCTCCTATTCCTTCACGAG 1	20
gz2nl	TTCATCCTTCCATTTATCATCTCAGCTCTAGCAGCAGCTCCACCTTCTATTCCTTCACGAG 1 TTCATCCTTCCATTTATCATCTCAGCTCTAGCAGCAGCTCCACCTTCTTATTCCTTCATGAG 1	20
gz3ml	TTCATCCTTCCATTTALCATCTCAGCCCTAGCAGCAGTCCACCTCCTATTCCTCCATGAG 1 TTCATCCTTCCATTTATCATCTCAGCCCTAGCAGCAGTCCACCTCCTATTCCTCCATGAG 1	20
bhz23wt		20
bhz28wt	TTCATCCTTCCATTTATCATCTCAGCCCTAGCAGCAGCAGCACTCCTCCTATTCCTCCATGAG 1 TTCATCCTTCCATTTATCATCTCAGCCCTAGCAGCAGCAGCACTCCACCTCCTATTCCTCCATGAG 1	20
bhz22wt		20
bhz20wt		20
bhz63t		20
bhz56t		20
bhz26t bhz30t		
bhz45t		
bh=25t		
d=14sl		
d=15sl		
dzlésl		
gz21cl		
gz22cl		
chimss	TTCATCCTCCCATTTATCACCACCCTAACACACTCATCCTCATTCTTACACGAA 1 TTTATCTTACCCTTCATTATCACACCCCTAACACACTCATCCTCATTCTTACACGAA 1	20
humsk	TTTATCTTACCCTTCATTATTACAGCCCTAGCAGCACTCCACCTCCTATTCTTGCACGAA L	
	** *** * ** ** ** ** ** ** ** ** ** **	
	ACAGGATCTAATAACCCCTTCAGGAATGGTATCTGACTCAGGATAAAATTCCATTCCATCCA	90
o5:22al	ACAGGATCTAATAACCCCTCAGGAATGGTATCTGACTCAGATAAAATTGCATTGCATCCA :	90
ab=18al	ACAGGATC PARTANCES T. SHOWN CONTROL SHOW	

sbz39al	ACAGGATCTAATAACCCCTCAGGAATGGTATCTGACTCAGATAAAATTCCATTCCA
adil.flesh	A CAGGATOTA A CA A COCOTTO A GGAATAGTATO CGACTOAGACAAAAATTO CATTO CACCOA 100
	A CAGGATOTA A CALACCOTTCAGGAATAGTATCCGACTCAGACAAAATTCCATTCC
gzinl	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATTCCATTCCACCCA 190
g=2n1	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATTCCATTCCACCCA 133
gz3nl	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 130
bhz23wc	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTTCCACCCA 130
bhz18wt	ACAGGATCTÁACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 130
bn=22wc	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGGACAAAATCCCGTTCCACCCA 130
bhz20wc	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 180
bhz63t	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 190
bhz56t	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 180
bhz26t	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGGACAAAATCCCGTTCCACCCA 130
bhz30t	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 130
bhz45t	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 180
bh:25t	ACAGGATOTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 180
dz14sl	ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 180 ACAGGATCTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 180
dz15sl	ACAGGATOTAACAACCCCTCAGGAATAGTATCTGACTCAGACAAAATCCCGTTCCACCCA 180
dz16sl	ACAGGATCIAACACCCTCAGGAATGGTATCCGATTCAGACAAAATCCCGTTCCACCCG 130 ACAGGATCCAATAACCCCTCAGGAATGGTATCCGATTCAGACAAAATCCCGTTCCACCCG 130
gz21cl	ACAGGATCCAATAACCCCTCAGGAATGGTATCCGATTCAGACAAAATCCCGTTCCACCCG 180 ACAGGATCCAATAACCCCTCAGGAATGGTATCCGATTCAGACAAAATCCCGTTCCACCCG 180
gz22cl	ACAGGATCAAATAACCCCCTGGGAATCACCTCCCACTCCGACAAAATTACCTTCCACCCC 180
chimss	ACAGGATCAAACAACCCCCTAGGAATCACCTCCCATTCCGATAAAATCATCTTCCACCCT 190
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bh=20wt	TACTACACATCAAAGACATCCTGGGCCTTCTAGTACTAATCCTAACACTCATACTACTC 240
bhz635	TACTACACAATCAAAGACATCCTGGGCCTTCTAGTACTAATCCTAACACTCATACTACTC 240
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bhz25t	TACTACACAATCAAAGACATCCTGGGCCTTCTAGTACTAATCCTAACACTCATACTACTC 240 TACTACACAATCAAAGACATCCTGGGCCTTCTAGTACTAATCCTAACACTCATACTACTC 240
bh=30=	TACTACACAATCAAAGACATCCTGGGCCTTCTAGTACTAATCCTAACACTCATACTACTC 240 TACTACACAATCAAAGACATCCTGGGCCTTCTAGTACTAATCCTAACACTCATACTACTC 740
bhz45c	TACTACACAATCAAAGACATCCTGGGCCTTCTAGTACTAATCCTAACACTCATACTACTC 240 TACTACACAATCAAAGACATCCTGGGCCTTCTAGTACTAATACTAACACTCATACTACTC 240
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bhz28wt	GTCCTATTCTCACCAGACCTATTAGGGGACCCCGATAACTACATCCCCGCCAACCCTCTA	300
bh=22wt	GTCCTATTCTCACCAGACCTATTAGGGGACCCCGATAACTACATCCCCGCCAACCCTCTA	300
bhz20wt	GTCCTATTCTCACCAGACCTATTAGGGGACCCCGATAACTACATCCCCGGCCAACCCTCTA	300
bhz63t	GTCCTATTCTCACCAGACCTATTAGGGGACCCCGATAACTACATCCCCGGCAACCCTCTA	300
	GTCCTATTCTCACCAGACCTATTAGGGGACCCCGATAACTACATCCCCGCCAACCCTCTA	300
bhz56t	GTCCTATTCTCACCAGACCTATTAGGGGACCCCGATAACTACATCCCCGCCAACCCTCTA	300
bhz26t	GTCCTATTCTCACCAGACCTATTAGGGGACCCCGATAACTACATCCCCGCCAACCCTCTA	300
bhz30t	GTCCTATTCTCACCAGACCTALIAGGGGACCCGATAACTACATCCCGCCATACCTATACATCCCGCCATACCTATACATCCCGCCATACCTATACATCCCGCCATACCTACATCCGCCATACCTATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCGCCATACCTACATCCATACATCCCCCATACATCCATACATCCCCATACATCCATACATCCATACATCCATACATCCATACATCCATACATCCATACATCCATACATCAT	300
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bhz25t	GTCCTATTCTCACCAGACCTATTAGGGGACCCCGATAACTACATCCCCGGCCAACCCTCTA	300
dz14sl	GTCCTATTCTCACCAGACCTATTAGGGGACGCCGATAACTACATCCCCGCCAACCCTCTA	300
dz15sl	GTCCTATTCTCACCAGACCTATTAGGGGACGCCGATAACTACATCCCCGCCAACCCTCTA	300
d:16sl	GTCCTATTCTCACCAGACCTATTAGGGGACGCCGATAACTACATCCCCGCCAACCCTCTA	300
g:21cl	GTTCTATTCTCCCCAGACCTACTAGGAGACCCTGACAATTACACTCCCGCCAACCCTCTA	300
_	GTTCTATTCTCCCCAGACCTACTAGGAGACCCTGACAATTACACTCCCGCCAACCCTCTA	300
gz22cl	ACACTATTCTCACCAGACCTCCTGGGCGATCCAGACAACTATACCCTAGCTAACCCCCTA	300
chimss	ACACIAI ICI CACAGACCI COGGAGACAGA CACAGA A TATA ACCETAGA CACACITA	300
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AATACCCCTCCCCATATCAAGCCTGAAT 328 AATACCCTTCCCCATATCAAGCCTGAAT 328
AATACCCTTCCCCATATCAAGCCTGAAT 328
AACACCCCTCCCCATATCAAGCGCGAAT 328
AACACCCCTCCCCATATCAAGCCGGAAT 328
AACACCCCTCCCCATATCAAGCCCGAAT 328
AATACCCCTCCCCATATCAAGCCCGAAT 328
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Table 7a

bhz22wt bhz23wt dz14sl [基] dz14sl [基] [] dz16sl [基] [] sbz22al sbz38al gz21cl bhz26t bhz30t bhz45t bhz56t bhz20wt bhz25t adil.flesh Position C 11 က ဩ ල G 150 :> 163 ġ **G** 156 169 ଦ ଦ **水** C 1. A CHICACHAINA

Table 7b

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Table 7c

Table 7d

Position	273	276	279	282	284	285	287	288	291	294	297	298	302	303	309	315	348	324	323	324
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Table 8. Percent similarity matrix calculated by pair-vise comparisons of cytochrome b gene sequences revealed from 'adil.flesh' and different felids

	bhz20wt	bhz25t	dz14si	humsk	chlmss	sbz22al	021L	gzSL	0z3L	gz21cl	adil.flesh
bhz20wt		100	99.1	81.7	78.7	93.3	95.1	95.4	95.4	9.68	95.4
bh225t	100		99.1	81.7	7.87	93.3	95.1	95.4	95.4	89.6	95.4
dz14si	99.1	99.1		81.4	78.4	93	94.8	95.1	95.1	89.3	95.1
humsk	81.7	81.7	81.4		86.9	79.6	81.1	80.2	80.2	79	81.4
chimss	78.7	7.8.7	78.4	86.9		78.7	79.6	7.8.7	78.7	76.8	79.9
sbz22ai	93.3	93.3	. 93	79.6	78.7		92.1	92.4	92.4	89	92.4
gz1L	95.1	95.1	94.8	81.1	79.6	92.1		98.5	98.5	89.3	2.66
gz2L	95.4	95.4	95.1	80.2	78.7	92.4	98.5	選 選	100	88.1	98.2
graf	95.4	95.4	95.1	80.2	7.87	92.4	98.5	100		88.1	98.2
9221cl	89.6	9.68	89.3	79	76.8	89	89.3	88.1	88.1		89.6
adil.flesh	h 95.4	95.4	95.1	81.4	79.9	92.4	2.66	98.2	98.2	89.6	

Table 10



BLASTN 2.1.2 [Nov-13-2000]

Reference:

Altschul, Stephen F., Thomas L. Madden, Alejandro A. Schäffer, Jinghui Zhang, Zheng Zhang, Webb Miller, and David J. Lipman (1997), "Gapped BLAST and PSI-BLAST: a new generation of protein database search programs", Nucleic Acids Res. 25:3389-3402.

RID: 984591695-10075-13605

Query-

(25 letters)

Database: nt

807,597 sequences; 2,863,827,885 total letters

If you have any problems or questions with the results of this search please refer to the BLAST FAOS

Taxonomy reports

Distribution of 500 Blast Hits on the Query Sequence

Mouse-over to show defline and scores. Click to show alignments

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Score
                                                                                                                                                                                                                                                                                               (bics) Value
        Sequences producing significant alignments:
       gb AF231651.1 AF231651 Strongylura notata clone HB-82 cytoc... 50 2e-05 gb AF231650.1 AF231650 Strongylura notata clone HB-159 cyto... 50 2e-05 ref NC 002672.1 Dinornis giganteus mitochondrion, complete... 50 2e-05 ref NC 002673.1 Emeus crassus mitochondrion, complete genome 50 2e-05
       ref NC 002673.1 Emeus crassus mitochondrion, complete genome
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       gb/AF232015.1/AF232015 Nothrotheriops shastensis cytochrome...
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       gb|AF232013.1|AF232013 Bradypus variegatus cytochrome b gen...
     gb|AY016015.1| Emeus crassus mitochondrion, complete genome
gb|AY016013.1| Dinornis giganteus mitochondrion, complete g...
gb|AY016014.1| Dromaius novaehollandiae mitochondrion, part...
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      gb|AF230167.1|AF230167 Bonasa umbellus cytochrome b (CYTB) ...
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      50
      2e-05

      qb/AF283631.1/AF283631
      Elaphe obsoleta
      LSUMZ 41189
      cytochro...
      50
      2e-05

  | GD|AF283631 1|AF283631 | Elaphe obsoleta LSUMZ 41197 cytochro | GD|AF283630 1|AF283630 | Elaphe obsoleta LSUMZ 41189 cytochro | GD|AF283629 | Elaphe obsoleta LSUMZ 41188 cytochro | Elaphe obsoleta LSUMZ 41188 cy
                                                                                                                                                                                                                                                                             50 2e-05
50 2e-05
50 2e-05
50 2e-05
   gb|AF283628.1|AF283628 Elaphe obsoleta LSUME 41187 cytochro...
                                                                                                   Elaphe obsoleta LSUMZ 41186 cytochro...
Elaphe obsoleta LSUMZ 40943 cytochro...
   qb|AF283627.1|AF283627
                                                                                                                                                                                                                                                                              50 2e-05
50 2e-05
50 2e-05
   gb | AF283626 . 1 | AF283626
                                                                                                      Elaphe obsoleta LSUMI 37499 cytochro...
   qb[AF28]625.1[AF28]625
                                                                                                    Elaphe obsoleta LSUME 44480 cytochro...
  qb|AF283624.1|AF283624
                                                                                                   Elaphe obsoleta LSUMZ 44451 cytochro...
Elaphe obsoleta LSUMZ 40444 cytochro...
  qb[AF283623.1[AF28]62]
                                                                                                                                                                                                                                                                                   50 2e-05
 gb[AF28]622.1[AF28]622
                                                                                                                                                                                                                                                                                  50 2e-05
50 2e-05
                                                                                                     Elaphe obsoleta LSUMZ 19925 cytochro...
  qb|AF283621.1|AF283621
                                                                                                     Elaphe obsoleta LSUMZ 37161 cytochro...
  qb[AF283620.1[AF283620
                                                                                                                                                                                                                                                                              50 2e-05
50 2e-05
                                                                                                     Elaphe obsoleta LSUMIJ9162 cytochrom...
  95 AF 28 36 19 . 1 AF 28 36 19
                                                                                                      Elaphe obsoleta LSUME HISB96 cycochr ...
 gb|AF283618.1|AF283619
gb|AF283617.1|AF283617
                                                                                                                                                                                                                                                                                 50 2-05
                                                                                                      Elaphe obsoleta LSUME H15892 cytoche...
```

Table 9. Animals selected for validation of minimum P'S score for efficient amplification of DNA templates in PCR

SL.	Name	P. S/AFF	P, S/AFR
1	Indian black buck (Antilope cervicapra)	97, 58	96, 54
2	Sheep (Ovis	87, 53	96, 54
3	Pig (Sus scrofa)	87, 52	87, 41
4	Fresh water dolphin (Platanista gangetica)	86, 49	82, 47

Sequences producing significant alignments:	Score E (bics) Value
	50 2e-05
gb[AF2]1651.1 AF231651 Strongylura notata clone HB-82 cytoc gb[AF231650.1 AF231650 Strongylura notata clone HB-159 cyto	50 2e-05
ref[NC 002672.1] Dinornis giganteus mitochondrion. complete	_50 2e-05
ref[NC_002673.1] Emeus crassus mitochondrion, complete genome	_ <u>5</u> 0
db AF232015.1 AF232015 Nothrotheriops shastensis cytochrome db AF232013.1 AF232013 Bradypus variegatus cytochrome b gen	<u>50</u> 2e-05 50 2e-05
gb[AF232013.1[AF232013 Bradypus variegatus cytochrome b gen gb[AY016015.1] Emeus crassus mitochondrion. complete genome	50 2e-05
gb[AY016013.1] Dinornis giganteus mitochondrion, complete g	<u>50</u> 2e-05
gb AY016014.1 Dromaius novaehollandiae mitochondrion, part	50 2e-05
gb AF230167.1 AF230167 Bonasa umbellus cytochrome b (CYTB)	<u>50</u> 2e-05 50 2e-05
gb AF074594.1 AF074594 Baeolophus bicolor cytochrome b gene gb AY005210.1 Poospiza melanoleuca isolate 3 cytochrome b	50 2e-05
gb AY005209.1 Poospiza melanoleuca isolate 2 cytochrome b	50 / 2e-05
gb[AY005208.1] Poospiza melanoleuca isolate 1 cytochrome b	<u>50</u> 2e-05
gb Ay005205.1 Poospiza hispaniolensis cytochrome b (cytb)	<u>50</u> 2e-05 50 2e-05
gb[AY005204_1] Poospiza garleppi cytochrome b (cytb) gene gb[AY005203_1] Poospiza erythrophrys cytochrome b (cytb) ge	<u>50</u> 2e-05 50 2e-05
<u>cb[AY005203.1]</u> Poospiza erythrophrys cytochrome b (cytb) ge <u>qb[AY005201.1]</u> Poospiza boliviana cytochrome b (cytb) gene,	50 2e-05
cb[AY005199.1] Poospiza alticola isolate 2 cytochrome b (cy	<u>50</u> 2e-05
gb AY005198.1 Poospiza alticola isolate 1 cytochrome b (cy	50 2e-05 50 2e-05
gb AF155870.1 AF155870 Heterocephalus glaber cytochrome b (gb AF189123.1 AF189123 Glyptotermes eukalypti cytochrome b	<u>50</u> 2e-05 50 2e-05
gb AF189123.1 AF189123 Glyptotermes eukalypti cytochrome b gb AF102099.1 AF102099 Criniferoides leucogaster cytochrome	50 2e-05
ch[AF102095.1[AF102095S1 Corythaixoides concolor cytochrome	50 2e-05
gb AF271065.1 AF271065 Mustela erminea specimen-voucher AF1	50 2e-05
GD AF243857.1 AF243857 Strongylura notata notata cytochrome	50 2e-05 50 2e-05
gb AF243856.1 AF243856 Strongylura notata forsythia cytochr ref NC 001567.1 Bos taurus mitochondrion, complete genome	50 2e-05
cb[AF306872.1]AF306872 Brachyramphus marmoratus haplotype M	50 2e-05
chlaf306871.1[Af306871 Brachyramphus marmoratus haplotype M	50 2e-05
gb AF306870.1 AF306870 Brachyramphus brevirostris haplotype	50 2e-05 50 2e-05
gb AF306869.1 AF306869 Brachyramphus brevirostris haplotype gb AF306868.1 AF306868 Brachyramphus brevirostris haplotype	50 2e-05
ghlafol0406.1 AF010406 Ovis aries complete mitochondrial ge	50 2e-05
gb AF248662.1 AF248662 Gryllus campestris haplotype 2 cytoc	50 2e-05 50 2e-05
gb AF248661.1 AF248661 Gryllus campestris haplotype 1 cytoc gb AF096462.1 AF096462 Rhipidura albicollis cytochrome b ge	<u>50</u> 2e-05 50 2e-05
gb AF096462.1 AF096462 Rhipidura albicollis cytochrome b ge gb AF283644.1 AF283644 Elaphe obsoleta cytochrome b gene, c	50 2e-05
gb AF283643.1 AF283643 Elaphe obsoleta cytochrome b gene, c	50 2e-05
gb AF283642.1 AF283642 Elaphe obsoleta cytochrome b gene, c	50 2e-05 50 2e-05
gb AF283641.1 AF283641 Elaphe obsoleta cytochrome b gene, c gb AF283640.1 AF283640 Elaphe obsoleta cytochrome b gene, c	50 2e-05
gb AF283639.1 AF283639 Elaphe obsoleta cytochrome b gene, C	50 2e-05
gb AF283637.1 AF283637 Elaphe obsoleta LSUMZ 45359 cytochro	<u>50</u> 2e-05 50 2 2 -05
gb AF283636_1 AF283636 Elaphe obsoleta LSUMZ 44662 cytochro gb AF283635_1 AF283635 Elaphe obsoleta LSUMZ 40443 cytochro	50 2e-05 50 2e-05
gb[AF283635.1[AF283635 Elaphe obsoleta LSUMZ 40443 cytochro gb[AF283634.1[AF283634 Elaphe obsoleta LSUMZ 44335 cytochro	50 2e-05
gb AF283633.1 AF283633 Elaphe obsoleta LSUMZ 42624 cytochro	<u>50</u> 2e-05
gb AF283632.1 AF283632 Elaphe obsoleta LSUME H1911 cytochro gb AF283631.1 AF283631 Elaphe obsoleta LSUME 41197 cytochro	<u>50</u> 2e-05 50 2e-05
qb AF283631.1 AF283631 Elaphe obsoleta LSUMZ 41197 cytochro qb AF283630.1 AF283630 Elaphe obsoleta LSUMZ 41189 cytochro	50 29-05
gb AF283629.1 AF283629 Elaphe obsoleta LSUME 41188 cytochro	50 2e-05 50 2e-05
gb AF283628.1 AF283628 Elaphe obsoleta LSUMZ 41187 cytochro gb AF283627.1 AF283627 Elaphe obsoleta LSUMZ 41186 cytochro	50 2e-05 50 2e-05
gb AF283626.1 AF283626 Elaphe obsoleta LSUMZ 40943 cytochro	50 2e-05
gb AF283625.1 AF283625 Elaphe obsoleta LSUMZ 37499 cytochro	50 2e-05 50 2e-05
gb AF283623.1 AF283623 Elaphe obsoleta LSUMZ 44480 cytochro gb AF283623.1 AF283623 Elaphe obsoleta LSUMZ 44451 cytochro	50 2e-05
gb[AF28]622.1[AF28]622 Elaphe obsoleta LSUM2 40444 cytochro	50 Ze-05
gb[AF28]621.1[AF28]621 Elaphe obsoleta LSUMI 19925 cytochro	50 2e-05
gb AF28]620,1 AF28]620 Elaphe obsoleta LSUM2]716] cytochro gb AF28]619,1 AF28]619 Elaphe obsoleta LSUM2]7162 cytochrom	50 2e-05 50 2e-05
GDIAF283618.11AF283618 Elaphe obsoleta LSUMZ H15896 cytochr	50 Ze-05
qb AF283617,1 AF283617 Elaphe obsoleta LSUM2 HIS892 cytochr	<u>50</u> 2e-05
qb AF283616.1 AF283616 Elaphe obsoleta LSUME 15831 cytochro	50 Ze-05 50 Ze-05
gblAF283614.11AF283614 Elaphe obsoleta LSUME H15887 cytochr	50 305
gb[AF78]61] 1[AF78]61] Elaphe obsoleca LSUM2 H15888 cytoche	52 205

gb[AF283612.1]AF283612	Elaphe obsoleta LSUMZ H15884 cytochr	50	2e-05
qh[AF283611.1[AF283611	and the checkets ISUME HISUII Cytochi	_50	2e-05
gb AF283610.1 AF283610	alle checlera LSUMI H15030 Cytocht	_50	2e-05 2e-05
gb AF283609.1 AF283609	minum checiera CAS 169468 CYCOCHIOM	50	2e-05
95 AF283608.1 AF283608	Planta checieta LSUME H14782 Cytocht	_50	2e-05
95 AF283607.1 AF283607	Flank checleta LSUME H14781 Cytocht	_50	2e-05
gb AF283606.1 AF283606	Flanks charleta LSUME H14724 Cytochi	50	2e-05
GD AF283605.1 AF283605	Elaphe obsoleta cytochrome b gene. C	_ <u>5</u> 0 _50	2e-05
gb/AF283604.1 AF283604	Elaphe obsoleta cytochrome b gene, C	50	2e-05
GD[AF283603.1[AF283603	Elaphe obsoleta cytochrome b gene, C	50	2e-05
gb AF283602.1 AF283602	Elaphe obsoleta LSUMI H3388 cytochro	50	2e-05
gb AF283601.1 AF283601	Elaphe obsoleta LSUMZ H3385 cytochro Elaphe obsoleta LSUMZ H3384 cytochro	50	2e-05
gb AF283600.1 AF283600	Elaphe bairdi LSUMZ H3382 cytochrome	50	2e-05
gb AF283599 . 1 AF283599	Elaphe bairdi LSUMZ H3381 cytochrome	50	2e-05
95 AF283598 .1 AF283598	Elaphe obsoleta LSUMZ H3379 cytochro	50	2e-05
95 AF283597 1 AF283597	Elaphe obsoleta LSUMZ 19616 cytochro	50	2e-05
gb AF283596 .1 AF283596	Flanka obsoleta LSUMZ H3376 cytochio	<u>50</u>	2e-05
95 AF283595 . 1 AF283595	Flanke obsoleta LSUMZ H3345 CYTOCHTO	_50	2e-05
gb AF283594 1 AF283594	Flanka obsoleta LSUMZ H3309 CYTOCHIO	_50	2e-05
gb AF283593.1 AF283593 gb AF283592.1 AF283592	Flanks obsoleta LSUMZ H3306 Cytochro	_50	2e-05
gb AF283591.1 AF283591	Flanks obsoleta LSUMZ H3Z76 Cytochio	50	2e-05
gb AF283590.1 AF283590	Finhe obsoleta LSUMZ H3246 CYTOCHIO	<u> 50</u>	2e-05 2e-05
gb AF283589.1 AF283589	Flanke obsoleta LSUMZ H3212 Cytochro	<u> 50</u>	2e-05
GD AF283588.1 AF283588	Flanka obsoleta LSUMZ H3209 CYtochio	<u> 50</u>	2e-05
cb AF283587.1 AF283587	Flanka chanleta LSUMZ H3206 cytochio	<u>50</u>	2e-05
Gb AF283586.1 AF283586	Elaphe obsoleta LSUMZ H3191 cytochro	<u> </u>	2e-05
CD AF283585.1 AF283585	Elaphe obsoleta LSUMZ H3190 cytochro	50	2e-05
CD AF283584.1 AF283584	Elaphe obsoleta LSUMZ H3189 cytochro	50	2e-05
cb AF283583.1 AF283583	Elaphe obsoleta LSUMZ H3188 cytochro	50	2e-05
cb AF283582.1 AF283582	Elaphe obsoleta LSUMZ H3186 cytochro Elaphe obsoleta LSUMZ H3169 cytochro	50	2e-05
GD AF283581 . 1 AF283581	Elaphe obsoleta CAS 203083 cytochrom	50	2e-05
SDIAF282580.1[AF283580	Elaphe obsoleta CAS 203079 cytochrom	_50	2e-05
gb AF283579.1 AF283579	Flanka obsoleta LSUMZ H2286 Cytochio	_50	2e-05
Gb AF283578.1 AF283578 Gb AF283577.1 AF283577	Finhe obsoleta CAS 208631 cycochrom	_50	2e-05
gb AF283576.1 AF283576	Flanka cheolera LSUMZ H2229 Cytochio	50	2e-05 2e-05
GD AF187030.1 AF187030	phinophylla numilio isolate TX46001	<u> 50</u>	2e-05
G5 AF310052.1 AF310052	pocenica historicolensis CyloChrome D	<u>50</u>	2e-05
GD AF310046.1 AF310046	Volatinia jacarina cytochrome b gene	50	2e-05
cb AF171919.1 AF171919	Deinagkistroden acutus cytochrome b Trimeresurus mucrosquamatus cytb gen	50	2e-05
GD AF171897.1 AF171897	Agelaius cyanopus cytochrome b (cytb	50	2e-05
gb[AF290174.1 AF290174	Agelaius phoeniceus cytochrome b (cy	50	2e-05
Gb AF290173.1 AF290173	Opinionalise major cytochrome b (cyto)	50	2e-05
gb AF290171.1 AF290171 gb AF290170.1 AF290170	Reblicators holosericeus cytochrome	_50	2e-05
gb AF290150.1 AF290150	Volarinia jacarina cytochrome b (cyt	50	2e-05
95 AF176252.1 AF176252	point adoptions acatecae cytochrome	50	2e-05 2e-05
gb AF176251 . 1 AF176251	pointrodontomys zacatecae cytochrome	<u>50</u> 50	2e-05
gb AF163907.1 AF163907	Microtus xanthognathus cytochrome b	<u> </u>	2e-05
GD AF163904.1 AF163904	Microtus pinetorum cytochrome b gene	50	2e-05
gb AF163901.1 AF163901	Microtus ochrogaster cytochrome b ge Microtus miurus cytochrome b gene. c	50	2e-05
gb[AF163899.1 AF163899	Microtus californicus cytochrome B (50	2e-05
95 AF163891 1 AF163891	wisconic abbreviatus cytochrome B (C	<u> 50</u>	2e-05
gb AF163890.1 AF163890 gb AF288524.1 AF288524	pipeophelye dussumieri isolate Germa	<u> 50</u>	Ze-05
GD AF288523.1 AF288523	nimochalve dussumieri isolate White	50	2e-05 2e-05
95 AF288522.1 AF288522	ninachelya dusaumieri isolate ALCY	<u>50</u> 50	2e-05
GD AF123530 . 1 AF123530	Psilopogon pyrolophus cytochrome b (50	2e-05
gb AF123512.1 AF123512	Eubucco bourcierii tucinkae cytochro Adolfus vauereselli cytochrome b gen	50	2e-05
95 AF206548 . 1 AF206548	comprehens ribiden cytochrome b gene	5.7	2e-05
95 AF197867.1 AF197867 95 U63397.2 SEU63397	viers auropaga cytochrome b gene, part	50	2e-05
The server of the Siener	in and aring the mitochondrion, compilies	50	2e-05 2e-05
TYEED 11.128:00 2012:1	any ampliantus mitochondrion, comple	<u>50</u>	2e-05
GDIAF141217.11AF141217	Designed incomes country Tanzania Cy	<u>50</u>	2e-05
ablaf101615.1 AF201615	Pancodon buchholzi cyoschrome b gene Bombus nevadensia cycochrome b gene	50	Ze-05
gb AF077920,1 AF077920	organica americanus cytochrome b (cy	52	Ze-05
95 AF190612.11AF190612	mirechondrion, complete genome	50	Ze-05
95 J01194 1180VMT 808	Cochlearius cochlearius cytochrome b	23	20-05

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gb/U89181.1/CAU89181 Chlorostilbon aureoventris cytochrome	50	24-05
gb UB4171.1 AFUP9171 Asio flammeus cytochrome b (cytb) gene		26-05
additional and the state of the	50	
gb AF217633.1 AF217813 Homoroselaps lacteus cytochrome b ge	50	
gb AF217822.1 AF217822 Hydrophis semperi cycochrome b gene		
qb AF217813.1 AF217813 Acanthophis antarcticus cytochrome b	_50	
<pre>gb[AF220408.1]AF220408 Calliophis kelloggi cytochrome b (cy</pre>	_5.0	
gb[AF126430.1[AF126430 Ellobius fuscocapillus cytochrome b	50	
gb[AF090337.1]AF090337 Aythya americana mitochondrion. comp	_50	2e-05
gb AF059111.1 AF059111 Sarkidiornis melanotos cytochrome b	_50	2e-05
gb[AF059053.1[AF059053 Aix sponsa cytochrome b gene, partia	50	2e-05
gb[AF099308.1[AF099308] Icterus wagleri wagleri cytochrome b	50	2e-05
	50	2e-05
	_50	2e-05
gb AF099294.1 AF099294 Icterus gularis tamaulipensis cytoch		2e-05
gb AF099293.1 AF099293 Icterus gularis gularis cytochrome b	_50	
gb[AF160610.1]AF160610 Cricetomys emini Cemi636 cytochrome	_50	2e-05
gb AF036280.1 AF036280 Tragelaphus strepsiceros cytochrome	_50	2e-05
gb AF036277.1 AF036277 Tragelaphus scriptus cytochrome b (c	<u>50</u>	
<pre>gb[AF036274.1] Tetracerus quadricornis cytochrome b (cytb)</pre>	_50	2e-05
qb[AF194218.1]AF194218 Phrynosoma platyrhinos cytochrome b	<u>_so</u>	2e-05
gb AF194216.1 AF194216 Urosaurus ornatus cytochrome b gene,	50	2e-05
ref[NC 002009.1] Artibeus jamaicensis mitochondrion, comple	50	2e-05
ref NC 001941.1 Ovis aries mitochondrion, complete genome	50	2e-05
ref[NC 000877.1] Aythya americana mitochondrion, complete g	50	2e-05
	50	2e-05
	50	2e-05
gb U27551.1 GCU27551 Grus canadensis tabida cytochrome b (c	50	2e-05
gb AF089058.1 AF089058 Quiscalus quiscula cytochrome b (cyt		2e-05
gb AF089055.1 AF089055 Quiscalus major cytochrome b (cytb)	<u> 50</u>	2e-05
gb AF089054.1 AF089054 Quiscalus lugubris cytochrome b (cyt	<u> 50</u>	2e-05
qb[AF089046.1]AF089046 Oreopsar bolivianus cytochrome b (cy	_50	
cb AF089042.1 AF089042 Molothrus badius cytochrome b (cytb)	_50	2e-05
gb AF089039.1 AF089039 Macroagelaius imthurmi cytochrome b	_50	2e-05
db AF089037.1 AF089037 Lampropsar tanagrinus cytochrome b (<u> 50</u>	2e-05
cb AF089026.1 AF089026 Gymnomystax mexicanus cytochrome b (<u> 50</u>	2e-05
gb/AF089025.1/AF089025 Gnorimopsar chopi cytochrome b (cytb	_50	2e-05
Gb AF089024.1 AF089024 Euphagus cyanocephalus cytochrome b	_50	2e-05
cb AF089023.1 AF089023 Euphagus carolinus cytochrome b (cyt	50	2e-05
cb AF089021.1 AF089021 Dives warszwewiczi cytochrome b (cyt	50	2e-05
cb AF089020.1 AF089020 Curaeus curaeus cytochrome b (cyto)	50	2e-05
cb AF089016.1 AF089016 Amblycercus holosericeus cytochrome	50	2e-05
gb/AF089013.1/AF089013 Agelaius xanthophthalmus cytochrome	_50	2e-05
gb AF089012.1 AF089012 Agelaius xanthomus cytochrome b (cyt	50	2e-05
gb AF089008.1 AF089008 Agelaius phoeniceus sub-species phoe	50	2e-05
qb AF089006.1 AF089006 .Agelaius humeralis cytochrome b (cyt	50	2e-05
cb/AF089005.1/AF089005 Agelaius cyanopus cytochrome b (cytb	50	2e-05
gb[AF108695.1]AF108696 Scolomys juruaense cytochrome B (cyt	50	2e-05
gb/AF108685.1/AF108685 Wiedomys pyrrhorhinos cytochrome B (50	2e-05
gb[AF108677.1]AF108677 Thomasomys oreas cytochrome B (cyt3)	50	2e-05
Gb AF145511.1 AF145531 Melanoplus foedus Cytochrome b gene,	50	2e-05
gb AF145511,1 AF145511 Melanoplus angustipennis cytochrome	50	2e-05
gb U89627.1 BMU89627 Bolicoglossa marmorea cytochrome b (cy	50	2e-05
q5 U89623.1 BPU89623 Batrachoseps pacificus cytochrome b (c	50	2e-05
gb AF181470.1 AF181470 Okapia johnstoni Cytochrome b gene,	50	2e-05
gb AF094075 1 AF084075 Lagenorhymchus acutus cytochrome b g	50	2e-05
gb[U90303.1[OMU9030] Ovibos moschatus cytochrome b (cytb) g	50	2=-05
ab U90302.1 OMU90302 Ovibos moschatus cytochrome b (cytb) g	50	2
gb[U90301.1]OMU90301 Ovibos moschatus cytochrome b (cytb) g	50	2 e -05
gb/U90300.1/OMU90300 Ovibos moschatus cytochrome b (cytb) g	50	Ze-05
gb[AF038883.1[AF03888] Deinagkistrodon acutus cytochrome b	_50	2e-05
gb[AF039268.1[AF039268 Agkistrodon contortrix cytochrome b	50	Ze-05
gb[AF0]9267.1[AF0]9267 Boa constrictor cytochrome b (cytb)	_50	24-05
gb S49215.1 S49215 apocytochrome b (sheep, domestic, Merino	_50	20-05
qb[AF158698.1[AF158698 Geomys pinetis dytochrome b gene. co	50	20-05
qb[AF158692.1]AF158692 Geomys bursarius jugossicularis cyto	50	Ze-05
qb[AF068193.1]AF068193 Ithaginia cruentus cytochrome b (cyt	50	2e-05
gb[AF091629.1]AF091629 Ancilocapra americana cytochrome b (_50	2=-05
db[AF02206] 11 Tragelaphus screpsiceros cytochrome b (cytb)	50	2=-05
gb[AF022062.t] Tragelaphus derbianus cytochrome b (cytb) ge	_5_7	24-05
qb[AF022060 1] Hippograque equinus cytochrome b (cytb) gene	50	25°-05
qb[AF022057.1] Tragelaphue oryx cytochrome b (cytb) gene, m	50	24-05
gb[AFII]500.1[AFI:]500 Lagenorhymchus acutus isolate LACU74	37	24-05
gblaF113499 1[AF113499 Lagenorhymchus acutus Lacus) .	20	203

		• ••
gb UC+645.1 LBUC9645 Loxocemus bicolor cycochrome b (cycb)	50	2e-05
This care a lammage to Eugentes notageus cytochrome b (cytb) 9	_50	2e-05 2e-05
This came a few reaches murinus cytochrome b (cyto) 9	_s <u>o</u>	
This care alreason Friends striatus fosteri Cytochrome	.50	2e-05 2e-05
Tirurant impressed Enjoyance striktus strigilatus CytoCRF	_50	2e-05
Throngos throughout Enigrapes strigitus strigilatus cytocht	_50	2e-05
Tillians, ilrovanta Frierand Striatus modraniel Cytogram	_50	2e-05
Lineards (Inchesors) Frictares striatus meetaniei eyoudu-om.	_50 _50	2e-05
		2e-05
gb U69790.1 EMU69790 Epicrates monensis cytochiome 5 (7)	_ <u>5,0</u> _5,0	2e-05
gb U69790.1 EMU69790 Epicrates fordi cytochrome b (cyto) ge	50	2e-05
gb U69784.1 EFU69784 Epicrates fordi cycochrome b (cycb) ge	50	2e-05
gb U69779.1 ECU69777 Epicrates cenchria cytochrome b (cytb) gb U69777.1 ECU69777 Epicrates cenchria cytochrome b (cytb)	50	2e-05
	50	2e-05
gb U69776.1 EAU69776 Epicrates angulifer cytochrome b (cytb	50	2e-05
gb[U69774.1] EAU69774 Epicrates angulifer cytochrome b (cyto) gb[U69772.1] CEU69772 Corallus enveris cytochrome b (cyto) g	50	2e-05
cb U69771.1 CEU69771 Corallus enveris cytochrome b (cyto) g cb U69771.1 CEU69771 Corallus enveris cytochrome b (cyto) g	50	2e-05
The same along careline envires even chrome b (cyto) 9	50	2e-05
Simple of the contract of the	_50	2e-05
threads 1/courses Candola aspera cytochrome b (cyto) gen	_50	2e-05
The same throws are constrictor evtochrome b (cyta) 98	_50	2e-05
discourse the construction of control ge	_50	2e-05
The state of the s	_50	2e-05
in the control of the americana mitochondrion, comple	_50	2e-Q5
Tilledocare illedocare Chemidobborus Eleris Strain 1514 Pay	_50	2e-05
Transport Timencast Chemidophorus tichis Strain 1814 omt	<u>50</u>	2e-05 2e-05
11-top and 11-top 4060 Connochaetes taurinus Cytochiome D 3	<u>50</u>	2e-05
GD/AF028822.1/AF028822 Alcelaphus buselaphus cycochiome b 9	<u>50</u>	2e-05
GD AF028821 1 AF028821 Damaliscus lunatus cytochrome D gall	50	2e-05
cb AF061340. I AF061340 Arcibeus Jamaicensis micochomatia	_50	2e-05
gb[AF076093.1[AF076093] Thalassarche impavida cytochrome b (50	2e-05
	50	2e-05
	50	2e-05
	50	2e-05
Thereses illegance Garrodia nereis cytochrome b (cytb)	50	2e-05
The series of th	_50	2e-05
Librariasa ilipareasa, niamedea dibsoni ovtochrome b (cyth)	_50	2e-05
Diomedea enomothora cytochrome b (CY	50	2e-05 2e-05
Limancore ilenacore Diomedea Chionoptera CytoChrome D (C	<u>50</u>	2e-05
ablar076047.1[ar076047 Diomedea antipodensis cycochiome 2	50	2e-05
gb/UE3314.1/MSU83314 Micrastur semitorquatus cytochrome b (50	2e-05
cb/U83318.1/MEU83318 Microhierax erythrogenys cytochrome b cb/U37303.1/SAU37303 Synthliboramphus antiquus cytochrome b	50	2e-05
	50	2e-05
	50	2e-05
- Language Brachy-amphus brevirostria cytochrome	_50	2e-05
The state of the s	50	2e-05
territor ilaminator derbia musilla eviochrome b gene, miso	50	2e-05 2e-05
db/U37087.1/ACU37087 Aechia cristatella cytochrome b gane,	<u>50</u>	2e-05
gb/U87525.1 HGU87525 Hecerocephalus glaber cytochrome-b gen	50	2e-05
gb/U87524.1 HGU87524 Heterocephalus glaber cytochrome-b gen gb/U87523.1 HGU87523 Heterocephalus glaber cytochrome-b gen	50	2e-05
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		2e-05
	50	
gb U17863.1 OAU17863 Oreamnos americanus cytochrome b gene	50	29-05
gb U17861.1 OAU17861 Oreamnos americanus cytochrome b gene gb U17862.1 OMU17862 Ovibos moschatus moschatus cytochrome	50 50	2e-05 2e-05
gb[U1786].1[OAU1786] Oreamnos americanus cytochrome b gene gb[U17862.1[OMU17862] Ovibos moschatus moschatus cytochrome gb[U17860.1[ODU17860] Ovis dalli cytochrome b gene. mitochon gb[U17859.1[OCU17859] Ovis canadensis cytochrome b gene. mito	50 50 50	2e-05 2e-05 2e-05
gb U1786].1 QAU1786] Oreamnos americanus cytochrome b gene gb U17862.1 QMU17862 Ovibos moschatus moschatus cytochrome gb U17860.1 QDU17860 Ovis dalli cytochrome b gene. mitochon gb U17859.1 QCU17859 Ovis canadensis cytochrome b gene. mitochon gb U65274.1 T8U65274 Thomomys bottae cytochrome b (cytb) ge	50 50 50	2e-05 2e-05
gb U1786].1 QAU1786] Oreamnos americanus cytochrome b gene gb U17862.1 QMU17862 Ovibos moschatus moschatus cytochrome gb U17860.1 QDU17860 Ovis dalli cytochrome b gene. mitochon gb U17859.1 QCU17859 Gb U65274.1 T8U65274 Thomomys bottae cytochrome b (cytb) ge gb U65267.1 T8U65267 Thomomys bottae cytochrome b (cytb) ge	50 50 50	2e-05 2e-05 2e-05 2e-05
gb U1786].1 OAU1786] gb U17862.1 OMU17862 gb U17860.1 ODU17860 gb U17859.1 OCU17853 gb U65274.1 TBU65274 gb U65260.1 TBU65260 gb U65260.1 TBU65260	50 50 50 50	2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05
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gb U1786].1 OAU1786] Oreamnos americanus cytochrome b gene gb U17862.1 OMU17862 Ovibos moschatus moschatus cytochrome gb U17860.1 ODU17860 Ovis dalli cytochrome b gene. mitochon gb U17859.1 OCU17853 Gb U65274.1 T8U65274 Thomomys bottae cytochrome b (cytb) gene. gb U65260.1 T8U65260 Thomomys bottae cytochrome b (cytb) gene. gb U65101.1 PAU65301 Perognathus amplus cytochrome b (cytb) gene. mitochrome b (cytb) gene.	50 50 50 50 50 50 50 50 50	2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05
gb U1786].1 OAU1786] Oreamnos americanus cytochrome b gene gb U17862.1 OMU17862 Ovibos moschatus moschatus cytochrome gb U17860.1 ODU17860 Ovis dalli cytochrome b gene. mitochon gb U17859.1 OCU17859 gb U65274.1 T8U65274 Thomomys bottae cytochrome b (cyto) gene. gb U65260.1 T8U65260 Thomomys bottae cytochrome b (cyto) gene. gb U65301.1 PAU65301 Perognathus amplus cytochrome b (cyto) gene. mitochon gb AF034719.1 AF034719 Gapra aegagrus cytochrome b (cyto) gene. mitochone.	50 50 50 50 50 50 50 50 50 50 50 50 50 5	2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05
gb U1786].1 OAU1786] Oreamnos americanus cytochrome b gene gb U17862.1 OMU17862 Ovibos moschatus moschatus cytochrome gb U17860.1 ODU17860 Ovis dalli cytochrome b gene. mitochon gb U17859.1 OCU17859 Gb U65274.1 T8U65274 Thomomys bottae cytochrome b (cyto) gene. gb U65260.1 T8U65260 Thomomys bottae cytochrome b (cyto) gene. gb U65101.1 PAU65301 Perognathus amplus cytochrome b (cyto) gene. gb AF034739.1 Capra caucasica cytochrome b (cyto) gene. gb AF034737.1 Capra cytochrome b (cyto) gene. gb AF034737.1 Capra cytochrome b (cyto) gene.	50 50 50 50 50 50 50 50 50 50 50 50 50 5	2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05
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gb U1786].1 OAU1786] Oreamnos americanus cytochrome b gene gb U17862.1 OMU17862 Ovibos moschatus moschatus cytochrome gb U17860.1 ODU17860 Ovis dalli cytochrome b gene. mitochon gb U17859.1 OCU17859 Gb U65274.1 T8U65274 Thomomys bottae cytochrome b (cyto) gene. gb U65260.1 T8U65260 Thomomys bottae cytochrome b (cyto) gene. gb U65101.1 PAU65301 Perognathus amplus cytochrome b (cyto) gene. gb AF034739.1 Capra caucasica cytochrome b (cyto) gene. gb AF034737.1 Capra cytochrome b (cyto) gene. gb AF034737.1 Capra cytochrome b (cyto) gene.	50 50 50 50 50 50 50 50 50 50 50 50 50 5	2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05 2e-05

gb[AF03472F.1] Ovis dall: dall: cytochrome t (cytb) gene, m	50	2e-
oblinosizata 11 Ovik ammon darwini cytochrome b (cyto) gene	<u>_5,0</u>	2e-
gb[AF034724.3]AF034724 Pantholops hodgsoni cytochrome b (cy	_50	2e-
gb[AF057130.1]AF057132 Taxides taxus cytochrome b (cytb) ge	.50	2e -
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The state of the s	_50	2e-
Liveries algories pelecanus onocrotalus cytochrome B gen	50	2e-
Anhinga cytochrome B gene, mit	_50	2e-
There is a languages Ambinga anhinga cytochrome B gene, mit	_50	2e-
illegate allangated Ambings appings evtochrome B gene, mit	<u> 50</u>	2e
Liverage ilcrusizes Chalodina longicollis cytochrome b gen	<u>50</u>	20
emblaJ277676.11ESC277676 Elaphe scalaris mitochondilat parc	50	2e-
emblaJ277675.11ESC277675 Elaphe scalaris mitochondrial partir	50	20
emb AJ277672.1 ELO277672 Elaphe longissima mitochondia	50	2e
emb[AJ277671.1]ELO277671 Elaphe longissima mitocholdi lairoch	<u></u>	20
emb[Y11832.1]MTDNCOMGN Dasypus novemcinctus complete mitoch	50	24
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The Tracera 1 Energes 78 Furronius depressirostris partial	<u>_50</u>	2=
This Tracert 11 Eproase 77 Futropius depressirostris partial	_50	24.
This Tracers 1/EDE745675 Europpius depressionstris partial	_50	26.
This tracere il ropassers Furropius depressirostris partial	_50	20
This theer, lightrasta Furtonius depressirostris partial	_ <u>50</u> _50	2e
TELLVISOR SIMPORFONDI Phea amedicana complete micochondia	_ <u>50</u>	Ze.
THIS PROPERTY DATES CALCULA CALCULATIONS DE GERE, MALOCALE	50	Ze:
q5/U48955.1/TMU48955 Thalassarche melanophris melanophris	50	2=
ablucass4 1 TCU48954 Thalassarche chrysostoma cytochrome b	50	7=
db[U48944.1]TCU48944 Thalassarche chlorothymchos chlorothym db[U48943.1]PPU48943 Phoebetria palpebrata cytochrome b (cy	50	2 e
- A	50	20
	_50	2 =
The grant throughout nicondea exulans dabbenena cycochiome	50	2=
The live and the live and D. omedes enomorbors saniard: cytochiom	50	2=
GETUNG 746 LIVEUNG 745 OLUMBURM APT TENTE	50	2=
luande il natuente n'emedea amsterdamensia cytochrome o l	_50	2e 2e
gb U49949 1 DAU49949 Diomedea amsterdamensia cytochrome b (
gb U48948 1 DAU48948 Diomedea amsterdamensia cytochrome b Company Diomedea amsterdamensia cytochrome b Diomedea amsterdamensia cytochrome b	_57	
gb[U48948.1]DAU48948 Diomedea amsterdamensia cytochrome 5 (10 10

gb[U66505.1]ALU66505 Artibeus lituratus cytochrome b (cytb)	50	26-05
gb[U6.6505.1]ALU66505 Artibeus lituratus cytochrome b (cyto) gb[U6.6504.1]AJU66504 Artibeus]amaicensis cytochrome b (cyto	50	2e-05
gb/U66504.1[AJU66504 Artibeus Jamaicensis cytochrome b (cyt gb/U66503.1[AJU66503 Artibeus Jamaicensis cytochrome b (cyt	50	26-05
	50	2e-05
gb[U66502.1]AIU66502 Artibeus intermedius cytochrome b (cyt	50	2e-05
db[066501.1]AIU66501 Artibeus inopinatus cytochrome b (cytb	<u>-</u>	2e-05
THISESON I LANGESON Arribeus hirsutus cytochrome o (cyto)		2e-05
Thirties and I lattices and Artibeus fracerculus cytochrome b (cyt	50	
qb[U66498.1]AFU66498 Artibeus fimbriatus cycochrome b (cytb	<u>. 50</u>	2e-05
	_50	2e-05
	50	2e-05
	50	26-05
	50	2e-05
gb/U63058.1/BBU63058 Brachyramphus brevirostris cytochrome	50	2e-05
There are alsting and scolomys augusense cytochrome o ley-b		2e-05
This is the incommentation of the control of the co	_50	
Thirder 1 1 MM 134672 Merachirus nudicaudatus cytochrome b I	<u> 50</u>	2e-05
Thirder 1 1 MM 134671 Merachisus nudicaudatus cytochrome b 1	50	2e-05
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Citation of the contract of th	<u>. 5</u> 0	2e-05
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GD L11904.1 CGYMTCYTBC Cratogeomys goldmani goldmani mitoch	50	2e-05
The variant of Contract S but or its mitochondrial DNA for Cytoch	50	2e-05
This care ilaphacare anches richard: Cytochrome b gene, mit		2e-05
Thirderes alastiagree Anthus berthelotic cytochrome b gene,	_50	2e-05
Third star ilegiasias Sciums stramineus cytochrome b gene,	_50	
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	50	2e-05
emb X36743.1 MTCACYT6 C.aura mitochondrial cyth gene	50	2e-05
dbi/A3035242.1/A3035242 Pantodon buchhelzi mitochondrial cy	50	2e-05
emb/X60946.1/MITDCB33 T. dorbignyi mitochondrial gene lot c		2e-05
This topogram I IMTETOVERS Rangifer tarancus mitochondrial Cy	_50	2e-05
THE LYGARD A MATRECYTE D FASCIATA MICOCHONGITAL CYTOCHTOMS	_50	2e-05
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amblyscan ilmidacyta o aries mitochondrion cyth gene for c	_50	Ze-05
la Topogra Ilmimecypar Dama dama mitochondrial cyth gene	<u> 50</u>	2e-05
emb X72005.1 MILWCYT3 L.weddelli mitochondrial gene for cyt	_50	2e-05
emb Y08814.1 MIHLCYTBG H.liberiensis mitochondrial cytochro	_50	2e-05
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	_50	Ze-05
	_50	2e-05
	50	2e-05
SD L19718.1 AIUMICYTB Artibeus lituratus mitochondrial cyto	50	2e-05
cb U27543.1 BRU27543 Balearica regulorum cytochrome b (cytb	50	2e-05
dellaggagge llaggagge Sciurus stramineus mitochondriai Cy	50	2e-05
gb U18258.1 SCU18258 Spharagemon campestris cytochrome b ge	50	2=-05
Third area ileminates Sobaragemon college cytochiome o gene,	50	2=-05
animages almost great Trimeror topis distrinaria Cytochrome b		20-05
Limiaren ilenmaren Campula pellucida eytoenzome o gene, m	50	2e-05
This rank ilection and disconnective carlinianus mitochonerion	_50	20-05
distinguished (Committee Canas falconeri mitochongrial UNA 101	<u> 50</u>	25-05
during age 1 hgrage and rayanicus micochondrial DNA for cyt	<u>50</u>	26-05
designation of the state of the	<u> 50</u>	2e-05
dhilparia ilcommonal Carricomis crispus mitochondrial ge	<u> 50</u>	29-05
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dhilanging: Hanging: Cervus mippon keramae mitochondrial	-50	2e-05
and languist? Ilanguist? Corrus slaphus mitochondrial DNA 10	50	2=-05
distancing itsparting sheep misochandrial DNA for cycochrom	50	2=-05
dhilbscholder over musimon mitochondrial DNA for cy.	-5.0	2=-05
dhe 1734636 1 BOUNTTON Bon tavanique micochondrial gene for	<u> 57</u>	2=-05
childles il novembe novine mitochondrial gene for cytoche.	22	24-05
obilotilus ilognamenta oreamnos americanus mitochondriai gr	-20	205
db-(D)1195 (NACMTERIC Nemorhaedus goral mitochondrial gene	23	34.02

	50	26-05
del[D32192.1] CEUMTCELL Cervus hippon mitochondrial gene for		26-05
Harlenging Theory are Character elaphus scoticus micochondit	<u>_5</u> 0	2e-05
deilaunnings ilagnings carris elaphus canadensis mitochong	.sn	2e-05
db] AB021093.1 AB021091 Cervus nippon nippon mitochondrial	SC	
	_50	2e-05
dbj AB001090.1 AB001090 Cervus nippon pulchellus mitochondi dbj (AB008339.1 AB008539 Dinodon semicarinatus mitochondrial	_5,0	2e-05
dbj AB00833.1 AB008535 Dinodon semicarinatus mitochondrial dbj AB008800 Ovis aries mitochondrial DNA for Cy	50	2e-05
	_50	2e-05
	50	2e-05
gb L08032.1 CPLMTCYTEA Carcharhinus plumbeus micochondrial	_5°0	2e-05
gb[L28941.1]URRCYS Uroderma bilobatum cytochrome b gene. 5'	50	2e-05
chling37.1[CDECY3 Chiroderma coriae cycochrome b gene, b	<u> 50</u>	2e-05
Citatological logyological Capita Dyranaica (incividual in/ mail	_ <u></u> 50	2e-05
The totage a licevologia Capta evienalica (individual 14/ marri		2e-05
later one 1 1 CDV010053 Capra Dyrenaica (individual 10) mi	_50	2e-05
il recognizione Carra pyrenaica (individual 9) mic	50	
Taranga alcayologa Canta avrenaica (individual 8) mit	50	2e-05
Tarance alcoverage Canta pyrenaica (individual 7) mic	_50	2e-05
(individual 6) mic	_50	2e-05
	<u>_5,</u> 0	2e-05
	_50	2e-05
emb[AJ010047.1]CPY010047 Capra pyrenaica (individual 4) mit	50	2e-05
emb X95777.1 CLMCB C.longirostris mitochondrial cytochrome emb AJ009879.1 ClB9879 Capra ibex nubiana mitochondrial cyt	50	2e-05
	50	2e-05
	50	2e-05
gb U08946.1 CAU08946 Coragyps atratus mitochondrion cytochr	50	2e-05
gb U08945.1 C3U08945 Cathartes burrovianus mitochondrion cy	50	2e-05
THE COUNTY OF THE CANADAL WILLIAM CANADAL CANA	50	2e-05
The state of the s	50	2e-05
timents tiperreach Dhoericopearus riber mitochonditon Cit	50	2e-05
The same throwes a referance mitochondrial cycodia one of general	<u> 50</u>	2e-05
		2e-05
Liverzon linemes a stherrist mitochondrial cytochrome b gene	_50	8e-05
Alces alces cytochrome b (cytb) gene	_43	3e-04
Tamandua tetradactiva clone 7 cytoch	45	3e-04
Tamandua terradactyla clone 6 miroch	45	
Tamandua tetradactyla clone 5 cytoch	45	3e-04
The state of the s	45	3e-04
Terrace 1127157465 Tenus Granatensis Cytochrome b (Cyb)	45	3e-04
Taristaca alamastaca Lenus considerus haplocype i cytochi	45	3e-04
Lanus corsicanus haplotype 3 cytocar	45	3e-04
The state of the s	<u>46</u>	3e-04
Triper transfer Triper secondilus crocodilus cyto	<u> </u>	3e-04
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Tringuis cracadilus clone H3-156 Cy	45	3e-04
Tylogy tis market Tylogy Tylogy acus pacificus cytochiome	_ * 5	3e-04
Tylosumus acus melanotus clone Santania	45	39-04
the manager the manager Tylosumus acus melanotus clone S.A	<u> 45</u>	3e-04
lampager ilampager Tylogumus acus imperialis cytochrome	45	3e-04
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Alerratica theratical ablances hians cytochrome b oxidase	<u> </u>	3e-04
terrance alignment and tetradactyla clone 3 cytoch	45	3e-04
Tamandua retradactivia clone i cytoch	45	3e-04
distribute allegates Mylodon darwinii cytochrome b gene	45	36-04
There are a large and the mississippiens is isolate S	_ 55	30-04
Alegansia ileganses miligator mississippiensis isolate S	45	34-04
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gb[AFJ18552.1[AFJ18552 Alligator mississippleners isolate	45	304
gb[AF]18551.1[AF]18551 Alliqueor mississipplensis testite	45	3-04
go[AFJ18550 1]AFJ18550 Alligator mississipplensis 130.31-	45	304
gb[AF]18549 1[AF]18549 Alligator mississippiensis isolate	_ 9.5 _ 9.5	3-04
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ghiafilgos: ilafilgos) Sorex monticolus specimen voucher Af		

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Alignments

tmpseq_0	1	taccatgaggacaaatatcattctg	25
AF231651	398		422
AF231650	398		422
NC 002672	15560		15584
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AF288523	401	425
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AF123530	303	327
AF123512	303	127
AF206548	303	
AF197867	401	425
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AF019268	192		416
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AF158693	398		425
AF069191	401		443

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AF022062	398		422
AF022060	398		422
AF022057	398	*********	422
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<u>U69799</u>	374		398
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U69772	374		398
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<u> U69769</u>	374		98
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U17859	329		353
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U65267	398		422
U65260 U65301	396 396		422
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AF034736	398		422
AF034735	398		422
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AF006213	303		327 327
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AJ277671	299		323 14592
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U63058	302	
U58386	398	422
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U34672	398	
U34671	398	
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Y14371	398	422
L11909	398	422
L11901	398	
L11904	398	422
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<u> X95768</u> X95767	303	327
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X86754	299	323
X86743	299	323
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U18257	169	
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U18250	169 169	193
<u>U17904</u> D84202	398	422
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D32195	243	267
D32191	398	
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AB021097	398	422
AB021095 AB021094	398 398	
ABUZIUYA	398	
8021091	396	422

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AB001612	398	• • • • • • • • • • • • • • • • • • • •	422
D84205	398	• • • • • • • • • • • • • • • • • • • •	422
D64203	398	•••••	422
D34636	398		422
D34635	398	• • • • • • • • • • • • • • • • • • • •	267
<u>D32198</u>	243		267
<u>D3318</u> ë	243		422
<u>D32192</u>	398		422
<u>AB021099</u>	398		422
<u>AB021096</u>	398		422
AB021093	398		422
AB021090	398		15326
AB008539	15302		422
AB006800	398		284
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L08032	401		422
L28941	398 398		422
L28937	269		293
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AJ010054	269		293
AJ010053	269		293
AJ010052 AJ010051	269		293
	269		293
AJ010050 AJ010049	269		293
	269		293
AJ010048 AJ010047	269		293
X95777	407		431
AJ009879	269		293
AJ010055	269		293
U08946	303		327
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U08941	303		327
U08940	303		327 327
X95775	303		327
X95774	303		327
X95764	303		310
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AF157466	324		346
AF157465 AF157464	324		346
AF157463	324		346
AF157460	321		343
AF231664	400		422
AF231663	400		422 422
AF231662	400		422
AF231660	400		422
AF231659	400		422
AF231658	400		422
AF231657	400 400		422
<u>AF231656</u> AF231644	400		422
AF231639	400		422
AF232019	400		422
AF232017	400		422 422
AF232014	400		
AF318564	345		_
AF318563	345		366
AF118562	344		
AF318561	344 344		_
AF318560	344		366
AF118552 AF118558	344		3.6.6
AF318557	361		403
AF118555	361		403
AFTIBSSS	361	**********	403
SETTE SET	361	• • • • • • • • • • • • • • • • • • • •	

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AF118551	381		403
AF318552	186	****************	403
AF318551	381	* * * * * * * * * * * * * * * * * * * *	403
AF318550	381		403
AF318549	381	• • • • • • • • • • • • • • • • • • • •	403
AF318548	381		403
AF238041	400		422
AF326272	400		422
AF326271	400		422
AF326270	400		422
AF326266	40C		422
AJ004340	302	n	326
AJ004264	302		326

Database: nt

Posted date: Mar 2, 2001 12:20 AM

Number of letters in database: 2,863,827,885 Number of sequences in database: 807,597

Lambda 0.711 1.37

Gapped

Lambda K

0.711 1.31 1.37

Matrix: blastn matrix:1 -3

Gap Penalties: Existence: 5, Extension: 2

Number of Hits to DB: 39355 Number of Sequences: 807597 Number of extensions: 39355

Number of successful extensions: 15066

Number of sequences better than 10.0: 5706

length of query: 25

length of database: 2,863,827,885

effective HSP length: 17

effective length of query: 8 effective length of database: 2,850,098,736

effective search space: 22800789888

effective search space used: 22800789888

T: 0 A: 30

X1: 6 (11.9 bits)

X2: 15 (29.7 bits)

S1: 12 (24.3 bits)

S2: 16 (32.2 bits)

Table 11. BLAST analysis of primers 'mcb869' in nr database of NCBI. It demonstrates that the 3' end of this primer is highly conserved among a vast range of animal species. It also shows the significant homology among the primer and templates (i.e. the cytochrome b gene fragment of different animal species), confirming the universal nature of our primer.



BLASTN 2.1.2 [Nov-13-2000]

Reference:

Altschul, Stephen F., Thomas L. Madden, Alejandro A. Schäffer, Jinghui Zhang, Zheng Zhang, Webb Miller, and David J. Lipman (1997), "Gapped BLAST and PSI-BLAST: a new generation of protein database search programs", Nucleic Acids Res. 25:3389-3402.

RID: 984593033-24247-14777

Query-

(26 letters)

Database: nt

807,597 sequences; 2,863,827,885 total letters

If you have any problems or questions with the results of this search please refer to the <u>BLAST FAQs</u>

Taxonomy reports

Distribution of 500 Blast Hits on the Query Sequence

Mouse-over to show defline and scores. Click to show alignments	!
×	
	1
1	
	}

901AF991779.11AF991779

Score

Ξ

Value (bits) Sequences producing significant alignments: 6e-06 _52 gb[AF189111.1|AF189111 Cryptotermes austrinus cytochrome b ... 52 6e-06 gb|U86834.1|U86834 Phyllotis wolffsohni MSB 67270 cytochrom... 52 60-06 qb[AF123633.1|AF121633 Perissocephalus tricolor cytochrome ... 52 52 6ė-06 gb|AF123617.1|AF123617 Pipreola arcuata cytochrome b gene, ...
gb|AF127202.1|AF127202 Hylopezus fulviventria cytochrome b ... 6e-06 52 6e-06 qb|AF127194.1|AF127194 Grallaria guatimalensis cytochrome b... gb|AF217828.1|AF217828 Aspidelaps scutatus cytochrome b gen... gb|AF160578.1|AF160578 Hypogeomys antimena Hant555 cytochro... gb|AF009931.2|AF009931 Archocentrus centrarchus cytochrome ... gb[AF091629.1|AF091629 Antilocapra americana cytochrome b (... gb[AF034967.1] Sigmoceros lichtensteinii cytochrome b gene,... gb|AF038290.1|AF038290 Antechinus sp. cytochrome b gene, mi... gb U07577.1 AMU07577 Antechinus melanurus mitochondrion cyt...
gb U81343.1 CFU81343 Chelus fimbriata cytochrome b gene, mi... emb|AJ222631.1|ABCYTOB Alcelaphus buselaphus mitochondrial ... gb|M99464.1|PNZMTCYTB Planigale sp. cytochrome b gene, comp... emb|AJ225116.1|DNJ225116 Dryomys nitedula mitochondrial gen... gb U25738.1 PRU25738 Paradisaea raggiana cytochrome b gene, ...
gb U25736.1 PRU25736 Paradisaea rubra cytochrome b gene, mi...
gb U15202.1 SMU15202 Seleucidis melanoleuca mitochondrion c... gb|U15204.1|PR15204 Paradisaea raggiana mitochondrion cytoc... emb|X56290.1|MIDDCYTB D.dama mitochondrion cyth gene for cy... emb|X56286.1|MIAACYTBA A.americana mitochondrion cyth gene ... dbj D88639.1 D88639 Anoa depressicomis mitochondrial DNA f...
dbj D82890.1 D82890 Bubalus depressicomis mitochondrial DN... gb AF119261.1 AF119261 Peromyscus maniculatus cytochrome b ...
gb AF123615.1 AF123615 Rupicola rupicola cytochrome b gene,... gb AF160603.1 AF150603 Apodemus sylvaticus Asyl588 cytochro... 45 3e-04 45 3e-04 gb|U62637.1|CCOLCYTB2 Charadrius collaris cytochrome b (cyt... gb|U62685.1|CBICCYTB2 Charadrius bicinctus cytochrome b (cy... 45 3e-04 gb|AF022071.1| Madoqua guentheri cytochrome b (cytb) gene, ...
gb|AF022070.1| Madoqua kirkii cytochrome b (cytb) gene, mit... 46 3e-04 46 3e-04 45 3e-04 46 3e-04 gb U83317.1 PSU83317 Polihierax semitorquatus cytochrome b ...
gb U37293.1 CCU37293 Cepphus columba cytochrome b gene, mit...
gb U37292.1 CCU37292 Cepphus carbo cytochrome b gene, mitoc... 45 3e-04 gb/U37291.1/8MU37291 Brachyramphus marmoratus perdix cytoch... 45 3e-04 45 3e-04 45 3e-04 gb/AF082055.1/AF082055 Rupicola rupicola cytochrome b gene,... gb|U72770.1|JMU72770 Jabiru mycteria cytochrome b gene, mit...
gb|U07578.1|DCU07578 Dasycercus cristicauda mitochondrion c... 45 3e-04
46 0.001
41 0.001
41 0.001
41 0.001 SDIAF031908.1|GOCCCYTB1 Geopsittacus occidentalis cytochrom... emb|AJ004231.1|SBAJ4231 Sula bassana mitochondrial cyth gen...
emb|AJ004230.1|SBAJ4230 Sula bassana mitochondrial cyth gen...
emb|AJ004229.1|SBAJ4229 Sula bassana mitochondrial cyth gen... emb|AJ004232.1|SBAJ4232 Sula bassana mitochondrial cyth gen... qb[U88865.1] Pomacentrus sp. cytochrome b (cytb) gene, mito... gb|U90001.1|MBU90001 Morus bassanus cytochrome b gene, mito...
gb|U63057.1|SMU63057 Brachyramphus marmoratus perdix cytoch... dbj[AB036404.1]AB036404 Rana porosa brevipoda micochondrial... dbj|AB036402.1|AB036402 Rana porosa brevipoda mitochondrial... dbj|AB036400.1|AB036400 Rana porosa brevipoda mitochondrial... dbj[AB0]6198.1[AB0]6198 Rana porosa porosa mitochondrial CN... gb|Ul9611.1|JMUl9611 Jabiru mycteria cytochrome b gene, mit...
emb|X92519.1|HACYTB H.ampullatus cytochrome b gene (complet... Galeocerdo cuvier mitochondrial cyto...

Galeocerdo cuvier mitochondrial cyto...

Galeocerdo cuvier mitochondrial cyto... qb[AY015012.1] Crypturellus tataupa mitochondrion, partial ... gb[AF074591.1]AF074591 Petrochelidon pyrchonota cytochrome ... gb[AY005212.1] Podepiza whitii isolate 2 cytochrome b (cytb... gb[AY005211.1] Poospisa whitil isolate 1 cytochrome b (cytb... qb|AF187122.1|AF187122 Cryptotermes tropicalis cytochrome b ... qb[AF187119.1[AF187118 Cryptotermes primus isolate 2 cytoch... Cryptotermes primus isolate i cytoch ... 95 AF182117. 1 AF142117 Cryptotermen dudlyl cytochrome b (Cy... GOLAFIASLIS LIAFIASLIS Quie canadenais cytochrome b gene. p ... GOLARIIZIA9. LIAPITZIA9 0.001 4.1 Ovie canadensis canadensis cytochrom. ... gbiafililia Liafililia gbiafililia Liafililia Ovie canadensis nelsoni cytochrome b 0.991 0.001 vires ceesinii ceesinii apecimen vou

gb[AF081989.1]AF081989 Vireo cassinii c	assinii specimen-vou	44	0.001
gb AF081988.1 AF081988 Vireo cassinii c	assinii specimen-vou	44	0.001
gb AF081987.1 AF081987 Vireo cassinii c	assinii specimen-vou	44	0.001
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db/AF081984.1/AF081984 Vireo cassinii c	assinii specimen-vou	44	0.001
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gb AF081982.1 AF081982 Vireo cassinii c	assinii specimen-vou	44	0.001
db AF081981.1 AF081981 Vireo cassinii c	assinii specimen-vou	44	0.001
gb AF081980.1 AF081980 Vireo cassinii c	assinii specimen-vou	44	0.001
gb AF081979.1 AF081979 Vireo cassinii c	assinii specimen-vou	44	0.001
gb AF081978.1 AF081978 Vireo cassinii c	assinii specimen-vou	44	0.001
qb AF081977.1 AF081977 Vireo cassinii c	assinii specimen-vou	44	0.001
gb AF081976.1 AF081976 Vireo cassinii c	assinii specimen-vou	44	0.001
gb AF081975.1 AF081975 Vireo cassinii c.	assinii specimen-vou	44	0.001
gb AF081974.1 AF081974 Vireo cassinii c	assinii specimen-vou	44	0.001
qb AF081973.1 AF081973 Vireo cassinii c	assinii specimen-vou	44	0.001
gb AF081972.1 AF081972 Vireo cassinii c	assinii specimen-vou	44	0.001
chiarogia71 1 AFO81971 Vireo cassinii G	assinii specimen-vou	44	0.001
chlaF081970 1/AF081970 Vireo solitarius	alticola country US	44	0.001
chlaros1969 1 AF081969 Vireo solitarius	alticola country US	44	0.001
chlarogies ilafosies Vireo solitarius	alticola country US	44	0.001
chlaros1967 1/AF081967 Vireo solitarius	alticola country US	44	0.001
chiarcaiges liaroniges Vireo solitarius	solitarius specimen	44	0.001
Thistograms tiafograms Viteo solitarius	solitarius specimen	44	0.001
gb AF081964.1 AF081964 Vireo solitarius	solitarius specimen	44	0.001
gb/AF081962.1/AF081962 Vireo flavifrons	specimen-voucher LS	44	0.001
gb AF081961.1 AF081961 Vireo flavifrons	specimen-voucher LS	44	0.001
	leucophrys specimen	44	0.001
	otis leucotis cytoch	44	0.001
	ytochrome b (cytb) g aris isolate 3 haplo	44	0.001
	aris haplotype 3DH11	44	0.001
	aris haplotype 3DH15	44	0.001
<u>qb AF144315.1 AF144315</u> Amphiprion ocella <u>qb AF144314.1 AF144314</u> Amphiprion ocella	aris isolate 2 haplo	44	0.001
-blass44313 1 AF144313 Amphiprion ocella	aris isolate l haplo	44	0.001
-blas144312 1/AF144312 Amphinrion ocella	aris haplotype 3DH12	44	0.001
-blass44311 1/85144311 Amphinrion ocella	aris haplotype 3DHl	44	0.001
-blast44310 11aF144310 Amphinrion ocella	aris isolate 2 haplo	44	0.001
gb AF144309.1 AF144309 Amphiprion ocella	aris isolate 1 haplo	44	0.001
ref NC 001567.1 Bos taurus mitochondrio	on, complete genome	44	0.001
	cytochrome b gene,	44	0.001
	nystina cytochrome b:	44	0.001
	ea cytochrome b gene	44	0.001
gb AF096452.1 AF096452 Platysteira cyane gb AF283619.1 AF283619 Elaphe obsoleta i	LSUMZ39162 cytochrom	44	0.001
qb AF283618.1 AF283618 Elaphe obsoleta	SUMZ H15896 cytochr	44	0.001
-blaszesene ilaszesene Flanke obsoleta i	LSUMZ H14782 cytochr	44	0.001
gb AF283602.1 AF283602 Elaphe obsoleta I	LSUMZ H3388 cytochro	44	0.001
gb[AF310069.1[AF310069 Elaenia martinica	cytochrome b gene,	44	0.001
gb AF146616.1 AF146616 Actophilornis at:	cicanus cytochrome b	44	0.001
	latus cytochrome b (44	0.001
Tablet 002504 1 Lama pacos mirochondrio	on, complete genome	44	0.001
Thiastelent liastelent Microrus ochsogas	ites cytochsome b gs	44	a.aal
chiarilazes 1 AF119263 Myonus schisticol	lor cytochrome b gen	44	0.001
chiarilazza liarilazza synantomya boreal	lis cytochrome b gen	44	0.001
gb AF288454 . L AF288454 Nyccereuces procy	ronoides koreensis C	44	0.001
	egulus ecriplatus cy	11	0.001
aniAF123647 1 AF123647 MachaeropteF19 PV	rocephalus cycochro	4.1	0.001
chlaft21646 llAF123646 Xenoplog acconice	na cytochrome b gen	44	0.001
aniarizidas liarizidas piera fasclicauda	cytochrome b gene	44	0.001
qb[AF123634.1[AF123634 Pyroderus scucacu	is cytochrome b gene	44	a.aat
qb[AF1216]2.1[AF1216]2 Cephalopterus och	istue cytochrome b g	-11	100.0
	phyrolasma cytochroms		0.001
chiagizidig liagizidiz Ampelioides techu	idll cytochrome b ge	4 4	0.001
miagrificia clariticia placegia chiocole	pldoca cycocheoma b	4.1	0.001
quiariziale Liariziste Rupicole perivien	la cytochcome b gena	4.4	0.07L
qb[AF12351]_1[AF12351] Collocale ecleced	t cytochrome b gene	_4.4	

			0.001
gb AF127201.1 AF127201 Myrmothera	campanisona cytochrome b	44	0.001
chiagi27192 liAF127192 Grallaria	ruficapilla cytochrome D g	44	0.001
chiagrazias 1 AF127189 Grallaria	varia cycochrome b gene, p	44	0.001
chiaria7849 1 AF197849 Sericomis	frontalis cytochrome b ge	44	0.001
chiarro7847 1 AF197847 Pardalotus	striatus cytochrome b gen	44	0.001
slwc nonggg 11 Hippopotamus amp	hibius mitochondrion, comp	44	0.001
flac 002079 11 Caragging auratu	s mitochondrion, complete	44	0.001
FINC 001794 11 Macropus robustu	s mitachandrion, complete	44	0.001
alva conena 11 Didelphia virgia	iana mitochondrion, comple	44	0.001
1.5201612 1/AF201612 Stomatorbi	nus sp. CU79703 Cyotchrome	-44	0.001
125007021 11A5097931 Amphinrion	clarkii cytochrome b gene	44	0.001
Laggarate 1 Agragage Amphiprion	ocellaris cytochrome b ge	44	0.001
LITOLINA I POINT BOS TAUTIS MITO	chandrion, complete genome	44	
Lamacated 1 ASI68760 Amalone so	initera isolate ixac cycoc	44	0.001
-blastcored tlast68759 Analone sp	inifera isolate TXki Cytoc	44	0.001
LACTORTER LIBETERTSR Analone SD	inifera isolate TXcc cytoc	44	0.001
- Lagrange Ilagicarse Analone so	inifera isolate NMIG CytoC	44	0.001
Lagranzai ilagianzai Perrocheli	don rufocollaris isolate E	44	0.001
	don rufocollaris isolate E	44	0.001
	canus cytochrome b (cytb)	44	0.001
-blastoners 11AF193833 Botaurus 1	entiginosus cytochrome b g	44	0.001
-blas102022 1[AF193822 Ardea alba	cytochrome b gene, partia	44	0.001
1 = 1 0 2 0 2 1 1 A = 1 9 3 8 2 1 A = dea hero	dias cytochrome b gene, pa	44	0.001
This correct times 17837 Paramaia m	ultifasciata cytochrome b	44	0.001
LAFRITARE LIAFRITARIS Naja kaout	hia cytochrome b gene, com	44	0.001
.	colubrina cytochrome b gen	44	0.001
· lagger 1 lagging Callionhis	japonicus cytochrome b ge	44	0.001
· la marana alagatagaa Micharaide	s euryxanthus cytochrome b	44	0.001
· lacoracio ilaccionio Decedalia	coronata cytochrome b gene	44	0.001
: 1	superbus cycochrome b gen	44	0.001
gb AF118156.1 AF118156 Terenura h	umeralis specimen-voucher	44	0.001
gb AF209938.1 AF209938 Euura atra	isolate 62 cytochrome b g	44	0.001
gb AF209933.1 AF209933 Euura atra	isolate C cytochrome b ge	44	0.001
	ta angustirostris cytochro	44	0.001
	specularoides cytochrome	44	0.001
	brasiliensis cytochrome b s barbouri haplotype PH.22	44	0.001
	s barbouri haplotype PH.13	44	0.001
	gambianus Cgam518 cytochr	44	0.001
	emini Cemi511 cytochrome	44	0.001
· last cocto ilast cocto Criceromys	emini Cemi530 cytochrome	44	0.001
· La cacata alagacogua Cricatomya	emini Cemi637 cytochrome	44	0.001
The second signification Criceromy	emini Cemi636 cytochrome	44	0.001
1251 COCCA 1 1251 60604 Calomyscus	bailwardi Cbal576 cytochr	44	100.0
Lamacosco ilasicosco Eliurua ma	iori Emai642 Cycochrome D	44	0.001
Winds Comme Times and The Comme Times The	jori Emaj641 cytochrome b	44	0.001
- Lagrances 1128160558 Fligging Ma	jori Emaj639 cytochrome b	44	0.001
gb AF160557.1 AF160557 Eliurus ma	jori Emaj638 cytochrome b	44	a.aal
	jori Emaj614 cytochrome b	44	0.001
	jori Emaj617 cytochrome b jori Emaj573 cytochrome b	44	0.001
	jori Emaj556 cytochrome b	44	0.001
	jori EmajS61 cytochrome b	44	0.001
FIRE COSED TIMES COSED FILLING MA	iori Emaj443 cycochrome b	44	0.001
TELESTICASAS ILASICASAS ELIUTUS MA	iori Emaj444 cycochrome b	44	0.001
-6/35016297 1/AF016297 Damaliscus	ovgarque cytochrome b (cy	44	0.001
-5125016286 1125016286 Oroc Leuco	Elx chrochrome p (chrp) de···	44	0.001
qb[AF036283.1[AF03628] Antilope C	erricapra cytochrome b (cy	44	g . gg L
gb AF036281.1 AF036281 Antidorcas	maraupialia cycochrome b	4.4	0.001
	s oryx cytochrome b (cytb) s euryceros cytochrome b (4.1	0.001
	ornia cytochrome b (cytb)	44	0.001
CINC 001941 11 Owin seins miroc	hondrion, complete genome	14	0.001
- Lactoread 1 ASIGREGA MICTORYTOM	va minutua cytochrome B (C	44	0.001
Winesages Ilacinges enintdomys	nicela cycochrome 8 (cyc8	44	0.001
qb[AF10867] L[AF10967] Thomasomys	daphne cytochrome B (cytb	44	a.aar
GOLAFIORSS LIAFIDESS SCAPERCOMY	e tumidue cytochrome 8 (cy	43	a.aat
LIAGOATTO IIADOATTO MAGAMUSELA	cue vuquangenele cytochrom	11	0.001
ablandaria II Munclacus muncjak	difficulting changing and a con-	3 3	0.001
gbiarragai.ileragai scenelle c	perstensibe cytocheome b g .	23	0.333
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qb AF084074.1 AF084074 Lagenorhynchus albirostris cytochrom	.44	0.001
gb AF090750.1 AF090750 Gobio gobio balcanicus cytochrome b	44	0.001
gb/AF090750.1/AF090750 Gobio gobio balcanicus cytochrome b	44	0.001
gb/AF157939.1/AF157939 Spermophilus columbianus columbianus		
gb AF157937.1 AF157937 Spermophilus washingtoni isolate 589	44	0.001
go Arts/93/. I (Arts/93/) Specimophilius washington	44	0.001
gb AF157936.1 AF157936 Spermophilus washingtoni isolate S88		0.001
chlasis7915 llasis7915 Spermophilus richardsoni isolate 561	44	
	44	a.aal
gb AF157914.1 AF157914 Spermophilus richardsoni isolate 562		0.001
gb AF157912.1 AF157912 Spermophilus undulatus isolate S60 C	44	
	44	0.001
gb AF157906.1 AF157906 Spermophilus undulatus isolate S55 C	44	q.qqI
gb/AF157891.1/AF157891 Spermophilus elegans elegans isolate		
gb AF157882.1 AF157882 Spermophilus columbianus columbianus	44	0.001
	44	0.001
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chings frace 1 lagis 7858 Spermophilus citellus isolate Sil/ C	44	
	44	0.001
	44	0.001
gb AF030497.1 AF030497 Crocidura brunnea cytochrome b (cyt		
qb U03541.2 LAU03541 Lenoxus apicalis cytochrome b gene. pa	44	0.001
	44	0.001
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Inconggai ilafonggai Tomocichla tuba cytochrome b (cyto)	44	
	44	0.001
	44	0.001
gb AF094633.1 AF094633 Stachyris whiteheadi cytochrome b ge		0.001
-hlasogaezi ilasogaezi Eminia lepida Cytochrome D gene, par	44	
	44	0.001
	44	0.001
gb/AF166348.1/AF166348 Phascolarctos cinereus cytochrome b		0.001
17 27 29 29 7 1 1 27 28 29 7 Geomys burgarius Ozarkensis Cytochio	44	0.001
	44	0.001
	44	0.001
db AF158693.1 AF158693 Geomys bursarius bursarius cytochrom		0.001
This sace that sace Geomys burgarius missouriensis Cytoc	44	
	44	0.001
	44	0.001
gb AF091632.1 AF091632 Bubalus depressicornis cytochrome b	44	0.001
This propers 1) as 107815 Dromicions gliroides cytochicme b 9e		
gb AF102814.1 AF102814 Vombatus ursinus cytochrome b gene,	44	0.001
gb As 102814. I As 102814 Vollabation of Crypt Services of Crypt Services	44	0.001
db AF022065.1 Tragelaphus euryceros cytochrome b (cytb) ge gb AF022059.1 Kobus ellipsiprymnus cytochrome b (cytb) gen	4+	0.001
Liampanco il tohus allinging/mnus CVEOCRIOME D (CYCO/ 9511		0.001
CD1A-022037.11 WOOLS C11PG-PG/" 1		
This records 11 Aprilone cervicabra cytochrome b (cyto) gene	44	
gb AF022058.1 Antilope cervicapra cytochrome b (cyto) gene	44	0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene	44	0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene gb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g	44	0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene gb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g	44	0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene gb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g gb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb	44 44 44	0.001 0.001 0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene gb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g gb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb gb U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene,	44	0.001 0.001 0.001 0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene gb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g gb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb gb U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene, gb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cyt	44 44 44	0.001 0.001 0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene gb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g gb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb) gb U69863.1 PSU69861 Python sebae cytochrome b (cytb) gene, gb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cyt gb AF143193.1 AF143193 Epinephelus sp. cytochrome b (cytb)	44 44 44 44	0.001 0.001 0.001 0.001 0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene gb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) gene, gb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb) gb U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene, gb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cyt) gb AF143193.1 AF143193 Epinephelus sp. cytochrome b (cytb)	44 44 44 44 44	0.001 0.001 0.001 0.001 0.001
db AF022058.1 Antilope cervicapra cytochrome b (cytb) gene db AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m db AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g db AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb) db U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene, db U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cyt) db AF143193.1 AF143193 Epinephelus sp. cytochrome b (cytb) db AF121222.1 AF121222 Amphiprion ocellaris isolate 8 cytochrome b (cytochrome b)	44 44 44 44 44	0.001 0.001 0.001 0.001 0.001 0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene gb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g gb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb) gb U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene, gb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cyt gb AF143193.1 AF143193 Epinephelus sp. cytochrome b (cytb) gb AF121222.1 AF121222 Amphiprion ocellaris isolate 8 cytoc gb AF096625.1 AF096625 Kobus ellipsiprymnus defassa cytochrome	44 44 44 44 44	0.001 0.001 0.001 0.001 0.001 0.001
gb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene gb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, m gb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) g gb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb) gb U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene, gb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cyt gb AF143193.1 AF143193 Epinephelus sp. cytochrome b (cytb) gb AF121222.1 AF121222 Amphiprion ocellaris isolate 8 cytoc gb AF096625.1 AF096625 Kobus ellipsiprymnus defassa cytochrome gb AF096624.1 AF096624 Kobus ellipsiprymnus ellipsiprymus c	44 44 44 44 44 44	0.001 0.001 0.001 0.001 0.001 0.001 0.001
qb AF022058.1 Antilope cervicapra cytochrome b (cytb) geneqb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, mqb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) gene, mqb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb)qb U69843.1 PSU69863 Python sebae cytochrome b (cytb) gene,qb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cytb)qb AF143193.1 AF143193 Epinephelus ap. cytochrome b (cytb)qb AF121222.1 AF121222 Amphiprion ocellaris isolate 8 cytocqb AF096625.1 AF096625 Kobus ellipsiprymnus defassa cytochrome b (cytochrome b)qb AF096624.1 AF096624 Kobus ellipsiprymnus ellipsiprymus cqb AF081052.1 AF081052 Eulemur rubriventer cytochrome b (cytochrome b)	44 44 44 44 44 44 44 44 44 44 44 44 44	0.001 0.001 0.001 0.001 0.001 0.001 0.001
qb AF022058.1 Antilope cervicapra cytochrome b (cytb) geneqb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, mqb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) gqb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb)qb U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene,qb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cytb)qb AF143193.1 AF143193 Epinephelus sp. cytochrome b (cytb)qb AF121222.1 AF121222 Amphiprion ocellaris isolate 8 cytochromeqb AF096625.1 AF096625 Kobus ellipsiprymnus defassa cytochromeqb AF081052.1 AF081052 Eulemur rubriventer cytochrome b (cytochrome b (cytochrome b)	44 44 44 44 44 44 44 44	0.001 0.001 0.001 0.001 0.001 0.001 0.001
qb AF022058.1 Antilope cervicapra cytochrome b (cytb) geneqb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, mqb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) gqb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb)qb U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene,qb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cytb)qb AF143193.1 AF143193 Epinephelus sp. cytochrome b (cytb)qb AF121222.1 AF121222 Amphiprion ocellaris isolate 8 cytochromeqb AF096625.1 AF096625 Kobus ellipsiprymnus defassa cytochromeqb AF081052.1 AF081052 Kobus ellipsiprymnus ellipsiprymus cqb AF081049.1 AF081049 Eulemur macaco macaco cytochrome b (cytochrome b (cytochrome b (cytochrome cytochrome b (cytochrome cytochrome cytochr	44 44 44 44 44 44 44 44 44 44 44 44 44	0.001 0.001 0.001 0.001 0.001 0.001 0.001 0.001
qb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene.qb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, mqb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) gene,qb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb).qb U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene,qb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cytb)qb AF121222.1 AF121222 Epinephelus sp. cytochrome b (cytb)qb AF096625.1 AF096625 Kobus ellipsiprymnus defassa cytochrome bqb AF096624.1 AF096624 Kobus ellipsiprymnus ellipsiprymus cqb AF081049.1 AF081049 Eulemur rubriventer cytochrome b (cyto)qb AF081049.1 AF081048 Eulemur fulvus albifrons cytochrome	44 44 44 44 44 44 44 44	0.001 0.001 0.001 0.001 0.001 0.001 0.001 0.001
qb AF022058.1 Antilope cervicapra cytochrome b (cytb) gene.qb AF022057.1 Tragelaphus oryx cytochrome b (cytb) gene, mqb AF022054.1 Antidorcas marsupialis cytochrome b (cytb) gqb AF016637.1 AF016637 Connochaetes gnou cytochrome b (cytb)qb U69863.1 PSU69863 Python sebae cytochrome b (cytb) gene,qb U69844.1 LTU69844 Lichanura trivirgata cytochrome b (cytb)qb AF143193.1 AF143193 Epinephelus sp. cytochrome b (cytb)qb AF121222.1 AF121222 Amphiprion ocellaris isolate 8 cytochrome b qb AF096624.1 AF096624 Kobus ellipsiprymnus defassa cytochrome b qb AF081042.1 AF081042 Eulemur rubriventer cytochrome b qb AF081048.1 AF081048 Eulemur fulvus albifrons cytochrome b qb AF082063.1 AF082061 Eulemur fulvus albifrons cytochrome b	44 44 44 44 44 44 44 44 44 44 44 44 44	0.001 0.001 0.001 0.001 0.001 0.001 0.001 0.001
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dbj AB023903.1[AB02190] Pecaurista leucogenys micocnondriat	44	0.001
dbj D88983 1 D88983 Bubalus bubalis mitochondrial DNA for c	44	0.001
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db1[0]2121.1[88UMTC82] Subalus senes buballs mitochondrial	44	0.001
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dbilas094974 11AB094974 Capra hiecus micochondelai UNA tot	_4.1	0.001
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db1 D88634.1 D88634 Bubalus bubalis micochondrial DNA for C	44	0.001
dbi D88631.1 D88631 Bubalus bubalis mitochondriai DNA LUL C	44	0.001
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emb X92531.1 DLCYTB D.leucas cytochrome b gene (complete se	44	0.001
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gb AF272636_1 AF272636 Clethrionomys gapperi specimen-vouch	40	0.021
gb AF272634.1 AF272634 Clethrionomys gapper: speciment	40	0.021
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achromathus cytochrome o (40	0.021
objAF155400.11AF155400 Peromyacua pectoralia laceianua cyto	40	0.021
ablaf155385 1/AF155385 Peromyacua accondent factore	40	0.021
gb/AF155184.1/AF155184 Peromyscus activates isolate AcizAC	47	0.021
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dblAF155599. LIAF155587 Sigmodon ochrognathus isolate Elemes.	47	0.021
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gbiaf121511 11AF121511 Trachyphonus diriaddir cycochrome b gene. co	42	0.021
and a louis cytocheome b gent, p	•3	g.g2l g.g2l
William Harris Barrier and Harris FR. 14	47	0.011
CELING 000954 LL Cavia porcellus complete mitochondelal genome		- · · - ·

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dD/AFU86732.1/AFU86732 SMINCHODSIS padminopiate 1/4-1-1-1-1-1	40	0.021
GDI (1959) 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	40	0.021
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QD U62 / U / T CASKCITES CIGITATION ASTRONO CLOSE - CALL		0.084
emb[AJ004315.1]HCAJ4315 Hippolais caligata mitochondrial cy	_38	0.304

Alignments

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D82890	869	• • • • • • • • • • • • • • • • • • • •	844
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<u>AF123615</u>	101	• • • • • • • • • • • • • • • • • • • •	844
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<u>U62685</u>	179 866		844
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AB036402	173	•••••	151 151
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D82894	869		844
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D34638	869	a	844
D34636	869	gg.	844
AB037602	869	aa	
AB018985	869		844
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AB004070	869	gg	844
X92531	869	gg.	844
U07565	869	gg	844
U10367	773		748
U10364	773		748
Z96068	869	g	844
	773		749
<u>076507</u>	791		772
AF157466	793		774
AF157464	793 793		774
AF157463	93		74
AY016019	93 869		850
AF027330	869		850
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AF027309	869		850 850
AF027308	869		850
AF027307	869		352
AF266188	371		389
AF324034 AF272639	408 869		850
AF272636	869		850
AF272614	869		850
AF272631	862		850
AF182711	712		673
AF182687	774		755
AF155422	669		850
AF155400	669		650 650
AF135285	669		850
82135384	669		030
AP135322	669 669		650
9617777	467		139
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WO 02/0772	78	114
		450
<u> AF155589</u>	669	850
<u> AF155588</u>	B69	850
AF123531	771	752
AF264047	869	850
AF206531	771	752
AF192706	863	. 844
NC 000884	15032	15013
AF004572	869	850
AF088932	869	850
U62697	176	157
U62681	179	160
U62707	179gn.	. 154
AJ004315	773	. 748
	•	
Database	e: nt	•
Poste	i date: Mar 2, 2001 12:20 AM	
Mimber o	of letters in database: 2,863,82	7,885
Number (of sequences in database: 807,5	97
Lambda	K H	
1.37	0.711 1.31	
Cannad		
Gapped Lambda	K H	
1.37	0.711 1.31	

	1	
Matrix: Di	lastn matrix:1 -3 ties: Existence: S, Extension: 2	
Gap Penal	cles: Existence: J, Extension, -	

Matrix: blastn matrix:1 -3
Gap Penalties: Existence: 5, Extension: 2
Number of Hits to DB: 19068
Number of Sequences: 807597
Number of extensions: 19068
Number of successful extensions: 7580
Number of sequences better than 10.0: 2441
length of query: 26
length of database: 2,863,827,885
effective HSP length: 17
effective length of query: 9
effective length of database: 2,850,098,736
effective search space: 25650888624
effective search space used: 25650888624
T: 0
A: 30
X1: 6 (11.9 bits)
X2: 15 (29.7 bits)
S1: 12 (24.3 bits)

Table 12. The other animals belonging to distantly related species analyzed by our primers to demonstrate its universal nature

SN.	Name of the animal
1.	Indian black buck no. l
2.	Indian black buck no 2
3	sh ee p
4	pig
5	dog
6	chimpanzee (chimss)
7	human (humsk)
8	Hamster
9	crocodile no l
10	crocodile no2
11	turtle no l
12	turtle no2
13	mouse
14	varanus
15	Naga-naga snake
16	Indian elephant
17	hen
18	dugong
19	lizard
20	weaver bird not
21	weaver bird no.2
22	buffalo no l
23	buffalo no 2

CLAIMS

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Universal primers named as 'mcb 398' and 'mcb 869' capable of amplifying a fragment of cytochrome b gene of any animal species in polymerase chain reaction (PCR) and revealing the identity of the biological material of any animal of unknown origin at species and sub-species level, said primers, having the sequences:

primers name Sequence (5'-3')
mcb 398 "TACCATGAGGACAAATATCATTCTG"
mcb 869 "CCTCCTAGTTTGTTAGGGATTGATCG

- Primers as claimed in claim 1 wherein the fragment of mitochondrial cytochrome b
 gene is capable of significantly discriminating amongst various evolutionary lineages
 of different animal species.
 - 3. Primers as claimed in claim 1 wherein the fragment of mitochondrial cytochrome b gene is flanked by the highly conserved sequences amongst a vast range of animal species.
- 4. Primers as claimed in claim 1 wherein the fragment on mitochondrial cytochrome b gene which is polymorphic inter-specifically, but monomorphic at intra species sources.
 - 5. Primers as claimed in claim 1 wherein in Antilope cervicapra species, the sequences of the fragment mentioned under claim 1 are as follows:
- 20 Mitochondrial cytochrome b gene sequence (398-869 bp) of Antilope cervicapra:
 "taccatgaggacaaatatetttttgaggagcaacagteatcaccaateteettteagcaateccatacateggtacaaacetag
 tagaatgaatetgaggagggttetcagtagataaagcaaceettaceegatttttegeettecaetttateeteeatttateattge
 agceettaccatagtacacetaetgttteteeacgaaacaggatecaacaaceecacaggaateteatcagacgcagacaaa
 attecattecaceectactacaactateaaagatateetaggagetetactattaattttaaeceteatgettetagteetatteteace
 ggacetgettggagacecagacaactatacaccagcaaacecaettaatacacceccacatatcaagecegaatgataette
 ctatttgcatacgcaateeteegateaatteetaacaaactaggagg"
 - 6. A method for the identification of the animal from a biological sample, said method comprising the steps of:
 - a) isolating and amplifying the DNA from the biological sample to be tested using the primers as claimed in claim 1,
 - b) sequencing the amplified products,
 - c) blasting the sequence resolved in step (b) against mito database of National Centre for Biotechnology Information (NCBI) using BLAST program and determining the most likely family of the animal source of the biological sample,

- d) blasting the sequence resolved in step (b) against non-redundant (nr) database of National Centre for Biotechnology Information (NCBI) using BLAST program and determining the most likely genus, species or more precisely the sub-species of the animal source of the biological sample,
- e) identifying the most significant alignment of the sequence resolved with cytochrome b gene sequence of the animal identified in steps (c) and (d) respectively and selection of these animals as 'reference animals' for further studies,

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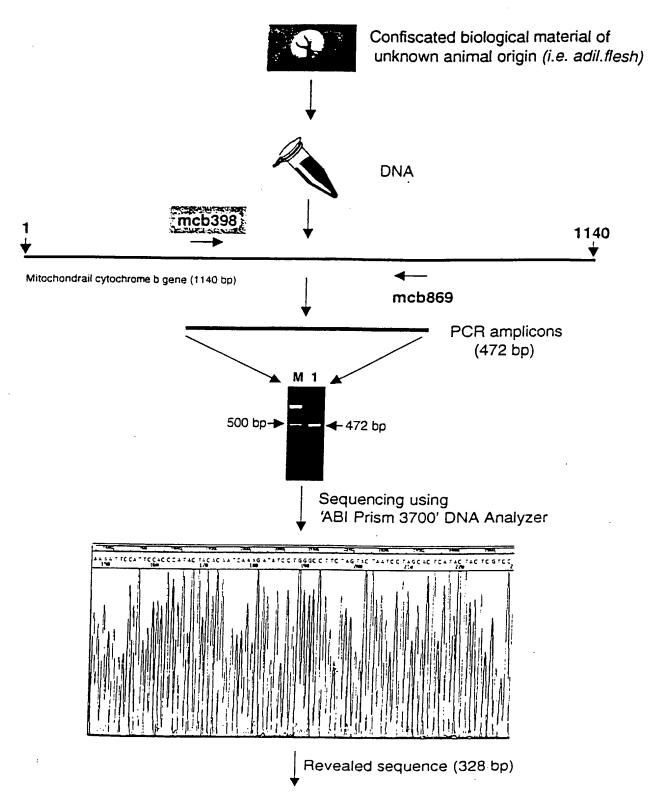
- f) isolating and amplifying and sequencing the DNA sequences from the reference animal on both strands in triplicate using the primers as claimed in claim 1,
- g) aligning the sequences obtained using CLUSTRAL program and identifying the variable sites amongst the animals analyzed,
- h) comparing the nucleotide sequences pair-wise to determine the variation among the animals resolved and identifying the nucleotide sequence to which the DNA sequence of the biological sample bears maximum similarity as the source animal of the biological sample.
- 7. A method as claimed in claim 6 wherein the universal PCR protocol works universally with the DNA template of any unknown animal origin and the universal primers mentioned under column 4.
- 8. A method as claimed in claim 6 wherein the Amplification reactions should be carried out in 20 μl reaction volume containing approximately 20 ηg of template DNA, 100μm each of dNTPs, 1.25 pmole of each primer, 1.5mM MgCl₂, 0.5 unit of Ampli*Taq* Gold (Perkin-Elmer-Cetus, USA) DNA polymerase and 1X PCR buffer (10mM Tris-HCl, pH 8.3, and 50mM KCl). The amplification profiles followed should be: an initial denaturation at 95°C for 10 min, followed by 35 cycles each of denaturation at 95°C for 45 s, annealing at 51°C for 1 min, and extension at 72°C for 2 min. The extension step at 35th cycles should be held for 10 min.
- 9. A method as claimed in claim 6 wherein the method enables identification of species of analyzed material (i.e. the DNA isolated from confiscated animal remain of unknown origin) using the public databases such as GenBank, NCBI etc.
- 10. A method as claimed in claim 6 wherein the method is used for animal identification to establish the crime with the criminal beyond a reasonably doubt.
- 11. A method as claimed in claim 6 wherein the method is used to establish the identity of

- biological materials such as skin, horns etc confiscated from animal poachers, if it is that of an endangered species.
- 12. A method as claimed in claim 6 wherein the method is used for establishment of the identity of confiscated animal parts and products of endangered animal species for the purpose of production of molecular evidence of animal hunting and related crime in the court of law, so that the human violation of the wildlife resources could be controlled.

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- 13. A method as claimed in claim 6 wherein the method is used to have an idea of the geographical location of the commitment of wildlife crime based on the cytochrome b gene haplotype of poached animal identified by the universal primer invented.
- 10 14. A method as claimed in claim 6 wherein the method is used for animal identification to detect the adulteration of animal meat in food products for the purpose of food fortification, by the food fortification agencies.
 - 15. A method as claimed in claim 6 wherein the method is used to provide a universal technique for detection of the origin of blood or blood stains etc collected from the scene of crime related to offences such as murder, rape etc, in order to establish the origin of blood found at scene of crime when it sounds as if criminals have wontedly spread the blood of an animal at the scene of crime, to confuse the crime investigation agencies and forensic scientists with human blood.
- 16. A method as claimed in claim 6 wherein the method is used so that it can be converted to a (a) COMMERCIAL 'MOLECUALR KIT' and (b) 'DNA CHIPS' based applications for wildlife identification in forensics.



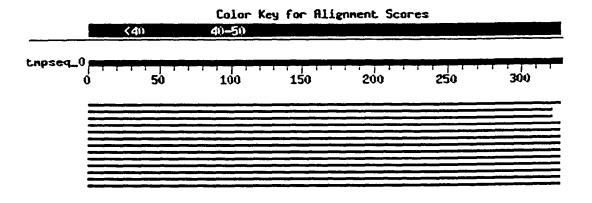
TGAATCTGAGGAGGCTTCTCAGTAGACAAAGCTACCCTGACAC
GATTCTTTGCCTTCCACTTCATCTTCATCTTCAGC
TCTAGCAGCAGTCCACCTCCTATTCCTTCACGAGACAGGATCT
AACAACCCCTCAGGAATAGTATCCGACTCAGACAAAATTCCAT
TCCACCCATACTACACAATCAAAGATATCCTGGGCCTTCTAGT
ACTAATCCTAGCACTCATACTACTCGTCCTATTCTCACCAGAC
CTGTTAGGAGACCCCGATAACTACATCCCTGCCAACCCTCTAA
ATACCCCTCCCCATATCAAGCCTGAAT

Figure 1 a

Sequence of cytochrome b gene (328 bp) revealed from biological material of unknown origin i.e. 'adil.flesh' using primers 'mcb398' and 'mcb869'

Homology search in 'nr' database using 'BLAST'

http://www.ncbi.nlm.nih.gov/BLAST/



Sequences producing significant alignments:	Score (bits)	E Value
gb AY005809.1 <i>Panthera pardus</i> cytochrome b gene, partial c gb AF053054.1 AF053054 <i>Panthera tigris sumatrae</i> isolate Su1 gb AF053053.1 AF053053 <i>Panthera tigris tigris</i> isolate B7 mi gb AF053050.1 AF053050 <i>Panthera tigris corbetti</i> isolate C2 gb AF053049.1 AF053049 <i>Panthera tigris corbetti</i> isolate C1	603 527 527 476 476	e-170 e-147 e-147 e-132 e-132

Selection of reference animals based on above information and further analysis using primers 'mcb398' and 'mcb869'

> Multiple sequence alignments using 'Autoassembler'

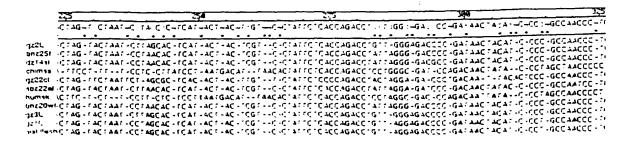


Figure 1 b

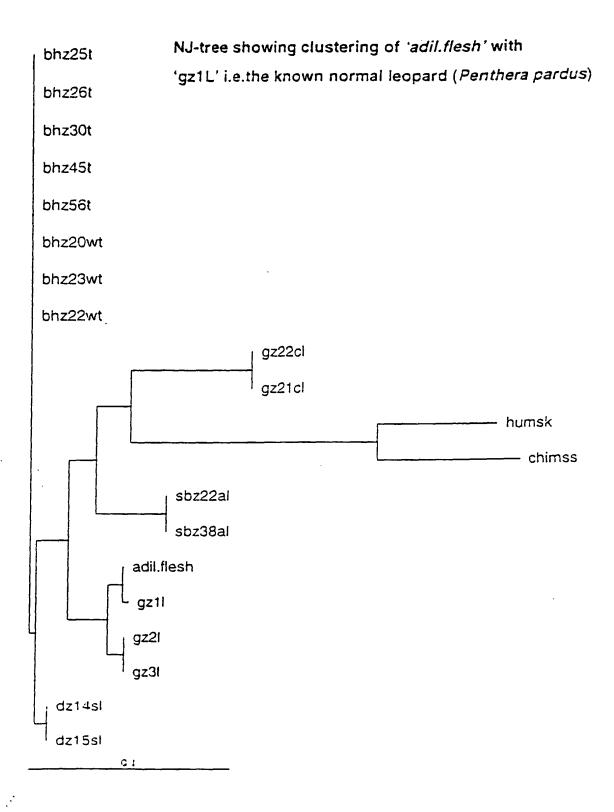


Figure 1c

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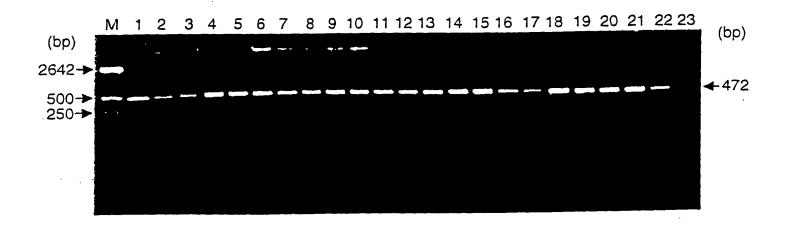


Figure 2

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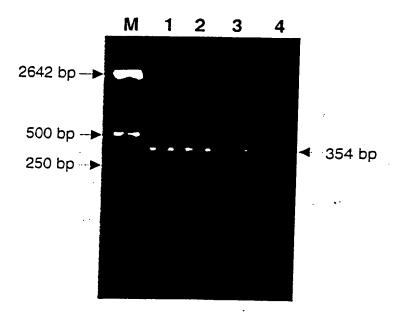


Figure 3

6/6

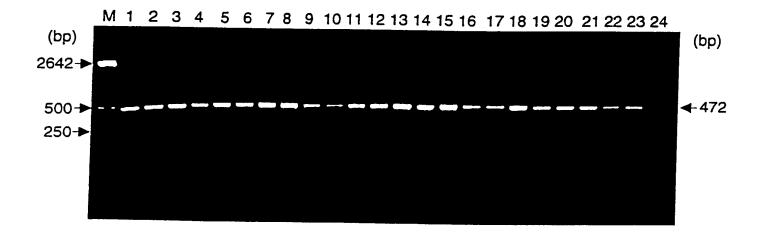


Figure 4

INTERNATIONAL SEARCH REPORT

Inte Application No PC I / IN 01/00055

	FICATION OF SUBJECT MATTER C12Q1/68		
,			
	International Patent Classification (IPC) or to both national classifica	ution and IPC	
Minimum do	SEARCHED cumentation searched (classification system followed by classification	on symbols)	
IPC 7	C12Q		
Documentat	ion searched other than minimum documentation to the extent that su	uch documents are included in the fields se	earched
D o o a moj man			
Electronic d	ata base consulted during the international search (name of data bas	se and, where practical, search terms used)
BIOSIS	, EPO-Internal, EMBL, WPI Data, PAJ,	MEDLINE, EMBASE	
C. DOCUMI	ENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the rele	evant passages	Relevant to claim No.
		, 5	
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	CONSERVED PRIMERS" PROCEEDINGS OF THE NATIONAL ACADE	MV OF	
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X Furti	her documents are listed in the continuation of box C.	χ Patent family members are listed	in annex.
		"T" later document published after the inte or priority date and not in conflict with	
consid	ent defining the general state of the art which is not lered to be of particular relevance	cited to understand the principle or the invention	
filing o	late	*X* document of particular relevance; the c cannot be considered novel or cannot involve an inventive step when the do	be considered to
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	ent referring to an oral disclosure, use, exhibition or means	document is combined with one or mo ments, such combination being obvio	ore other such docu-
	ent published prior to the international filing date but nan the priority date claimed	in the art. *&* document member of the same patent	family
Date of the	actual completion of the international search	Date of mailing of the international sea	arch report
2	8 February 2002	12/03/2002	
Name and	nailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2	Authorized officer	
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Integral Application No

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